

# Bioinspired nanomaterials

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nanoSeminar  
Institute of Materials Sciences  
11.11.2010



Forschungszentrum  
Dresden Rossendorf



- **Forschungszentrum Dresden-Rossendorf**
- **Institute of Radiochemistry**
- **Microbe radionuclide interaction**
- **Bacterial S-layers**
- **Metall binding by bacterial S-layers**
- **Application potential of bacterial S-layers**
- **S-layer based materials**
- **Biomass productions**
- **Summary**
- **Future cooperation IRC-FZD with IfWW-TUD**



As of 01.01.2011

**HZDR**

HELMHOLTZ  
ZENTRUM DRESDEN  
ROSSENDORF

**Ion-Beam Physics and Materials  
Research**

**Radiation Physics**

**Radiochemistry**

**Radiopharmacy**

**Safety Research**

**High Magnetic Field Laboratory**

**Advanced Materials  
Cancer Research  
Nuclear Safety  
Research**

**Ion-Beam Center**

**Radiation Source ELBE**

**Rossendorf Beamline  
(ESRF, Grenoble)**

**PET-Center**

**TOPFLOW-Facility**

**Dresden High Magnetic  
Field Laboratory**

- **Member of the Leibniz-Association**

Change to the **Helmholtz Association**  
as of 01.01.2011, as **HZDR**  
(Result of the evaluation 2007)

- **Foundation:** 01.01.1992 (e.V.)
- **Employees:**
  - ~ 800 total (incl. PhD, Annex, projects, trainees)
  - ~ 400 basic funding
  - ~ 370 scientists
- **Budget 2009:**
  - ~ 60 Mio € basic funding
  - ~ 20 Mio € third party funding

Dresden



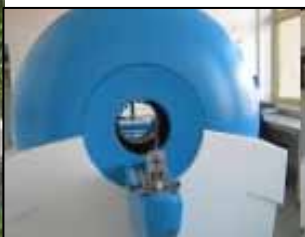
Radiochemical lab building 8b



Office and lab building 8a  
Microbiological and radiochemical labs,  
Divisions Biogeochemistry, Surface Processes, Biophysics



Leipzig



Radiochemical laboratories  
FZD research site Leipzig, Division Reactive Transport



ESRF Beamline ROBL, France  
Division Molecular Structures

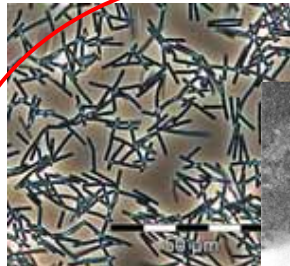
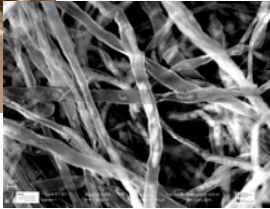
Grenoble



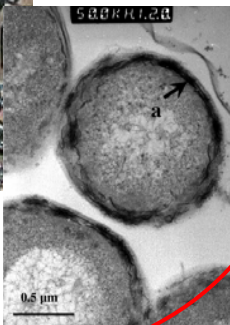
# Radio-ecological research on the behaviour of actinides in nature



