

TAC Meeting

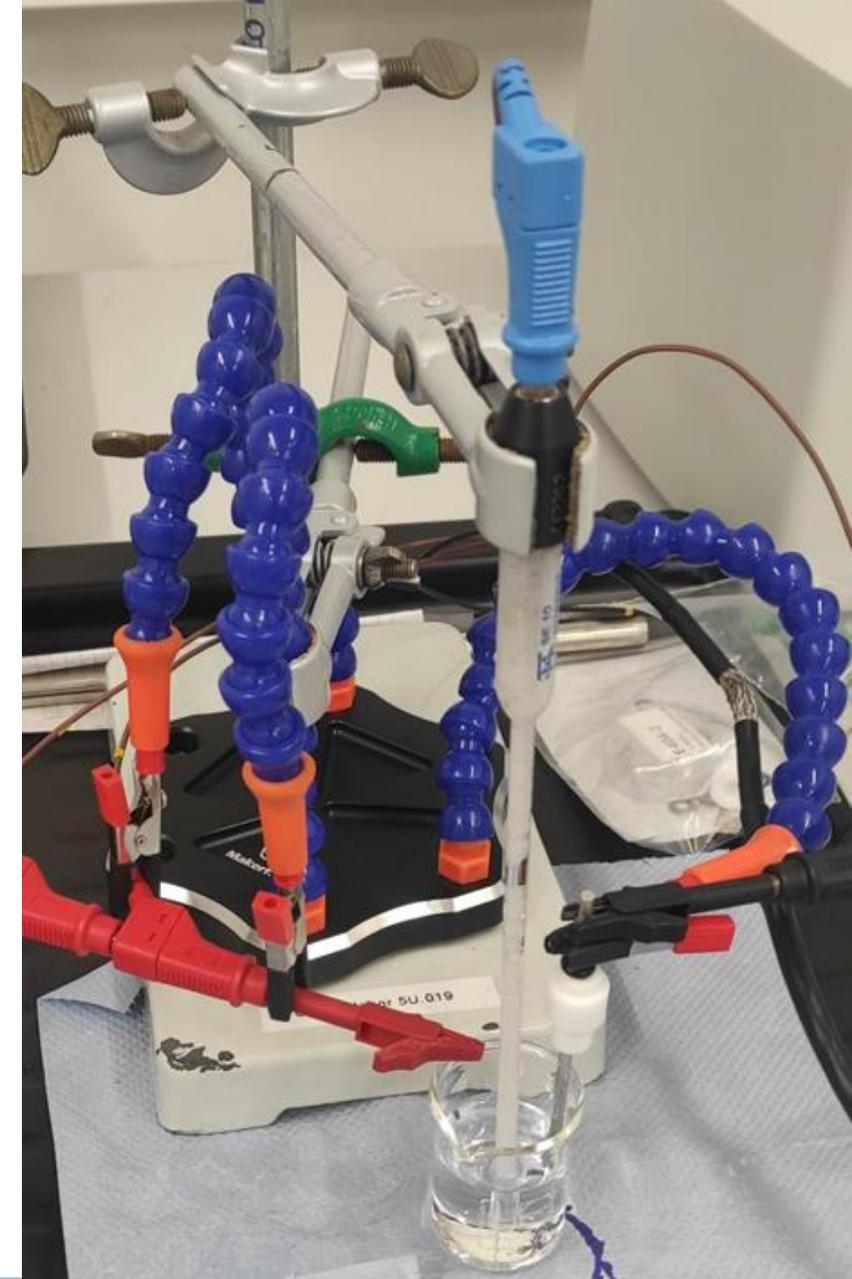
Development of electrochemical aptamer-based sensor for
the early detection of cardiac troponin I

10-08-23

Ivan Lopez Carrasco

Content

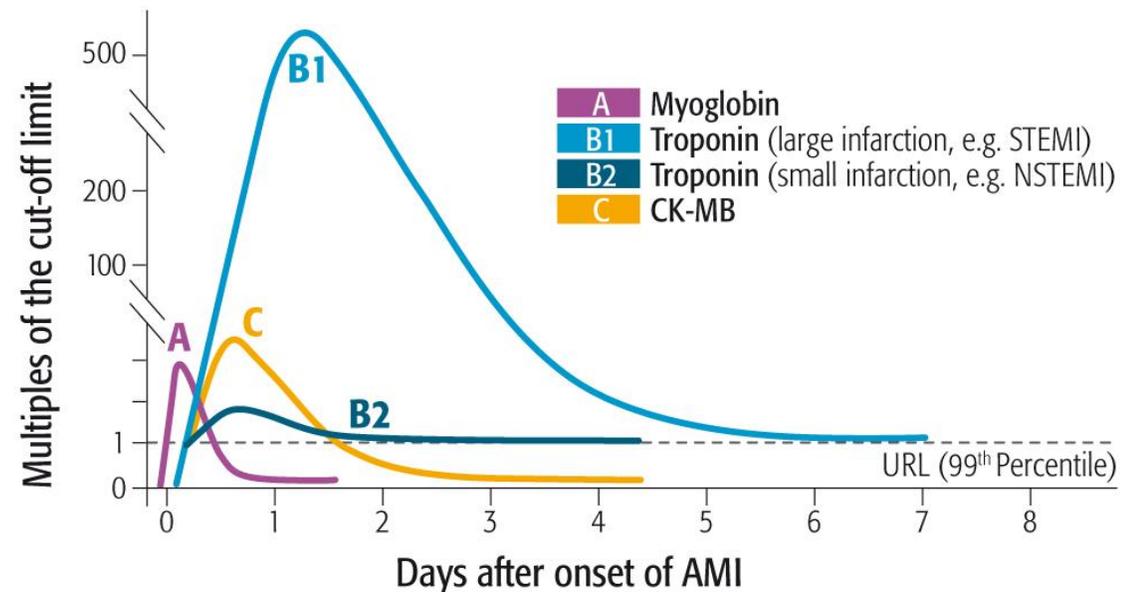
1. Research topic
2. Objective of research
3. Current status of research
4. Next steps
5. Publications



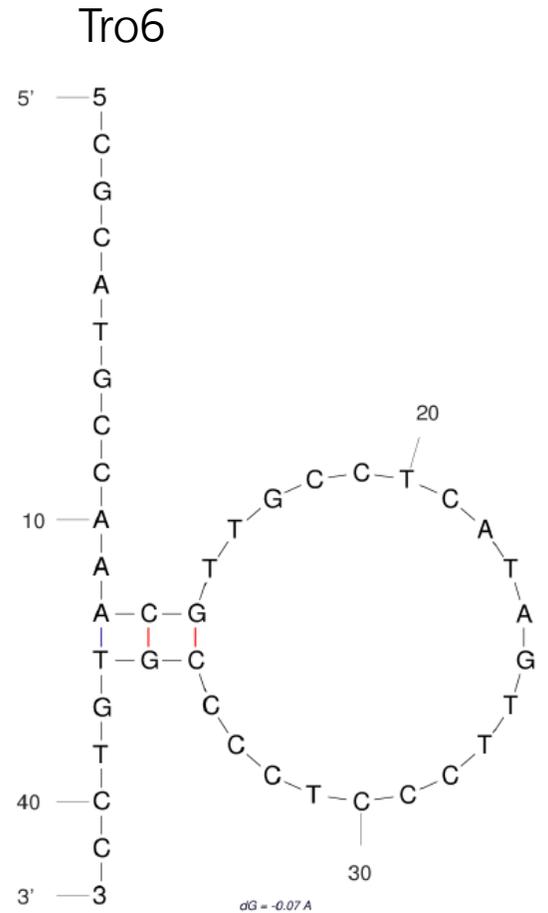
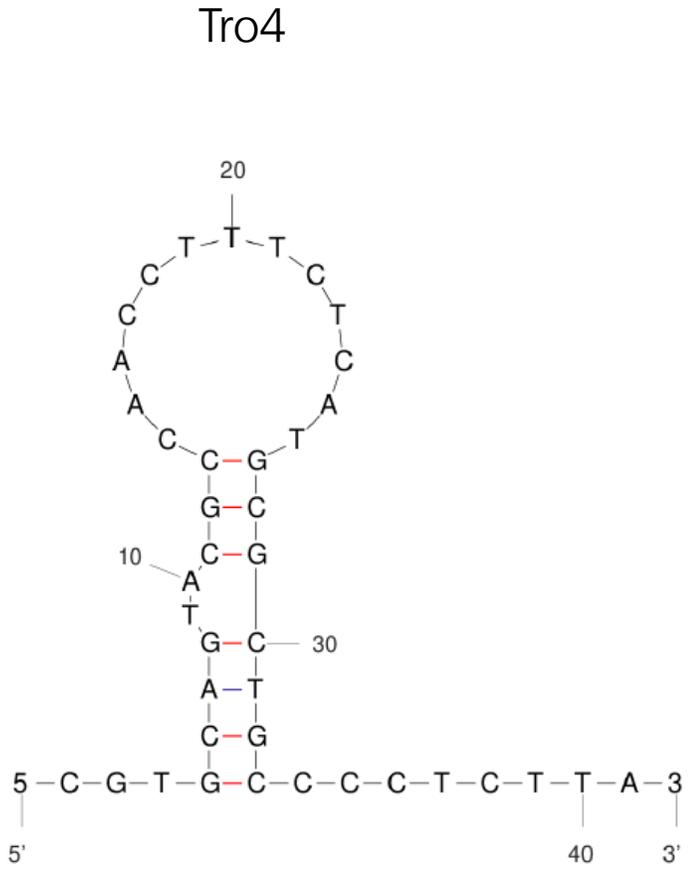
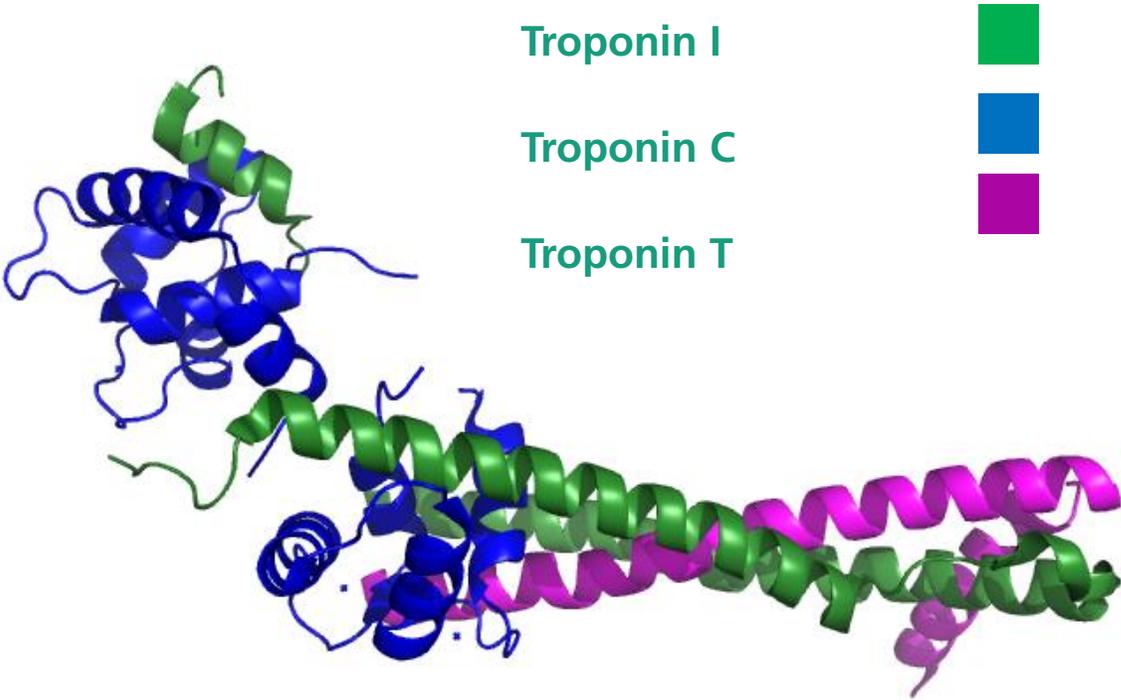
Ability of Troponin I to Predict Cardiac Events in Patients Admitted from the Emergency Department

Michael C. Kontos, MD, FACC, F. Philip Anderson, PhD, Ramin Alimard, MD,
Joseph P. Ornato, MD, FACC, James L. Tatum, MD, Robert L. Jesse, MD, PhD, FACC
Richmond, Virginia

- Reference value Troponin I: 0 - 0.04 ng/mL.
- Serum levels rise within the next 12 h after AMI.
- Optimal cutoff value for MI 1 ng/ml.
- Cutoff values as markers of CT-induced cardiotoxicity cTnI > 0.50ng/ml cTnI > 0.08ng/ml

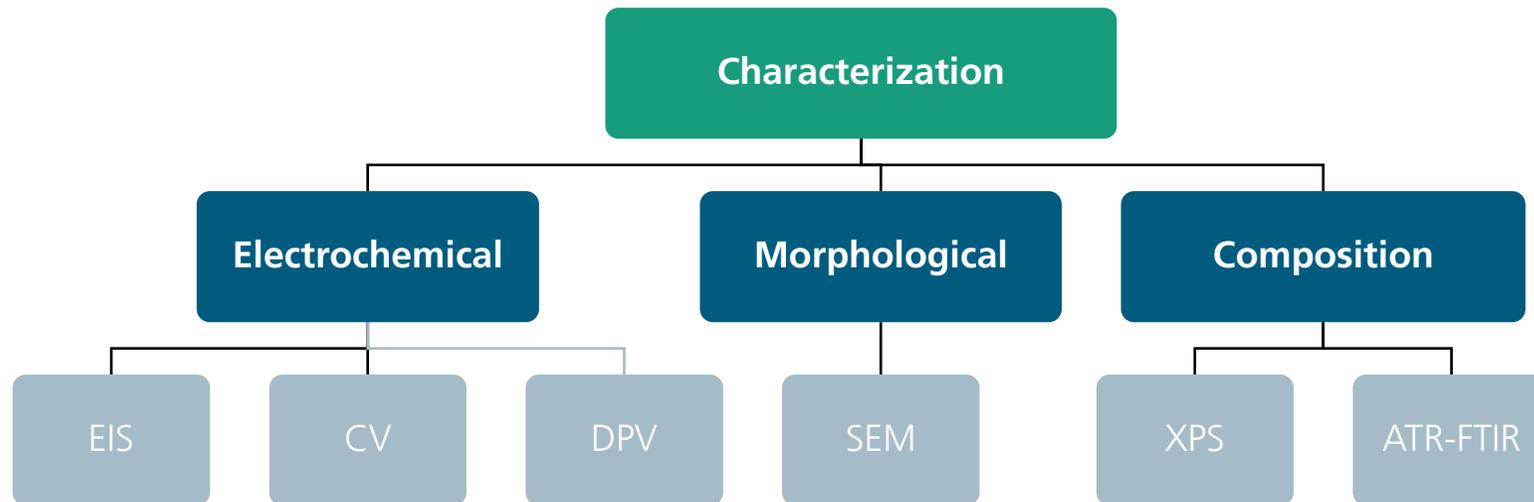


Troponin complex as biomarker of heart injury and aptamers that bind to troponin I



<https://en.wikipedia.org/w/index.php?title=Troponin&oldid=1044978411>

Development of Aptamer-based sensor for cardiac troponin I



Procedure follow for electrochemical characterization

The screenshot displays the Nova 2.1.5 software interface for electrochemical characterization. The main window is titled "DATA" and shows a procedure sequence for "PEN-Tro4+MCH-+100nMcTnI20-06-23-S2 (20/06/2023 18:46) AUT52115". An orange arrow points to the "CV staircase" step in the sequence.

Procedure Sequence:

- Autolab control
- OCV 0.189 V
- Apply 0 V
- Cell on
- Wait 5 s
- FRA measurement
- Wait 5 s
- CV staircase (highlighted with an orange arrow)
- Wait 10 s
- Square wave
- Wait 10 s
- Square wave
- Wait 10 s
- Differential pulse
- Wait 10 s
- Differential pulse
- Wait 10 s
- Cell off
- Wait 10 s

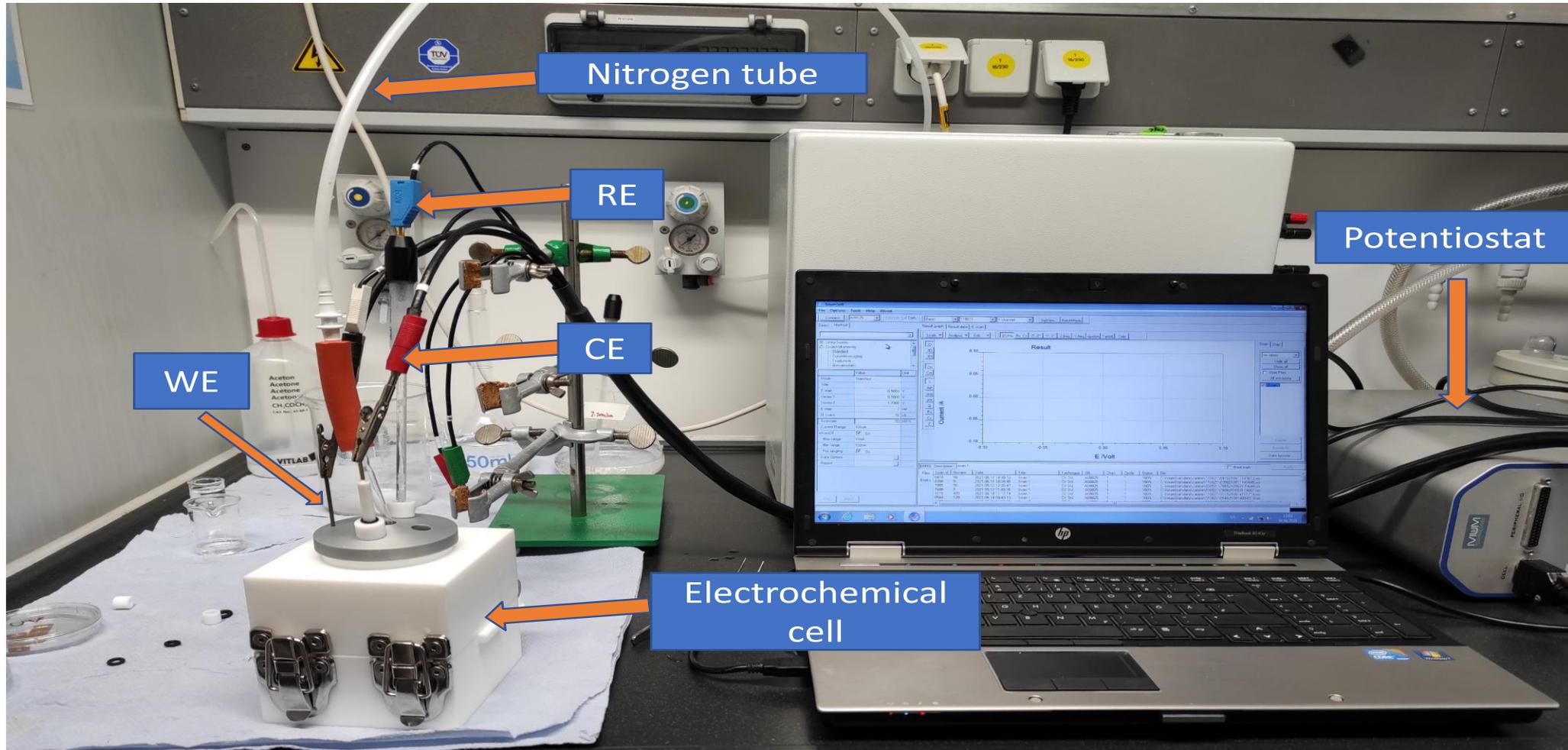
Plots:

The "Plots" section at the bottom shows several electrochemical data plots:

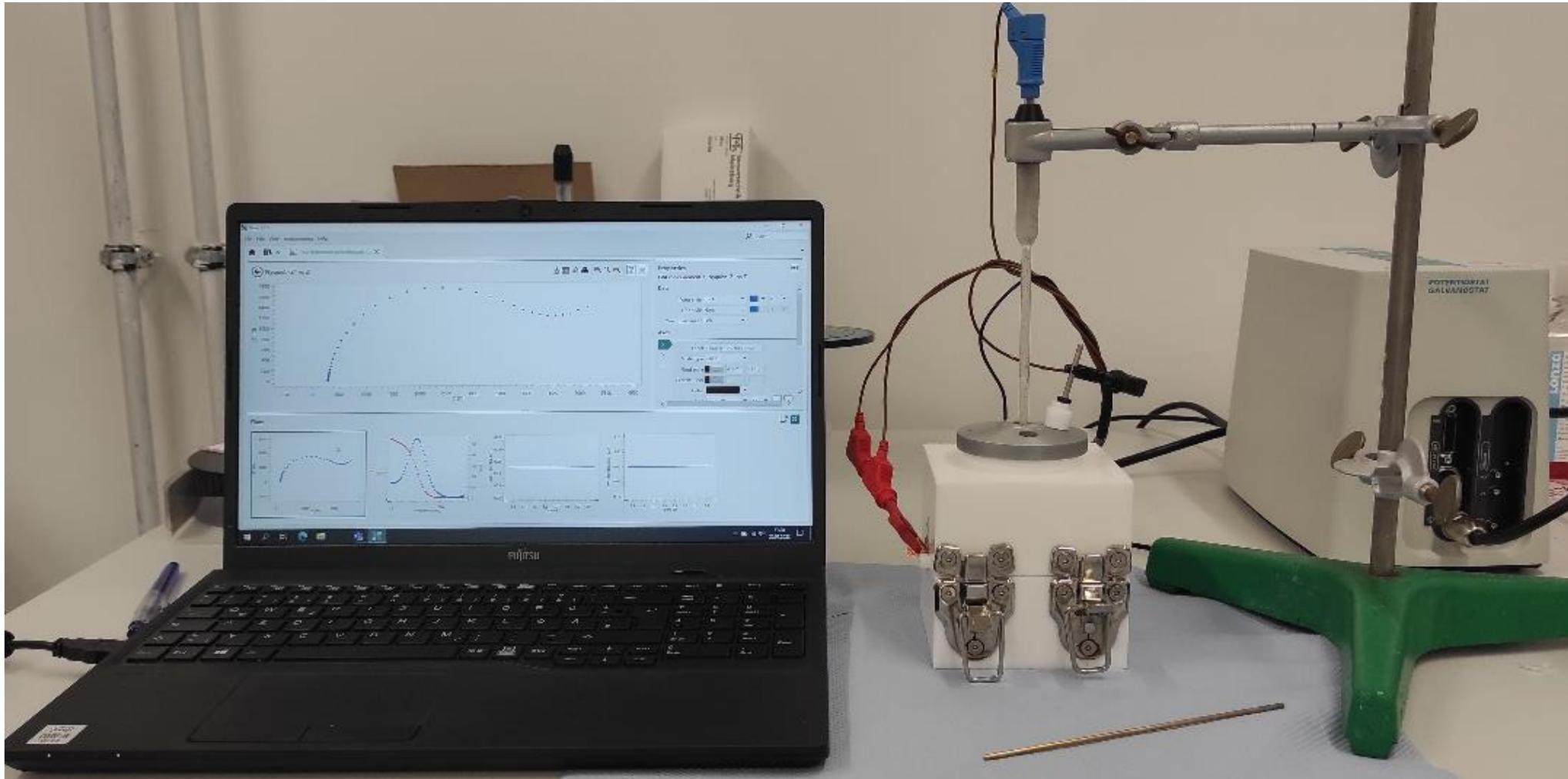
- Impedance plot (Z' vs Z'')
- Frequency response plot (Current vs Frequency)
- Cyclic voltammogram (Current vs Potential)
- CV staircase plot (Current vs Potential)
- Electrochemical Impedance Spectroscopy (EIS) plot (Current vs Potential)
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The Windows taskbar at the bottom shows the system time as 22:45 on 01/08/2023.

Experimental setup for electrochemical characterization



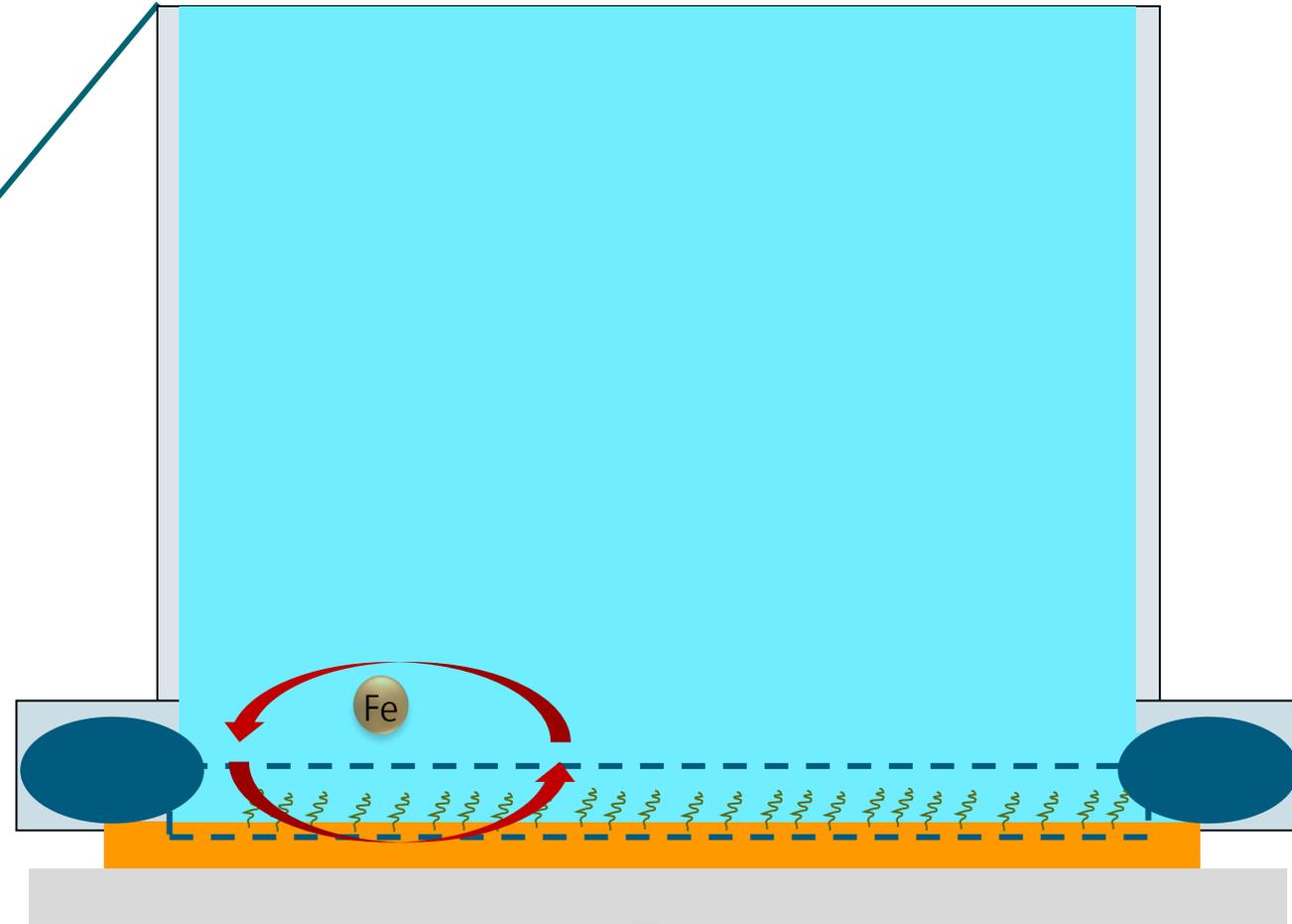
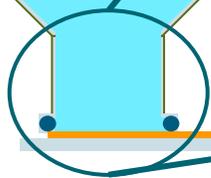
Experimental setup for electrochemical characterization