Fungi



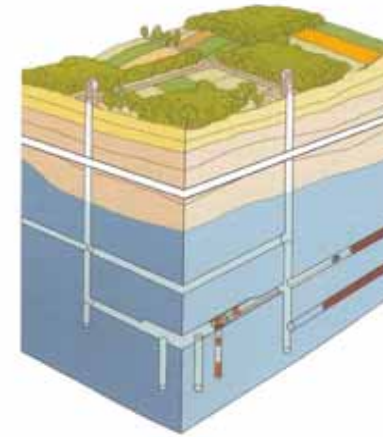
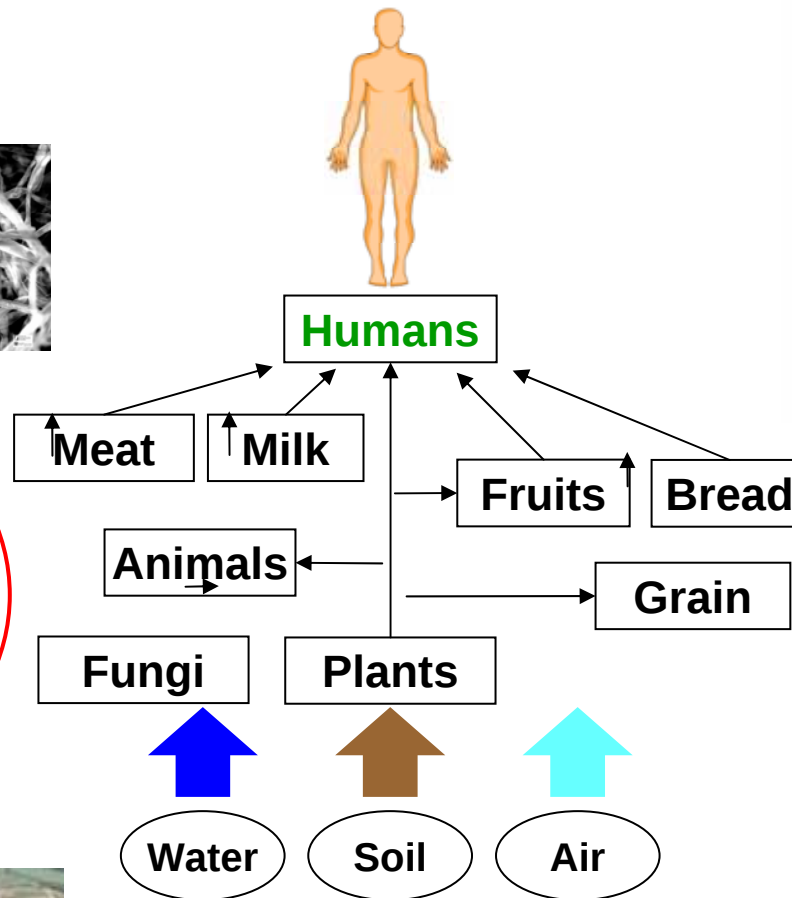
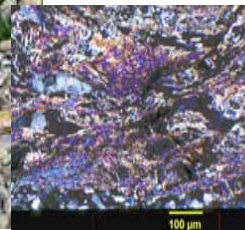
Bacteria