▪ Reference electrode

▪ Counter electrode

▪ 20mL volume

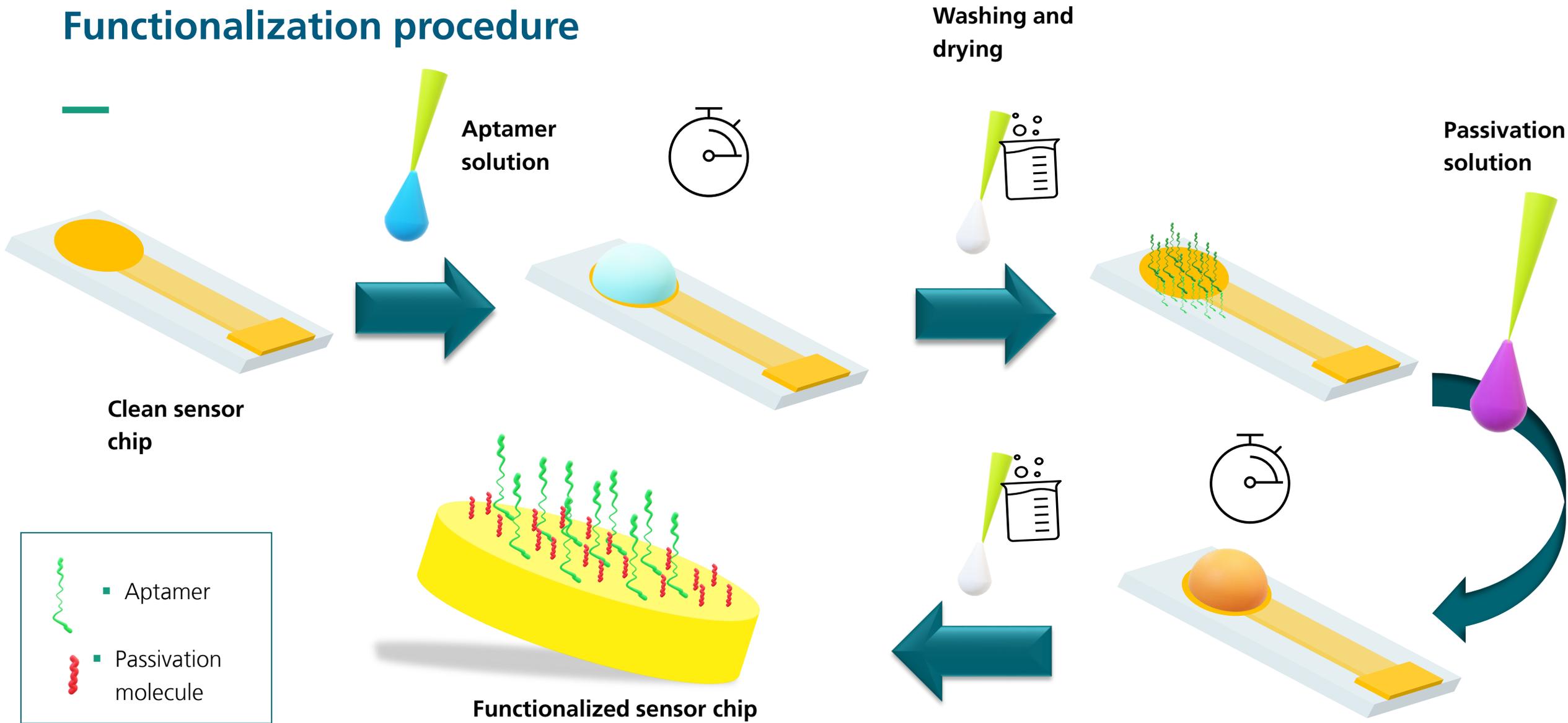


Transversal view

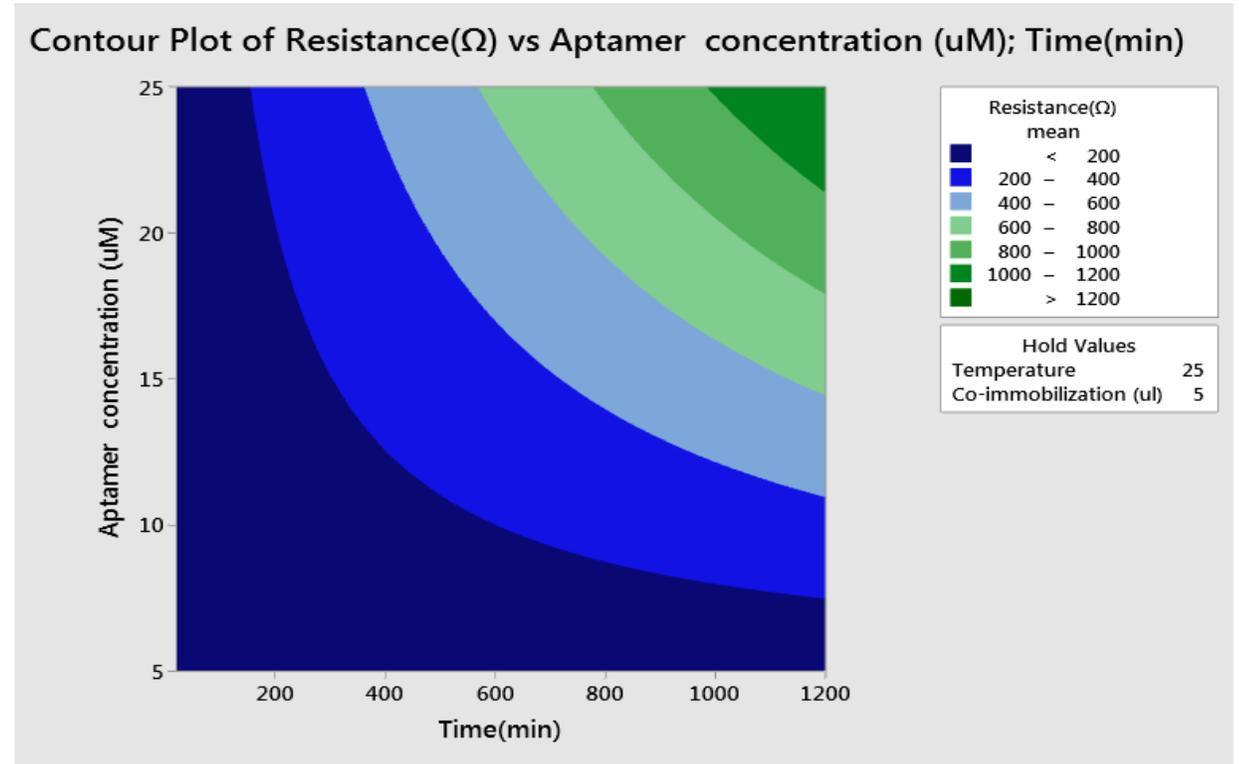
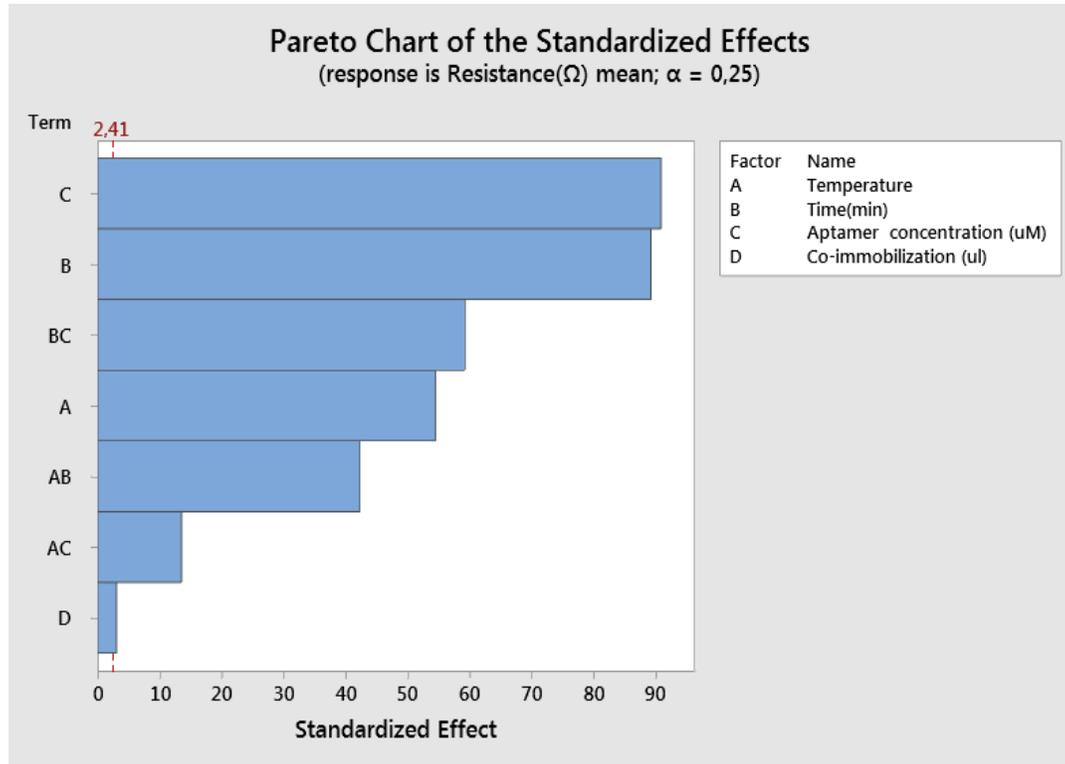
Parameter that are control in immobilization step

Code	Temperature	Time	Aptamer concentration (μM)	Co-immobilization	pH
Medium value 0	RT	10 hrs	15	50%	7.5
Low value -	4°C	20min	5	0	7
High Value +	46 °C	20 hrs	25	100%	8

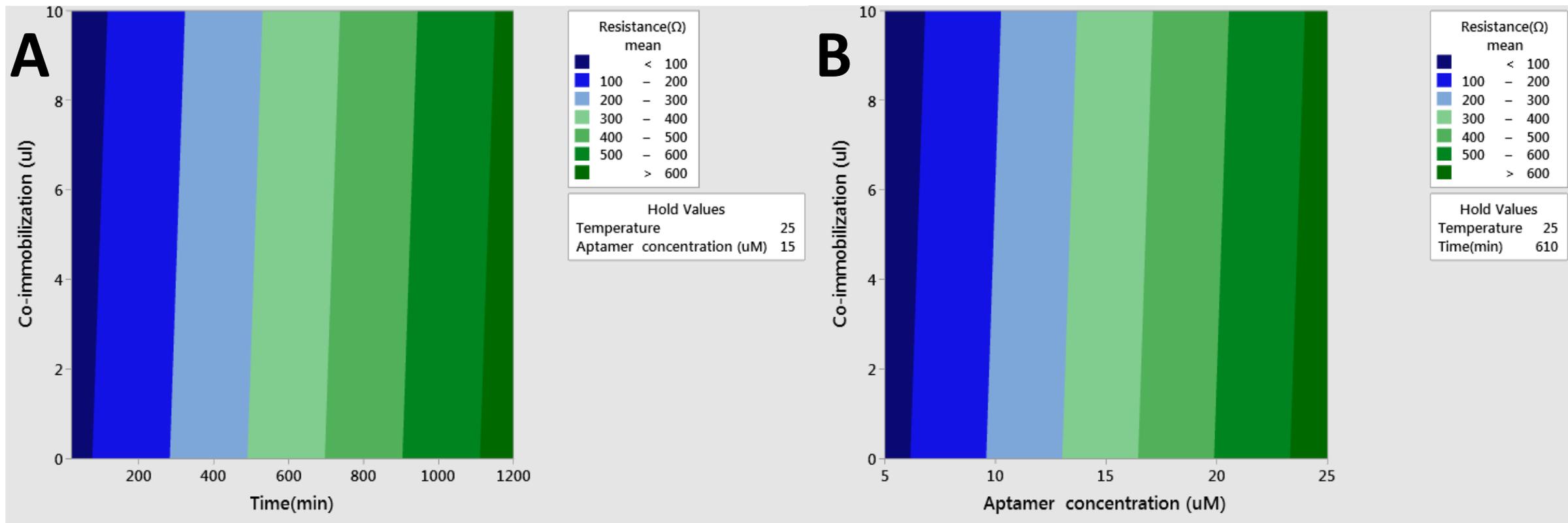
Functionalization procedure



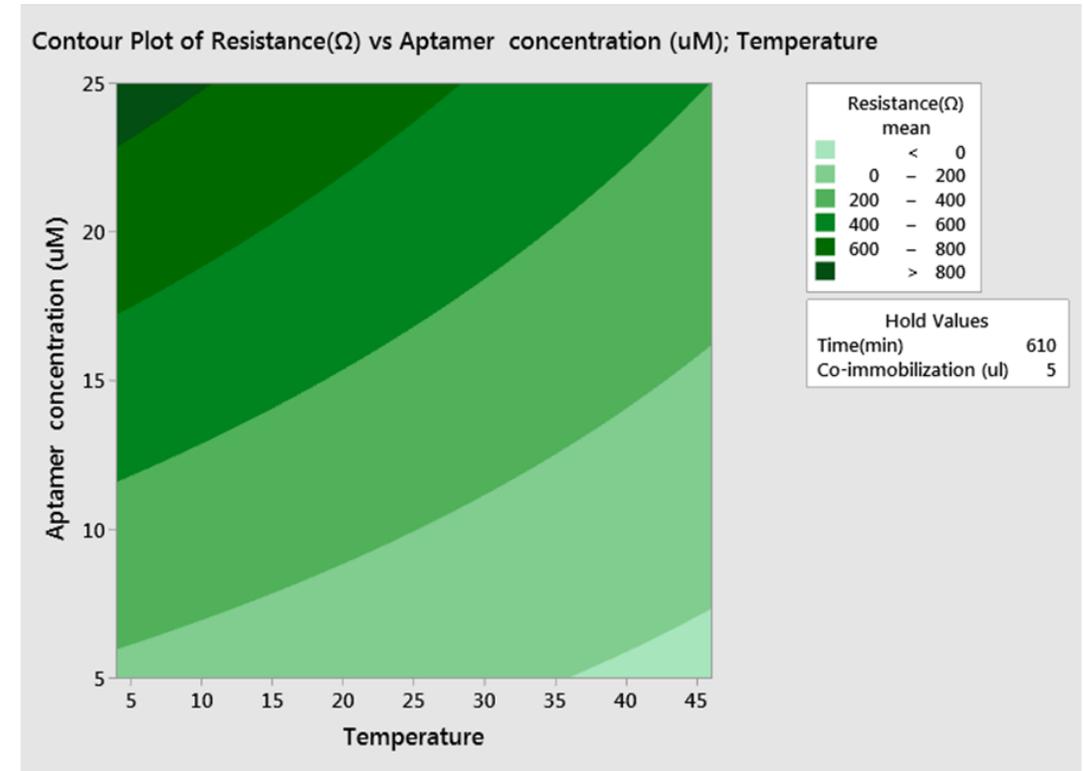
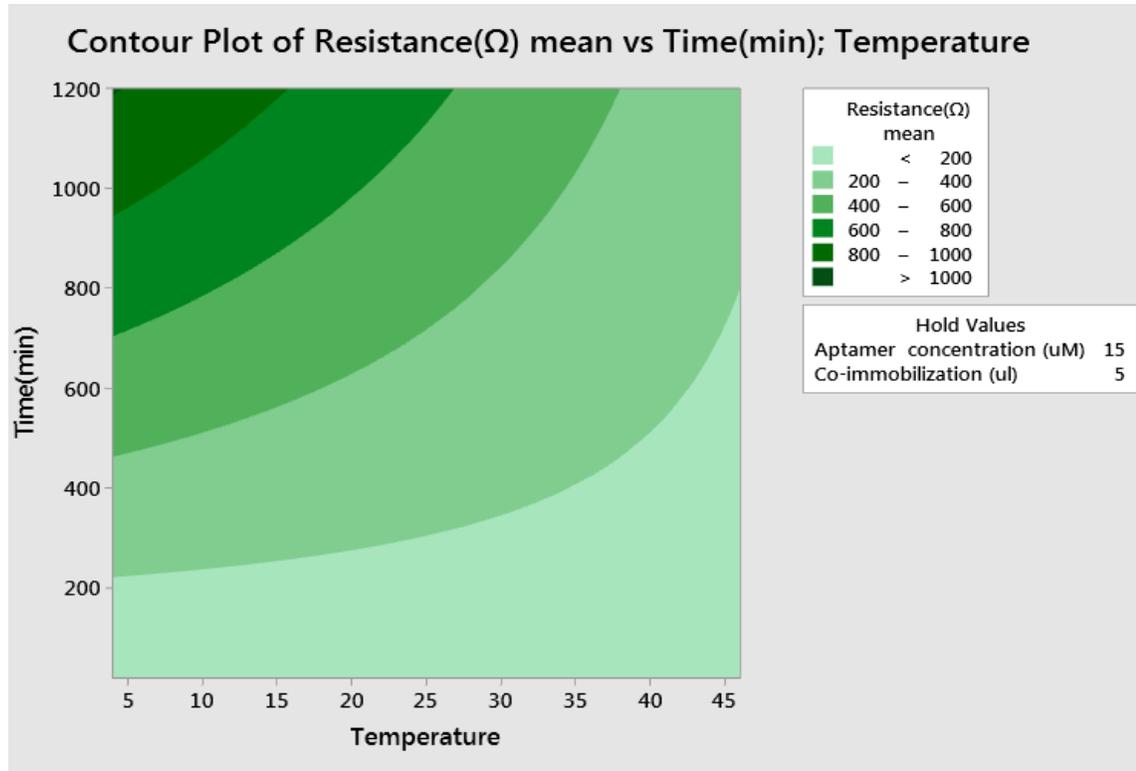
Pareto charts of the controlled parameters



Co-immobilization vs the two main parameters

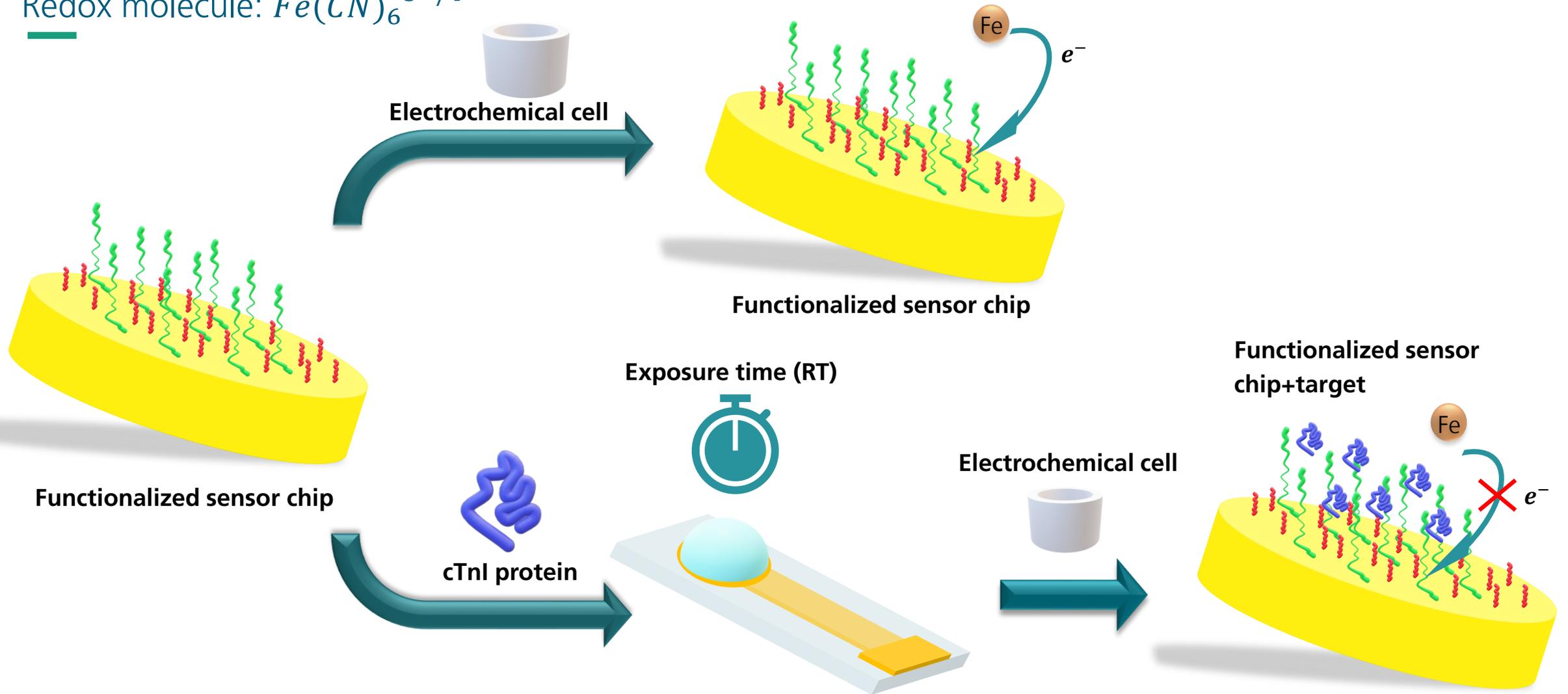


Contour plots for aptamer concentration and time



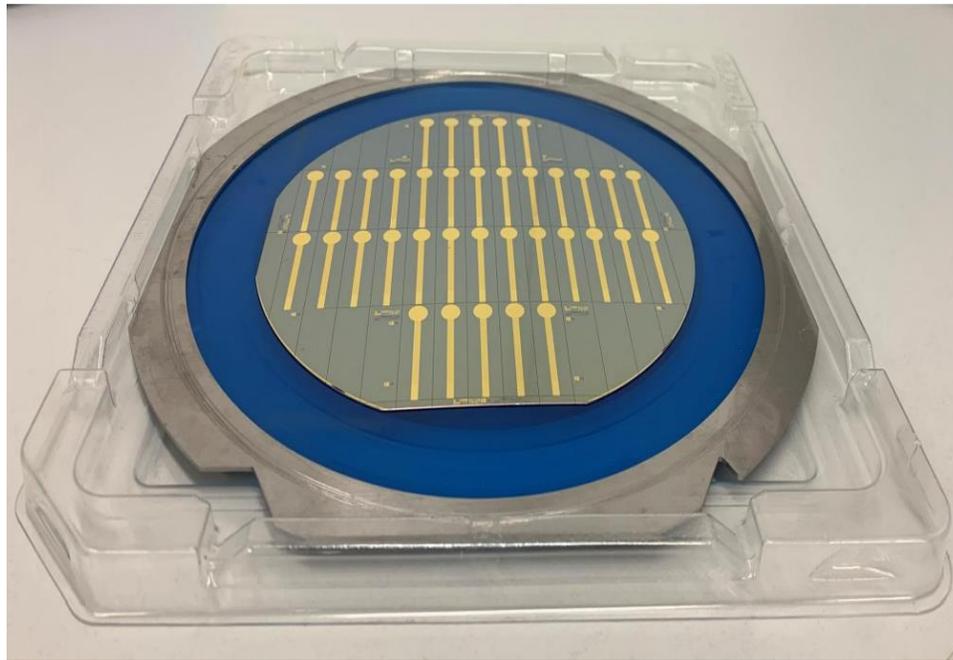
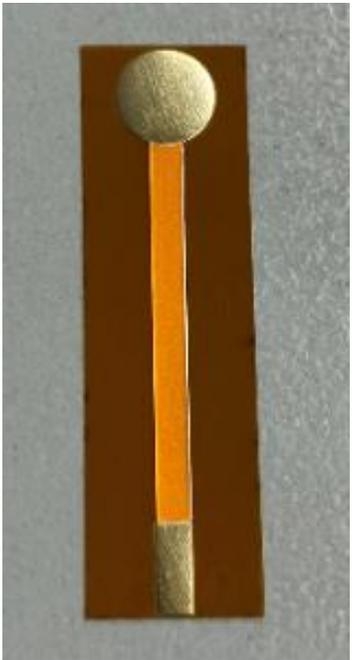
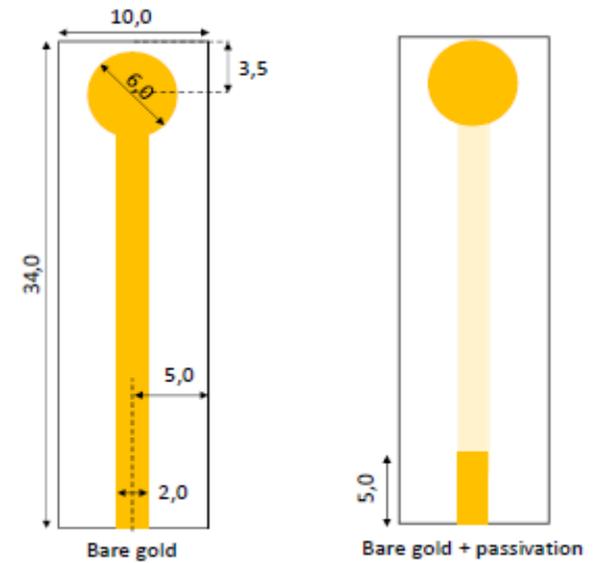
Detection methodology and electrochemical characterization

Redox molecule: $Fe(CN)_6^{3-/4-}$



Sensor chip initial options

- PCB electrode with a chemical AuNi and with galvanic soft gold(with not Nickel)-add technology
- Silicon-base electrodes with a gold layer
- PEN-base gold electrodes



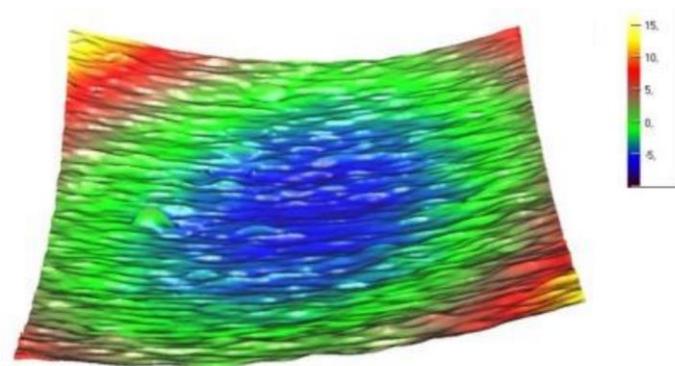
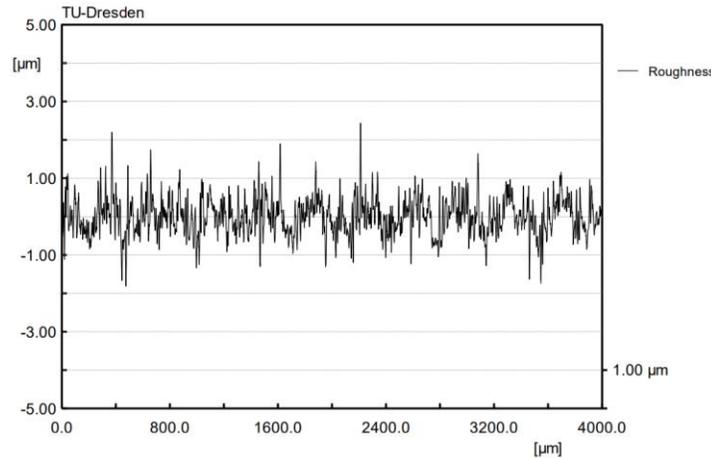
PEN-gold electrodes