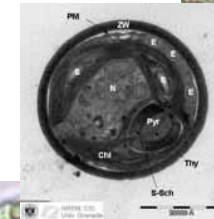
Water



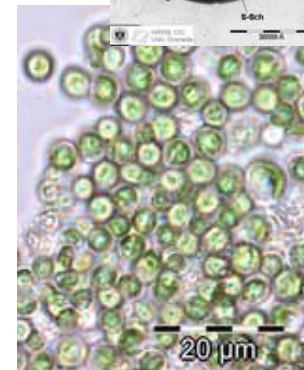
Soil



Long term disposal of radioactive waste



Algae/plants



### Natural and anthropogen sources of actinides and radionuclides

Uranium deposits and minerals



Nuclear power plants



Reprocessing plants



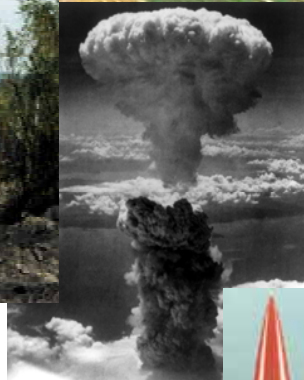
Storage tanks



Nuclear warhead, USA



Uranium mining waste piles



Nuclear explosions

Uranium ammunition

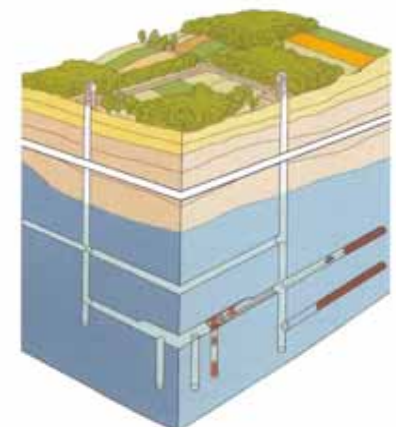


Cement production



Phosphate fertilizer

Storage



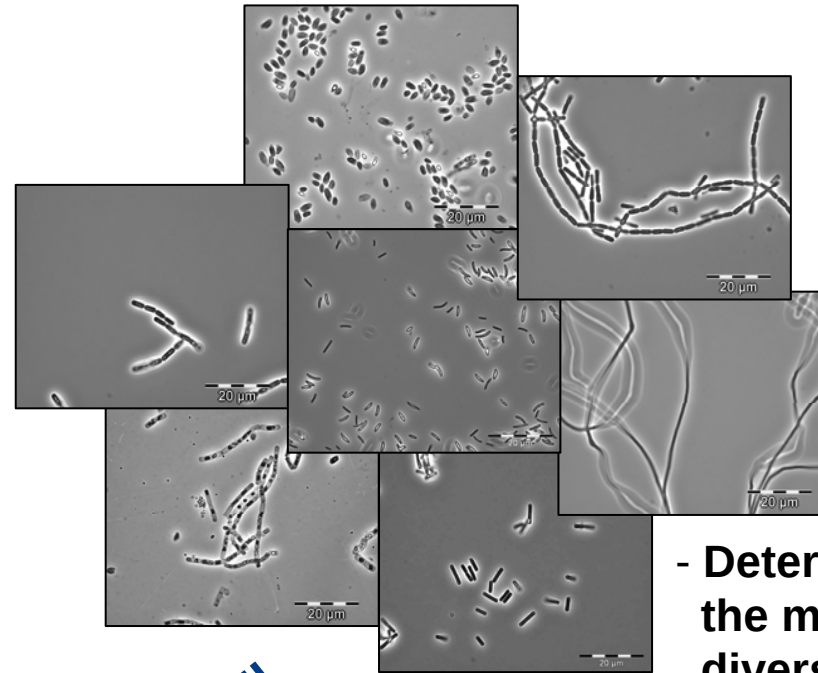
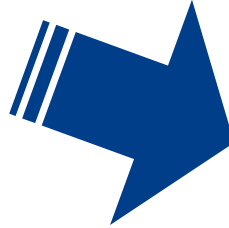
→ Our aim: protect people and environment from being affected by actinides

# Fundamental research

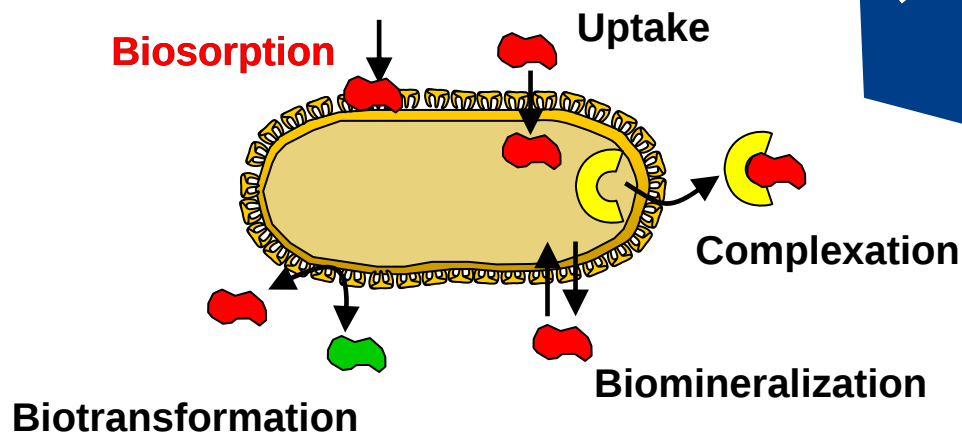
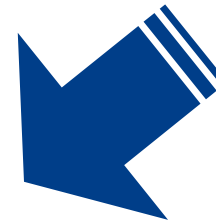