Polyethylene naphthalene, pure gold , inkjet-printing

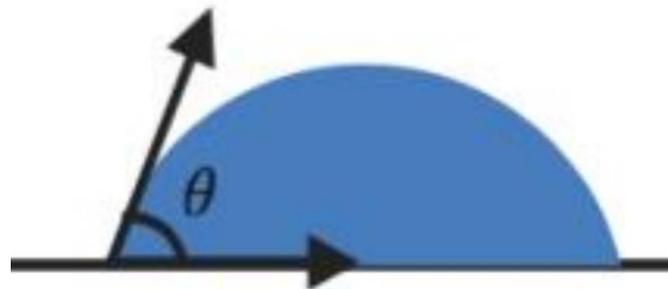


Laser profilometry measurements

- Roughness average 0.3710 ± 0.04

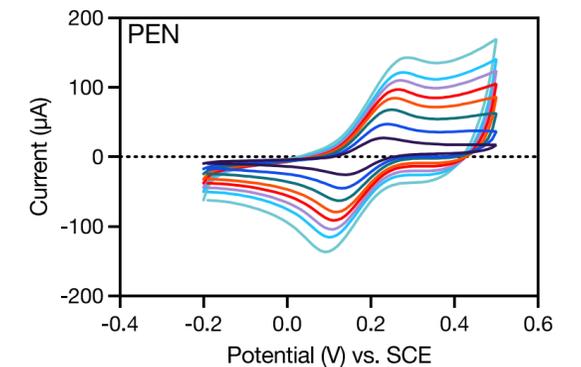
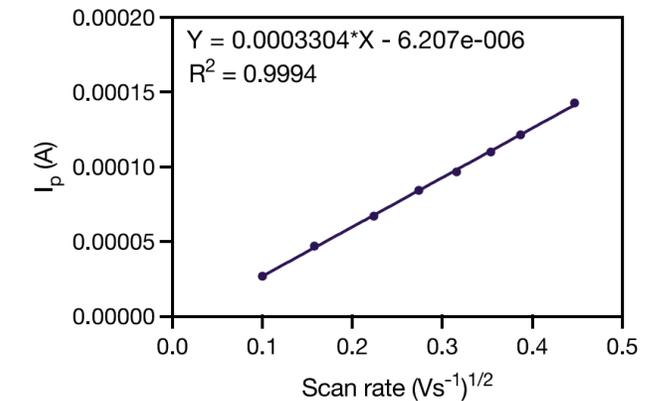


Contact angle	Water	DMSO	DIM
PEN	60.8 ± 2.6	16.7 ± 0.3	11.8 ± 0.3



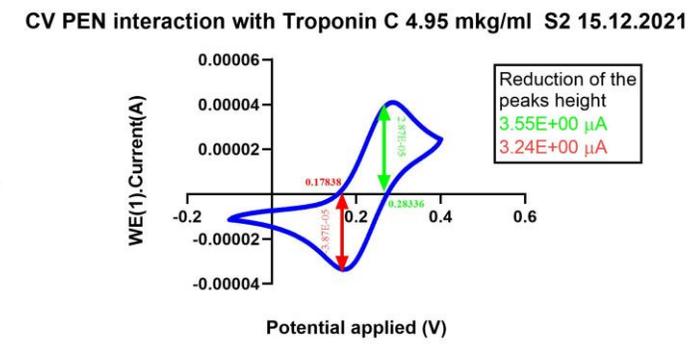
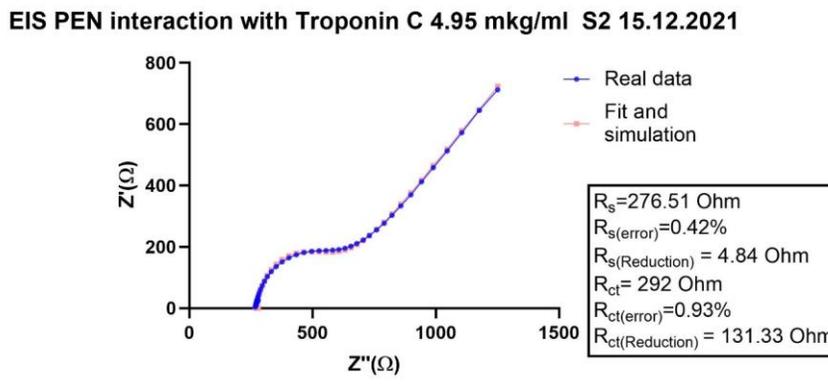
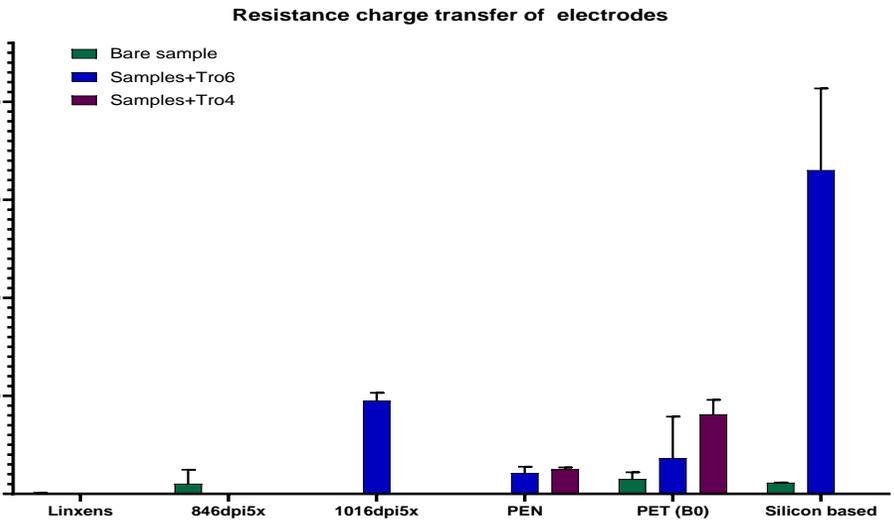
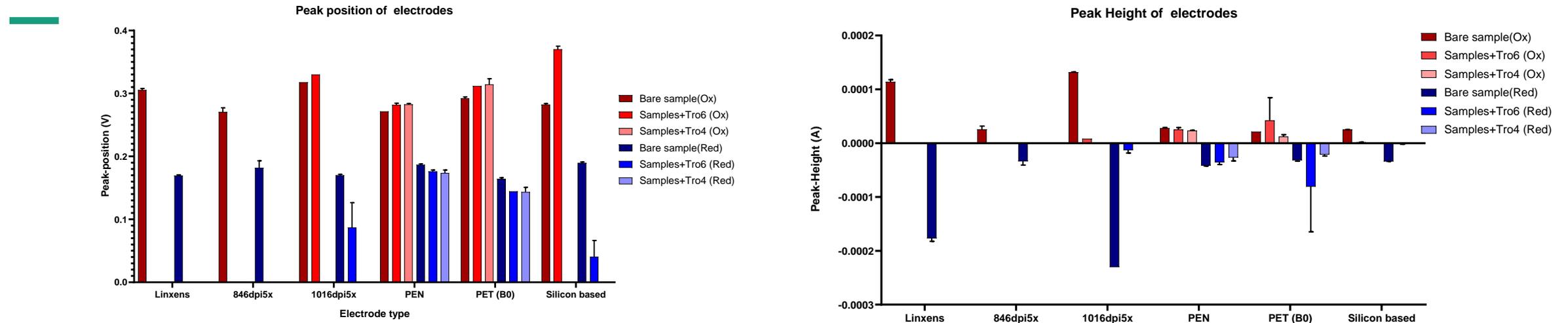
Electrochemical Surface area

- $A_{effective} = 0,4558 \text{ cm}^2$
- $A_{geometric} = 0,2826 \text{ cm}^2$



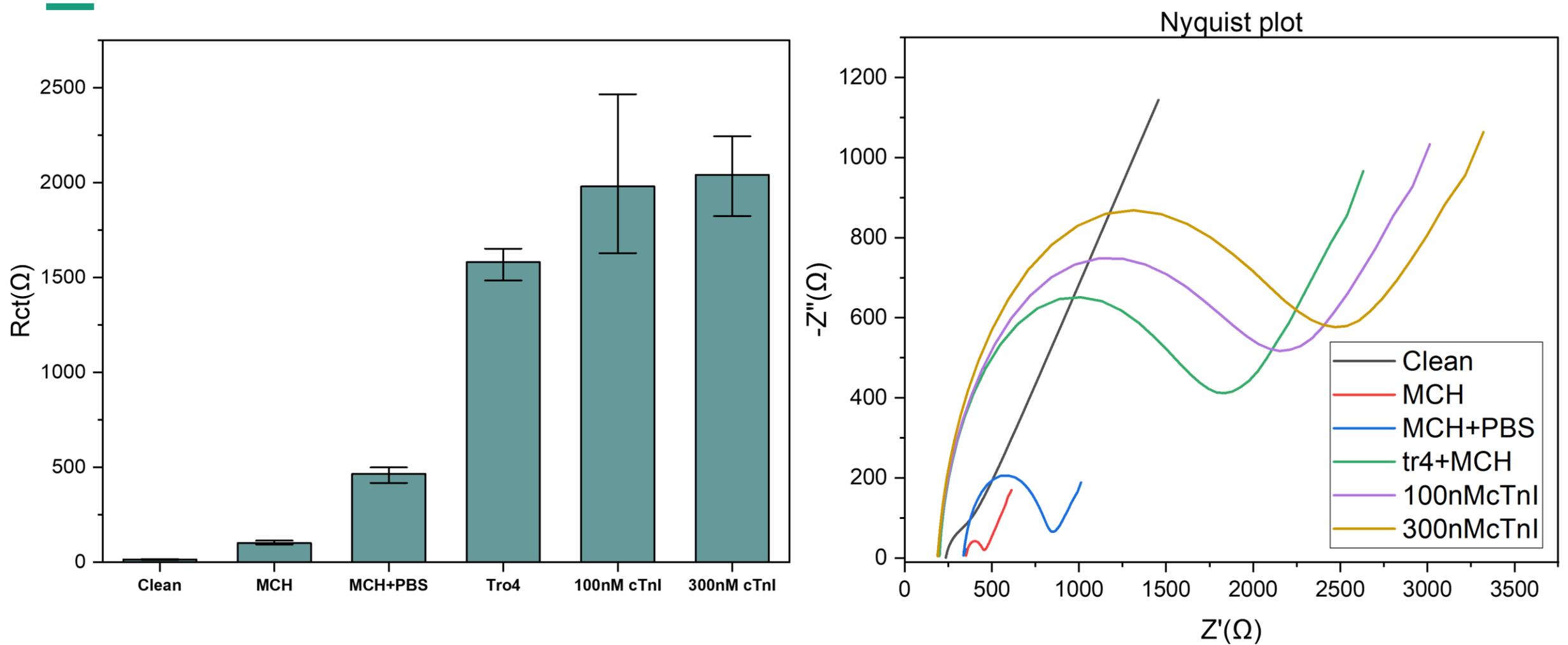
Summary of results of immobilization on different manufacturing electrode technologies

Results use in a publication



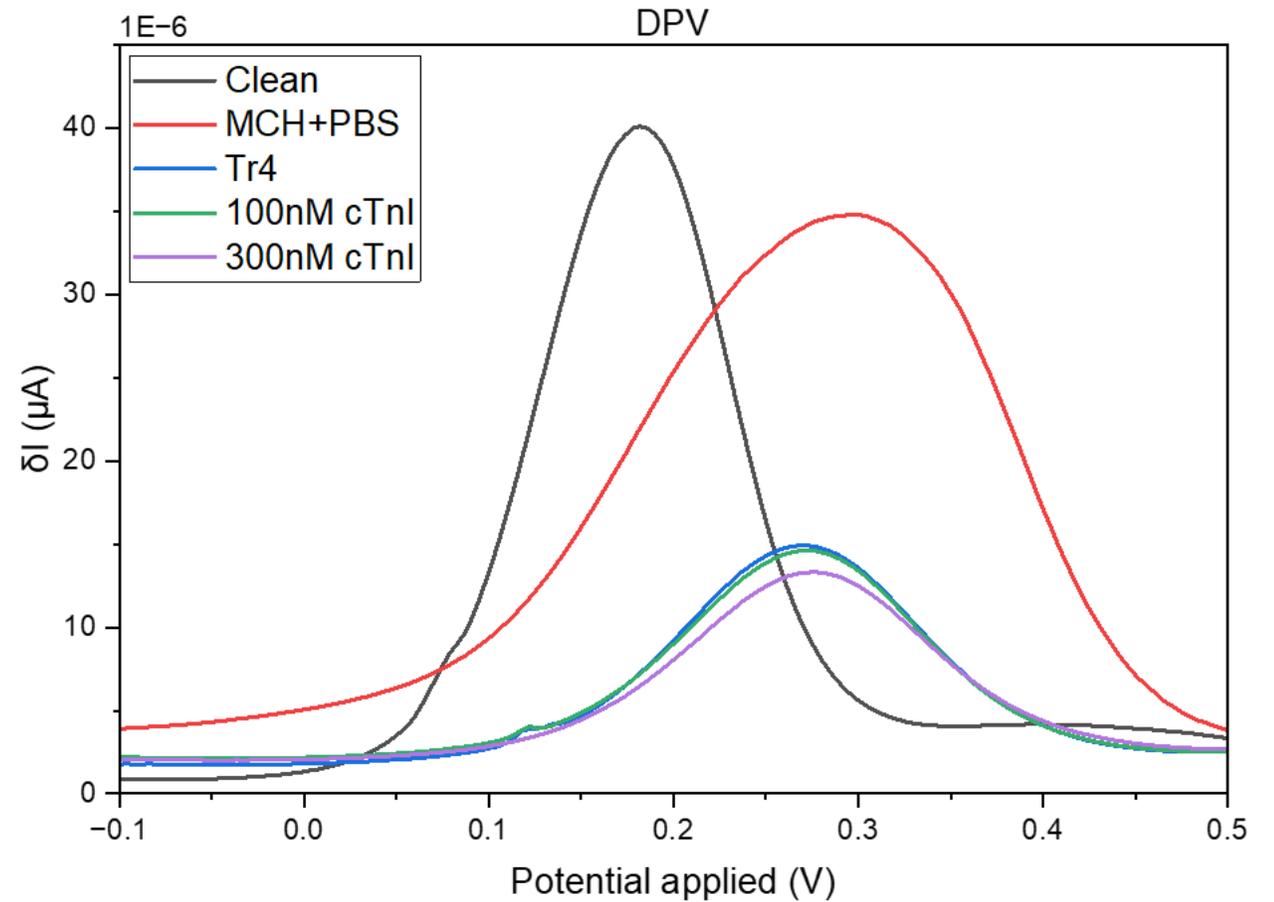
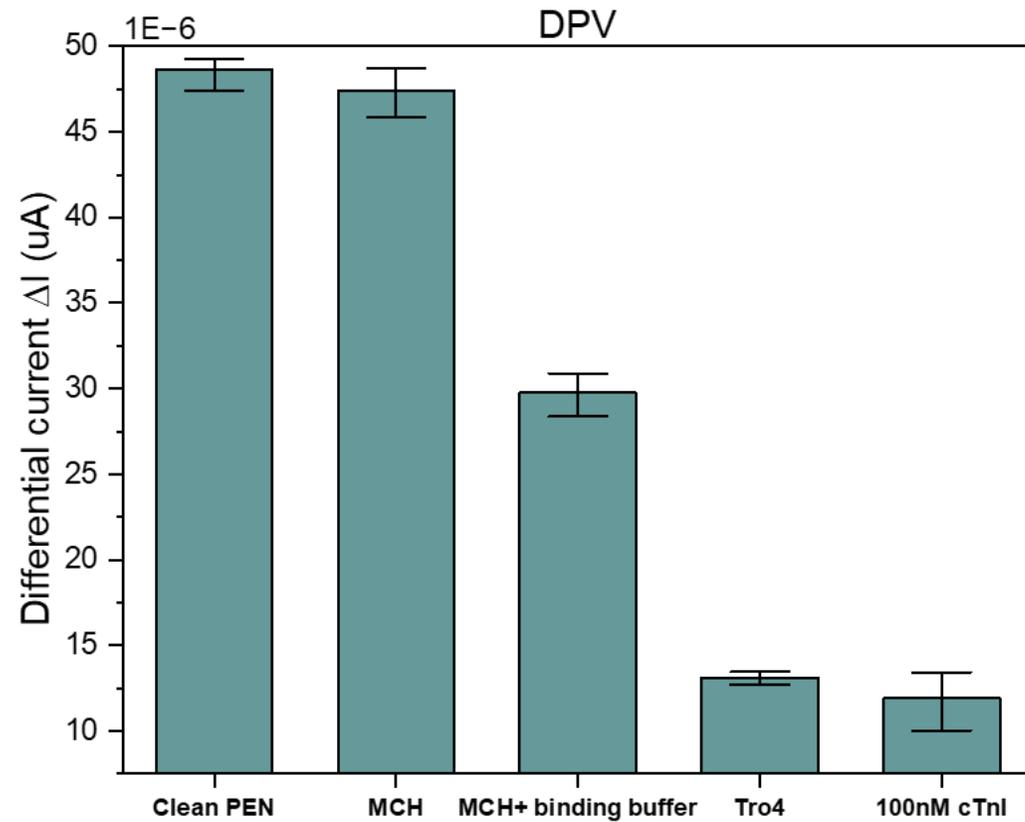
Results of immobilization steps and binding events obtain with EIS

PEN substrate, 1mM $Fe(CN)_6^{3-/4-}$



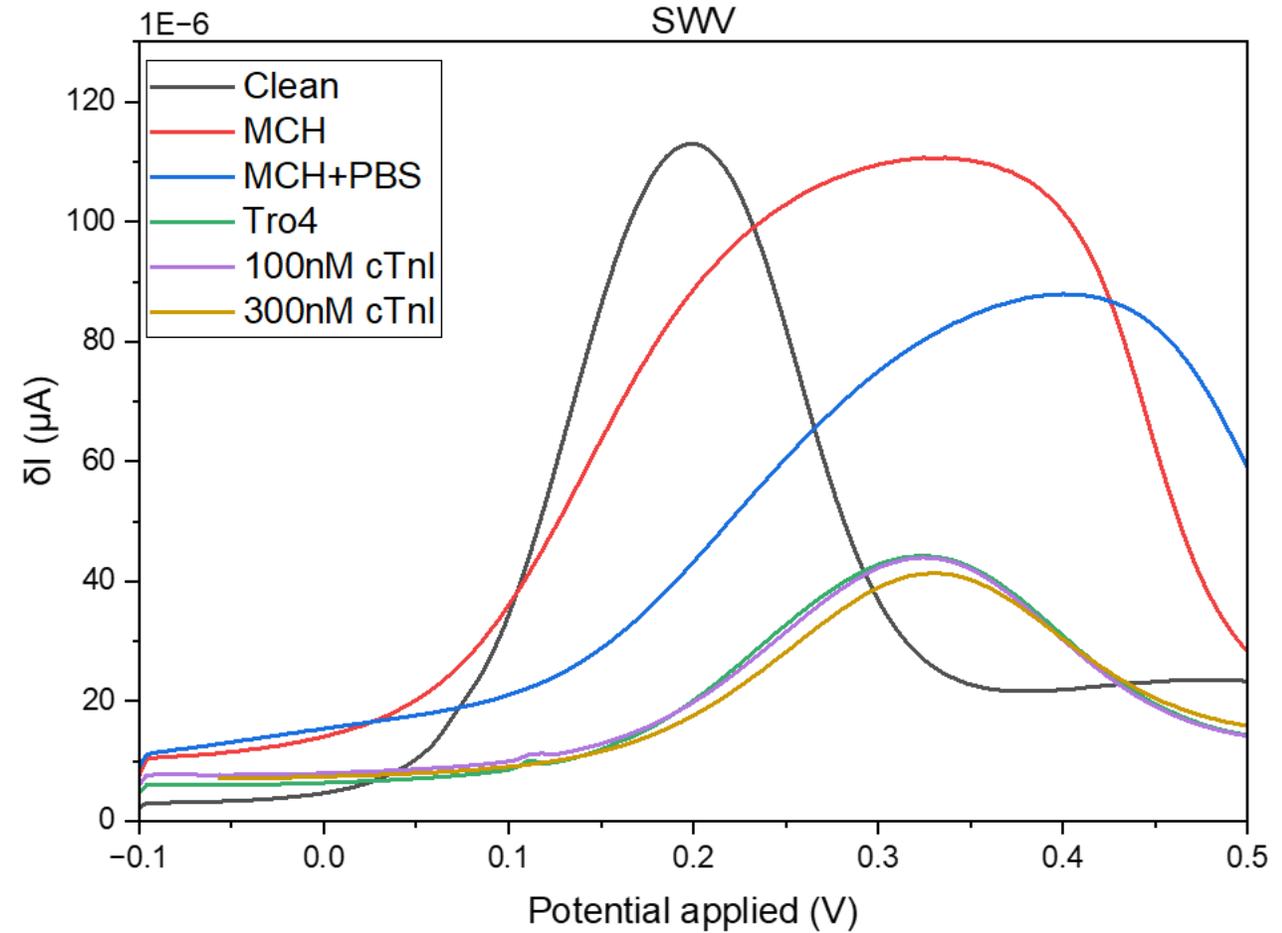
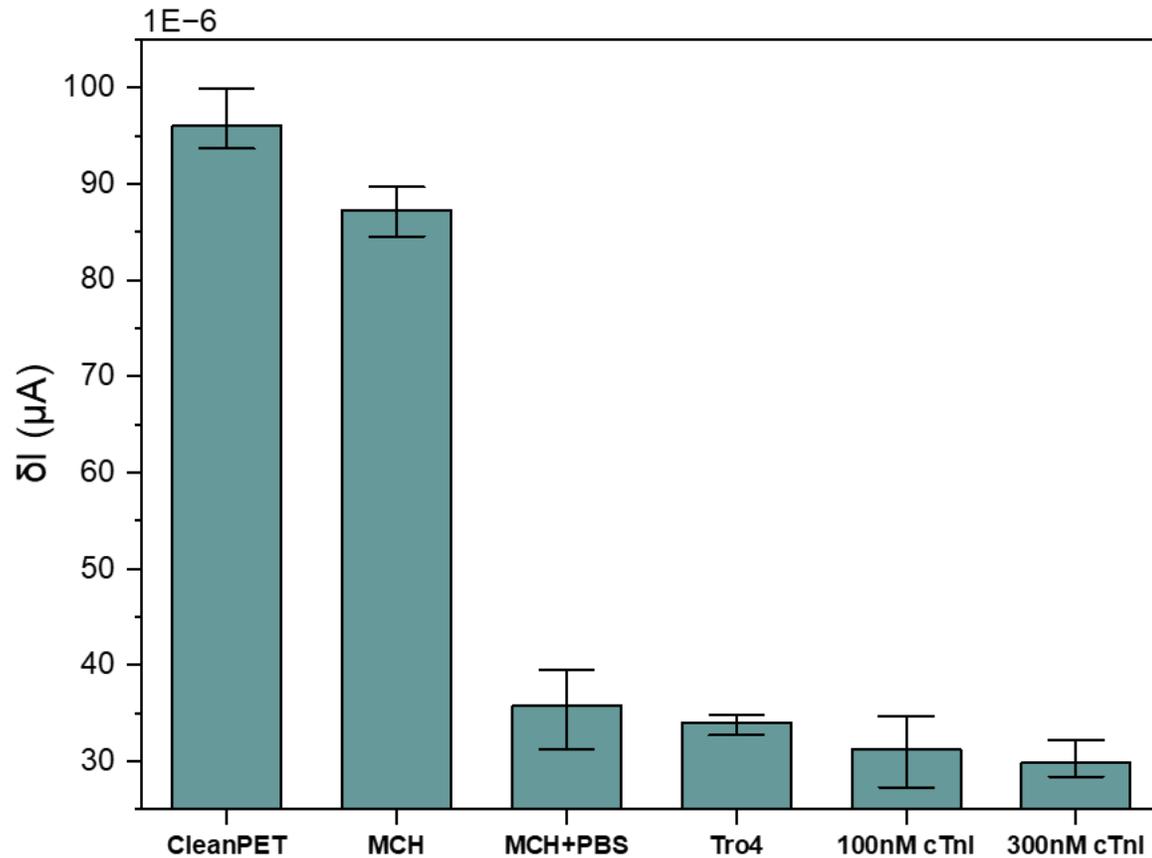
Results of immobilization steps and binding events obtain with DPV

PEN substrate, 1mM $Fe(CN)_6^{3-/4-}$

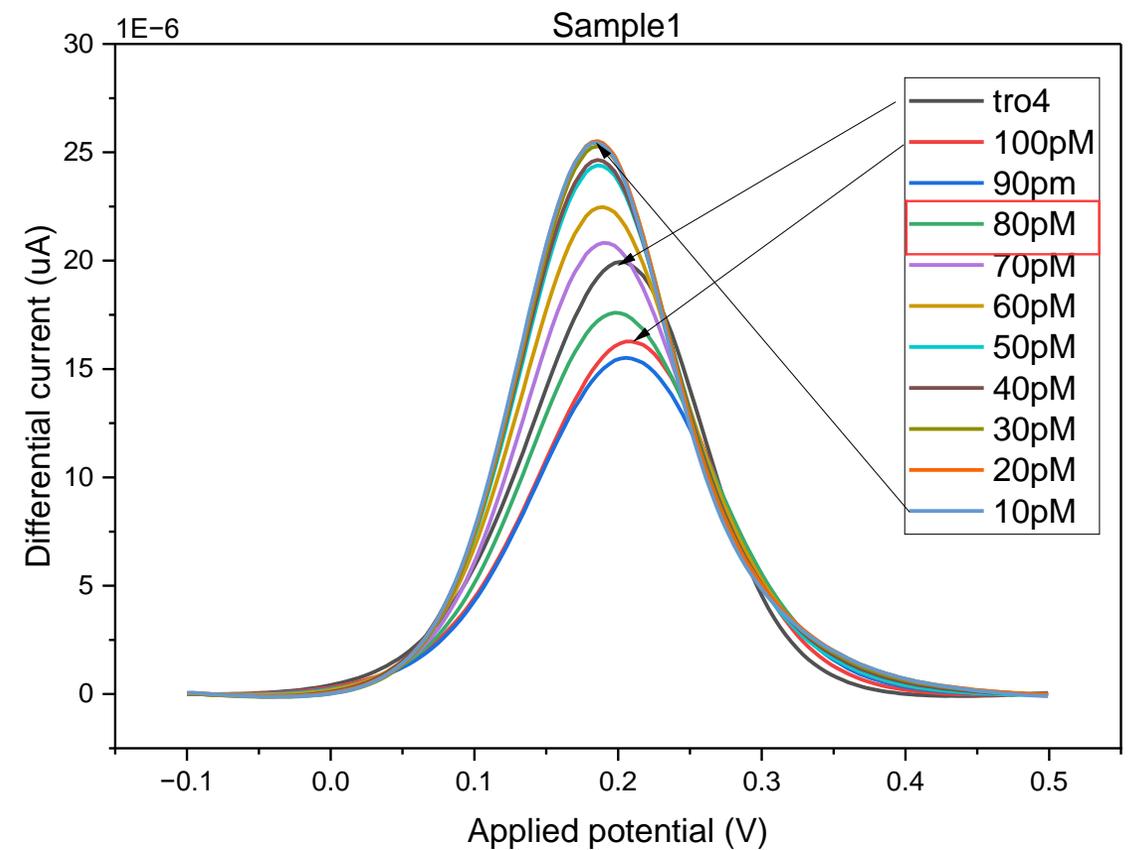
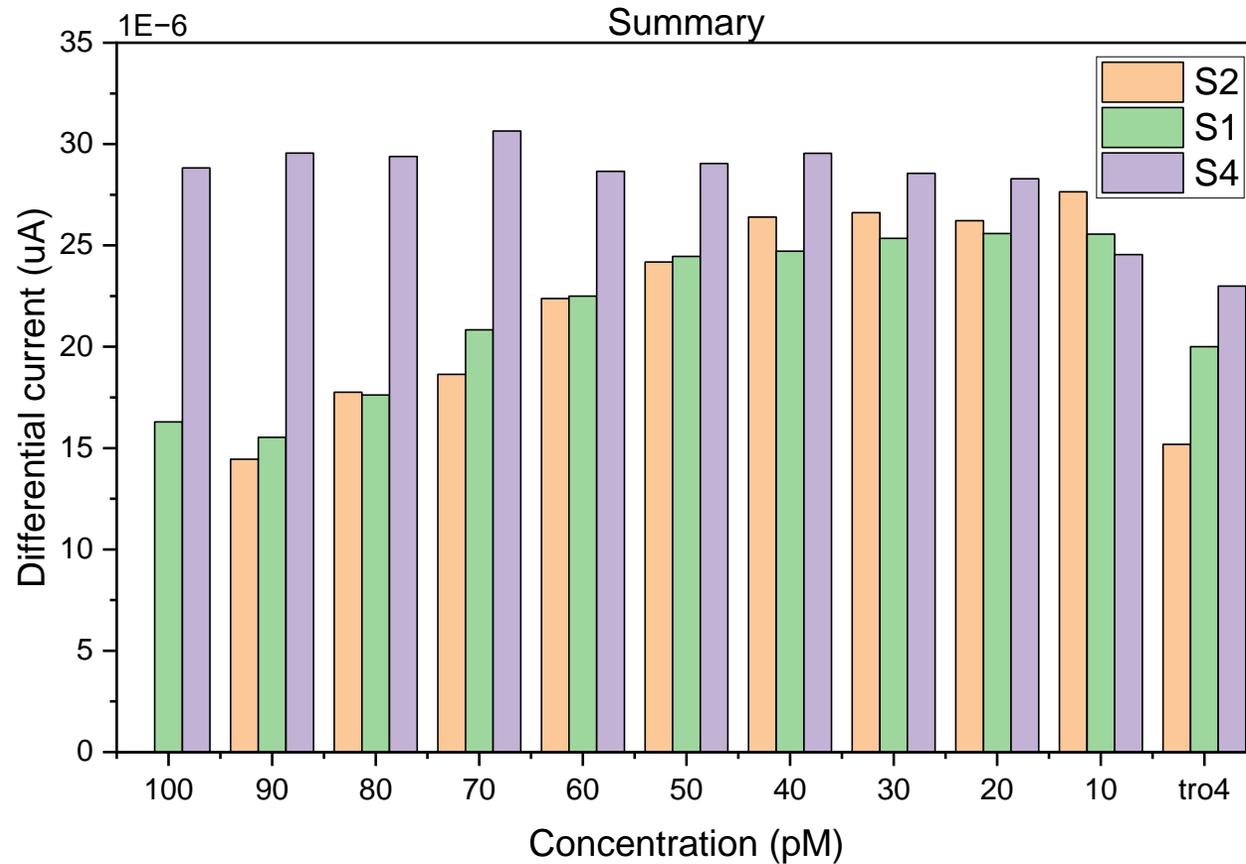