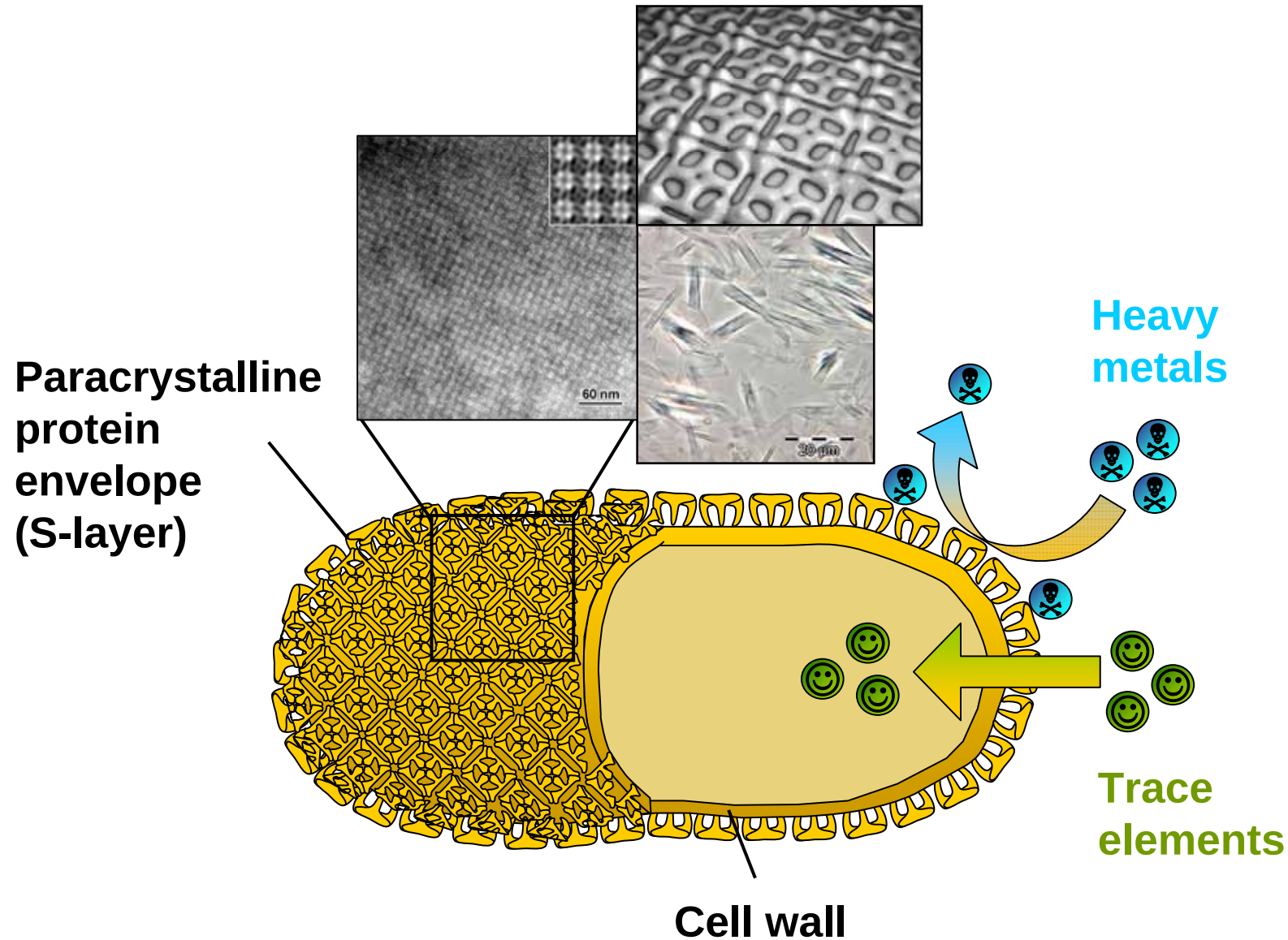
Uranium mining waste pile  
“Haberland” nearby Johann-  
georgenstadt, Saxony



- Determination of the microbial diversity via gen analysis
- Isolation of bacteria