Results of immobilization steps and binding events obtain with SWV

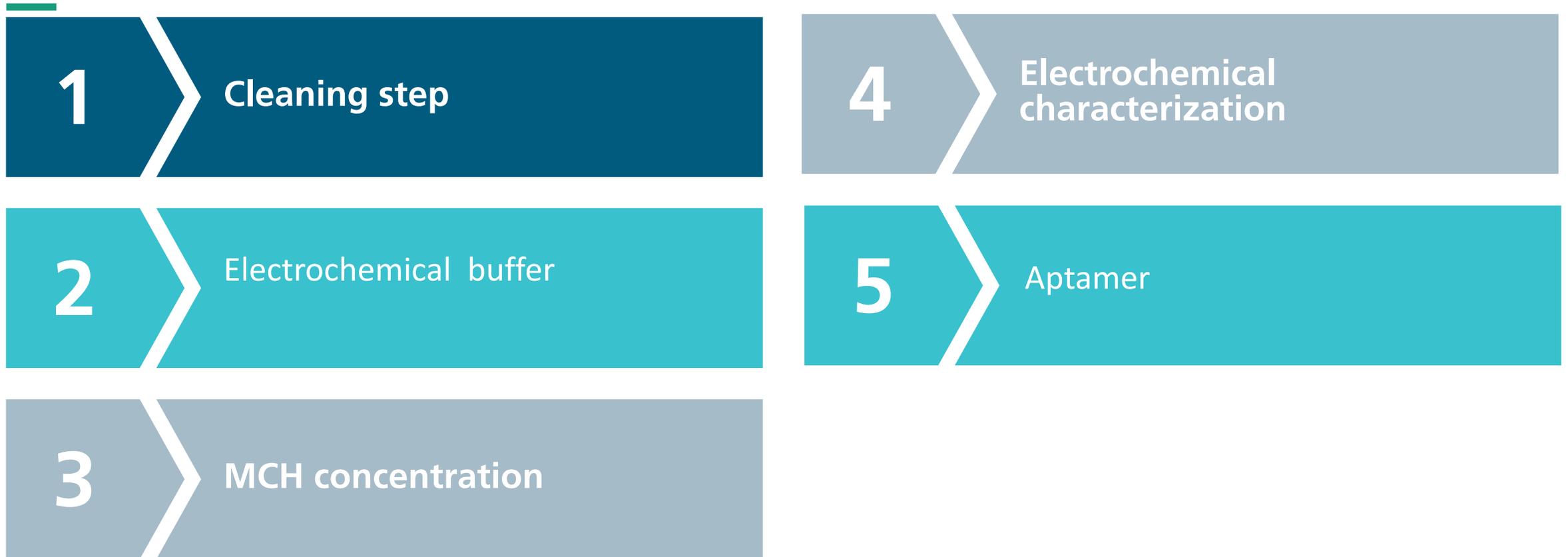
PEN substrate, 1mM $Fe(CN)_6^{3-/4-}$



Results of the tests done with functionalized PEN-based electrodes in binding experiments at different protein concentrations

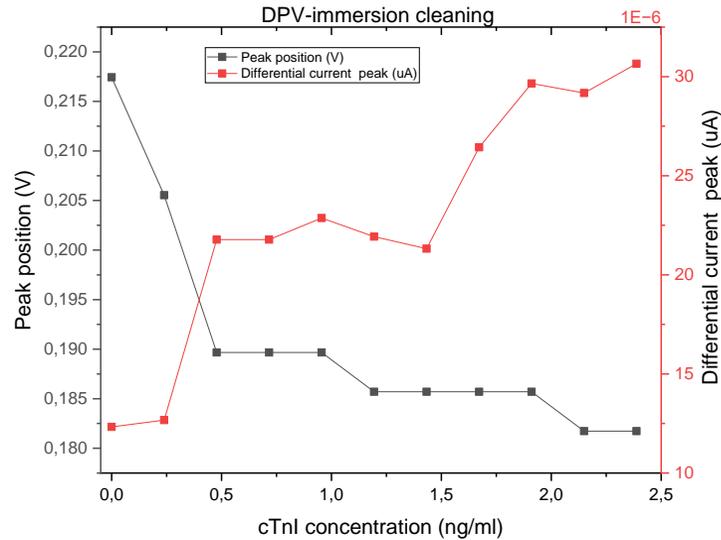


Identification of behaviour for functionalized samples

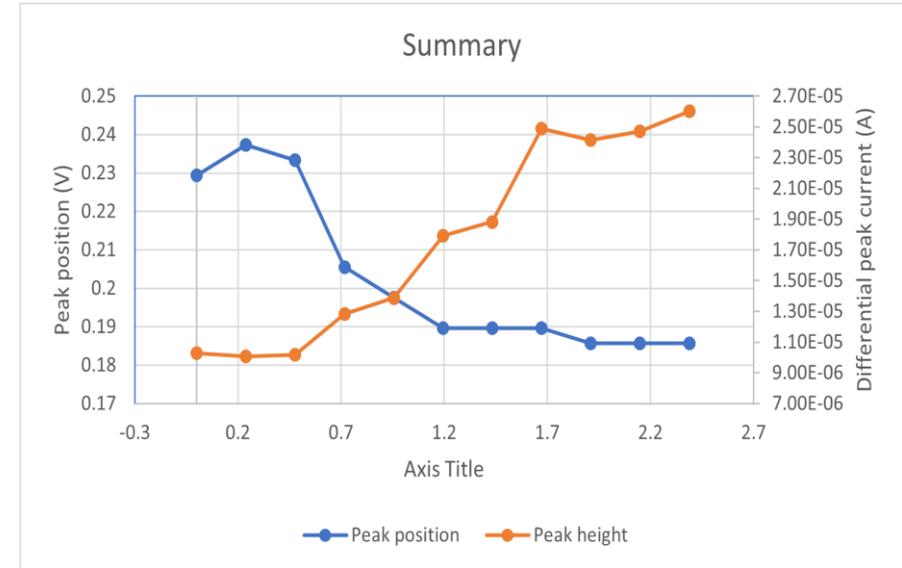


Summary of results for the behaviour of functionalized samples against different concentrations of cTnI protein

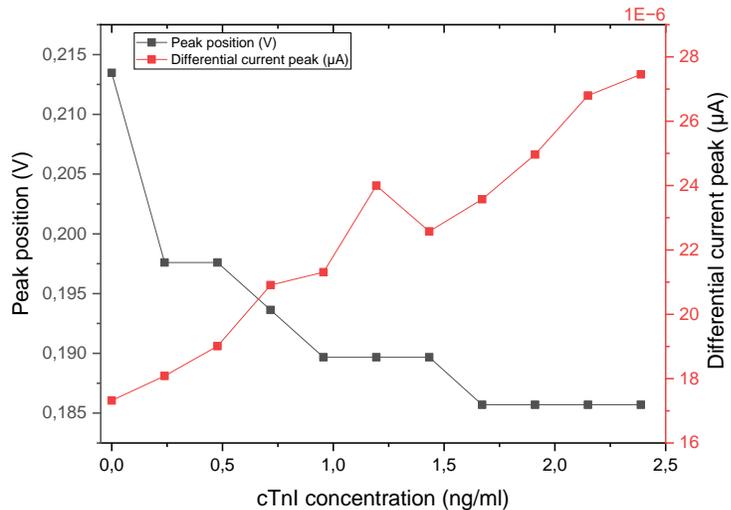
Cleaning after exposure



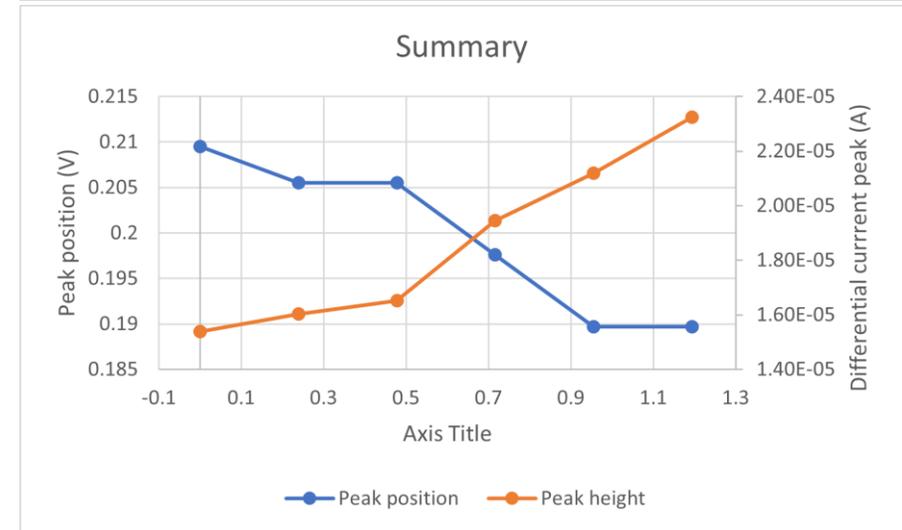
Change of electrochemical buffer



Change of MCH concentration



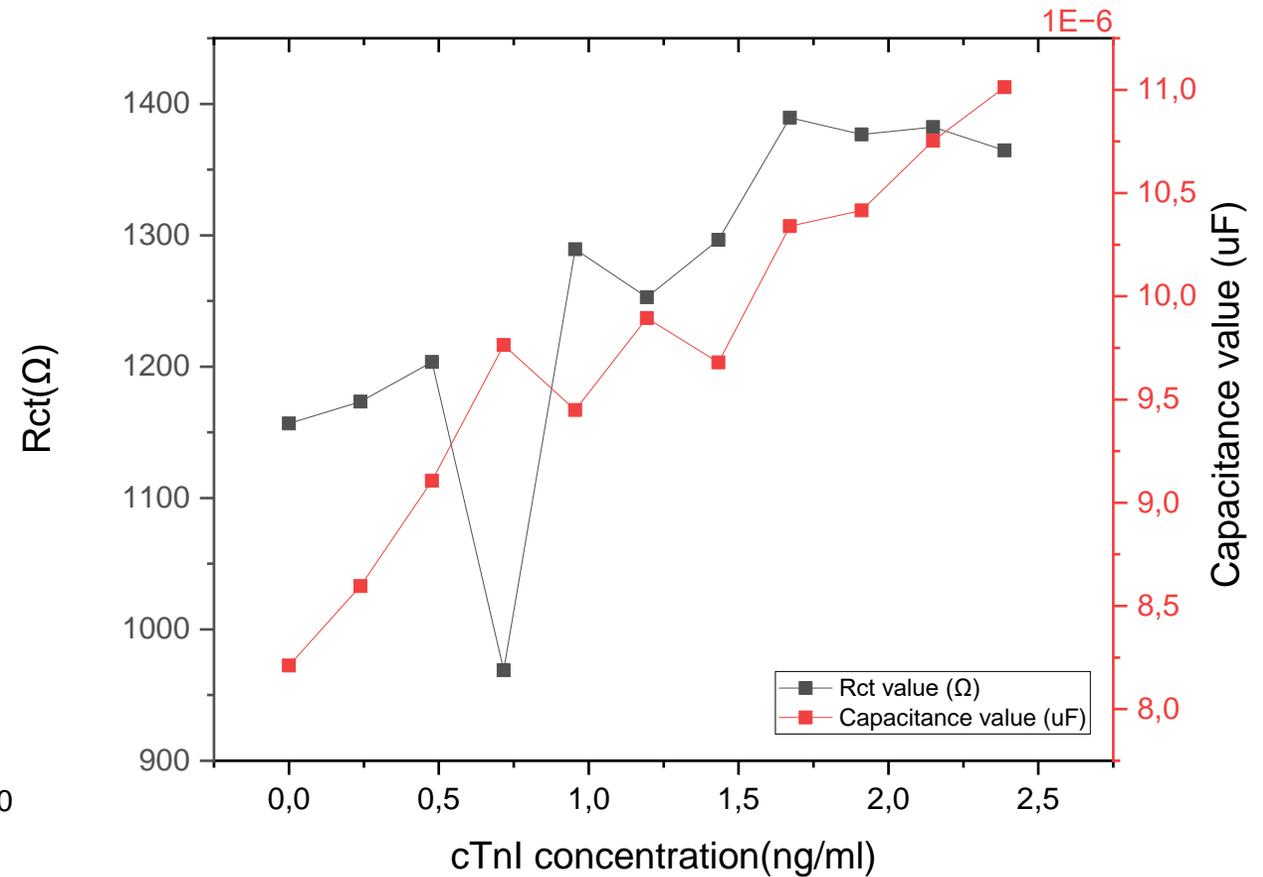
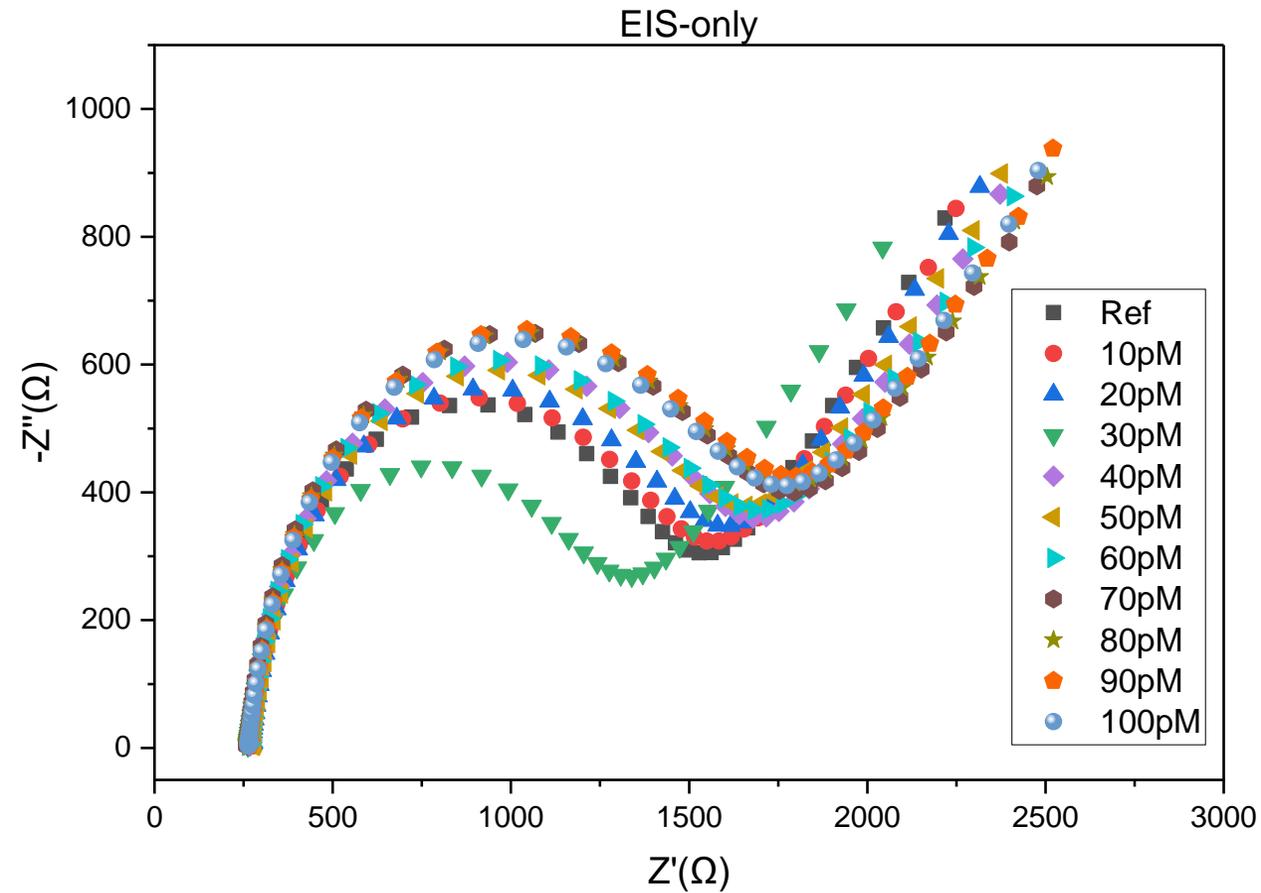
Change of aptamer



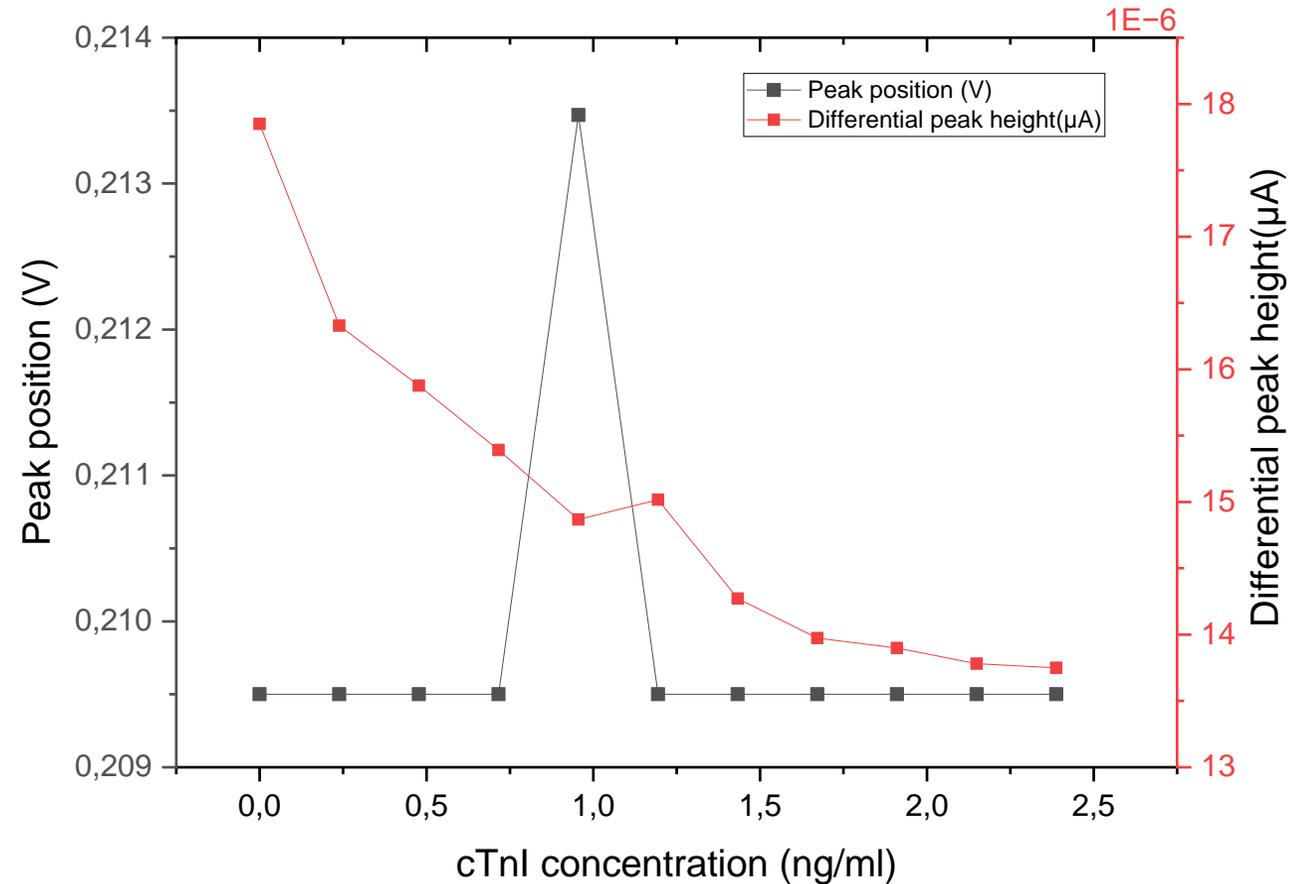
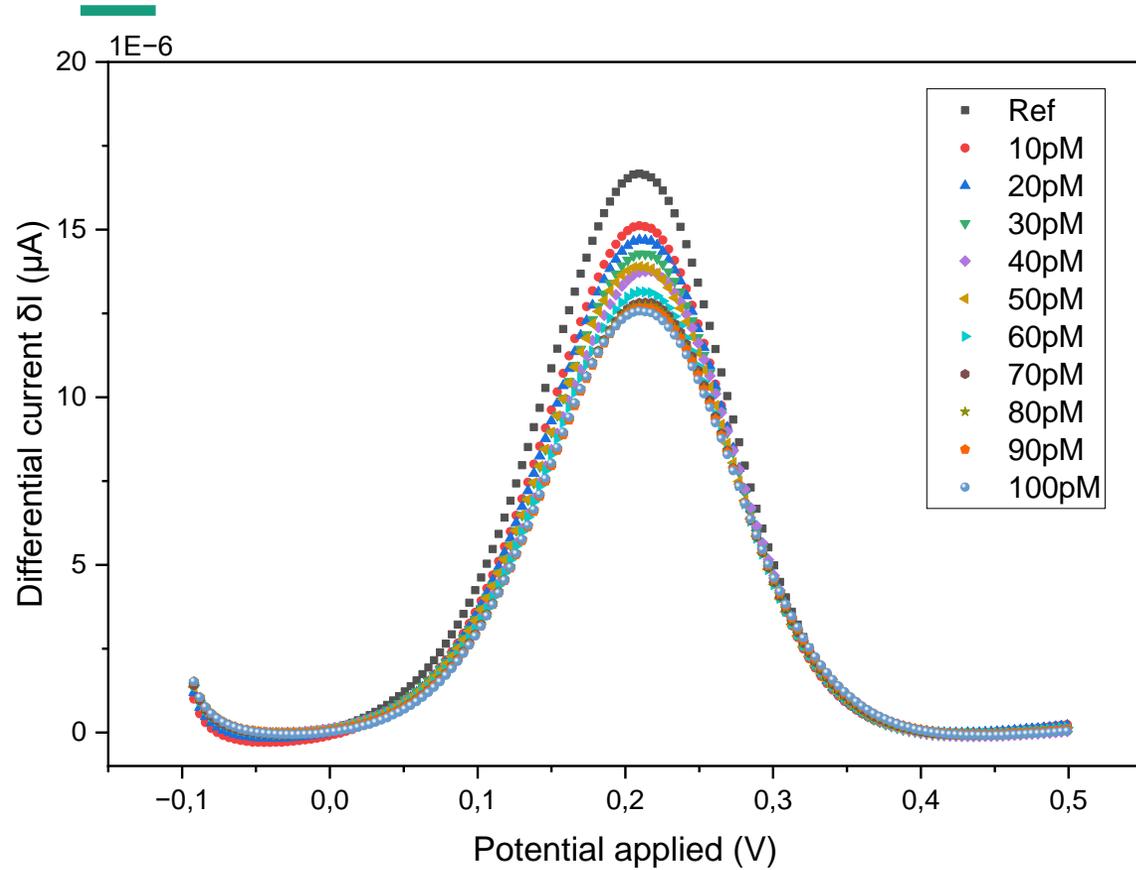
Single technique approach



Results obtained with EIS technique (single technique approach)

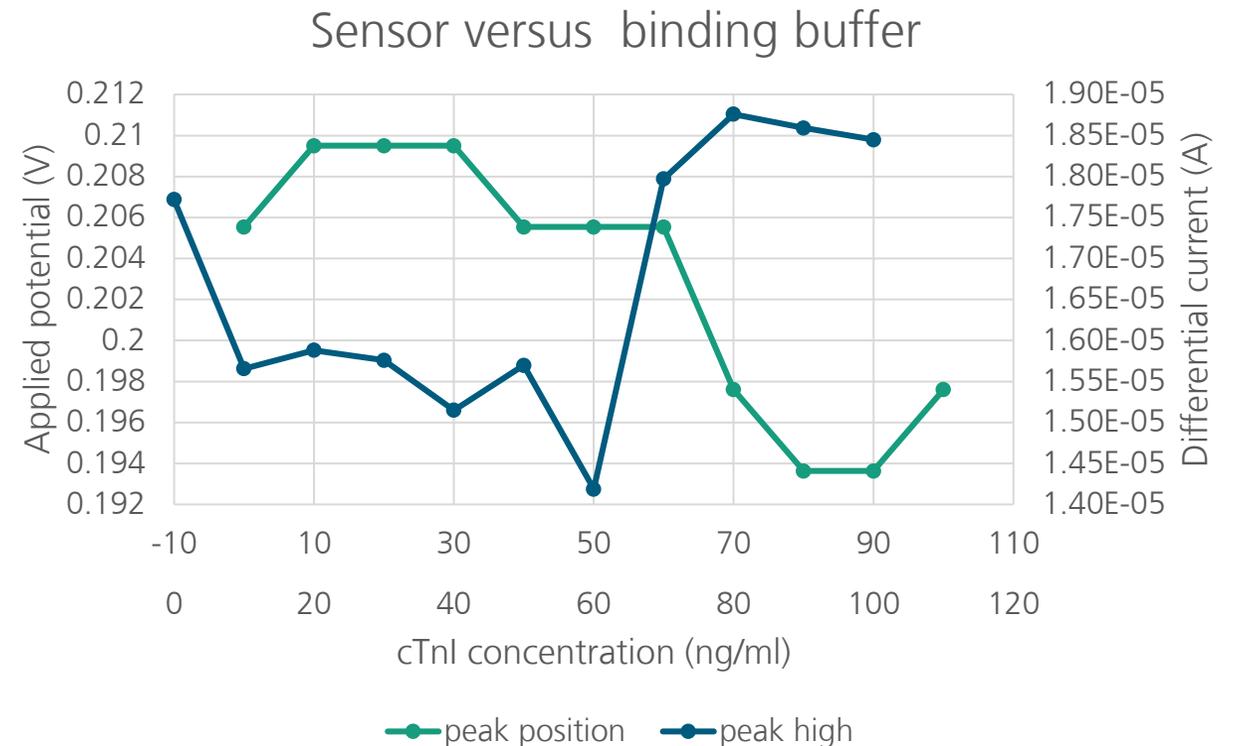


Results obtained with DPV forward (single technique approach)



Next step in the research

- Test of several sample to have a statistical analysis
- Test more complex solutions in the detection experiments
- Enhance response signal of the system
- Miniaturization of de system



Publications status

- **Immobilization of thiolate-aptamer on different manufactured electrodes (main author)**
- **Influence in electrochemical response of PEN-based gold electrodes treated with Laser-induced periodic surface structure for its use as biosensor of cardiac biomarkers(main author)**
- **Cleaning of LTCC, PEN, and PCB Au electrodes towards reliable electrochemical measurements (co-author)**
- **Advanced Hydrogel-based Wearable Electrochemical Biosensors with integrated Microfluidics(review, co-author)**

Thank you for your attention

Kontakt

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