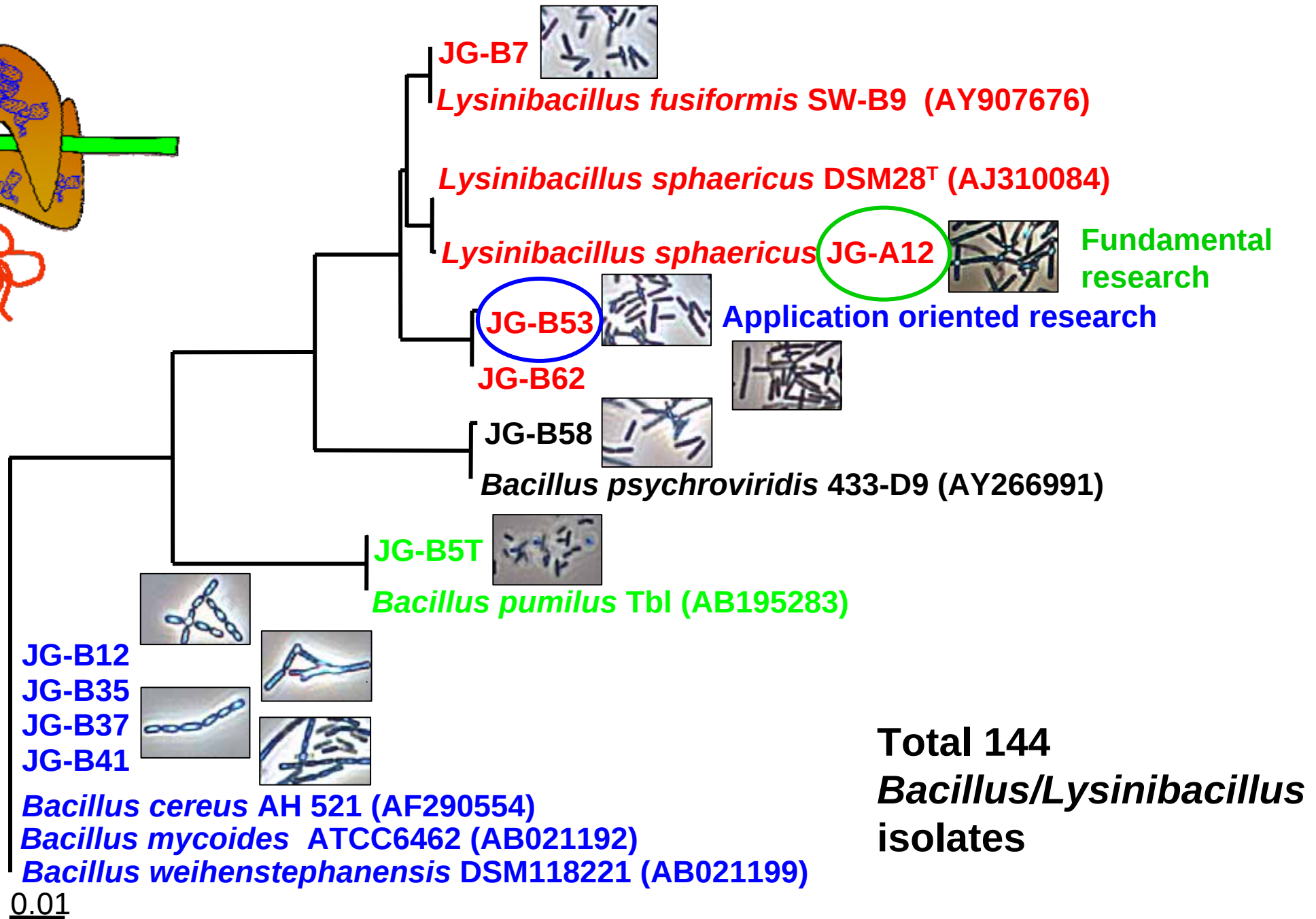
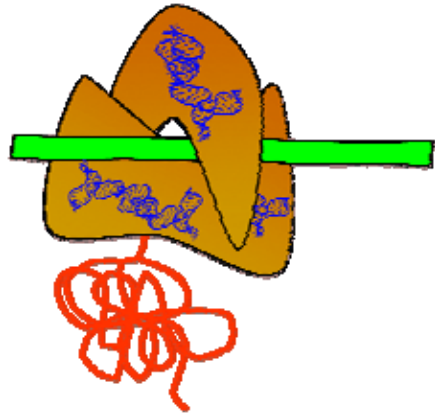


Research on the interaction  
of actinides with  
microorganisms



## Interesting properties of S-layer

- Self assembling proteins
- Multifunctional
- Intelligent interface to the bacterial environment
- Metal binding
- Some with high stability



## Elements



## Molecular biology

## ICP-MS

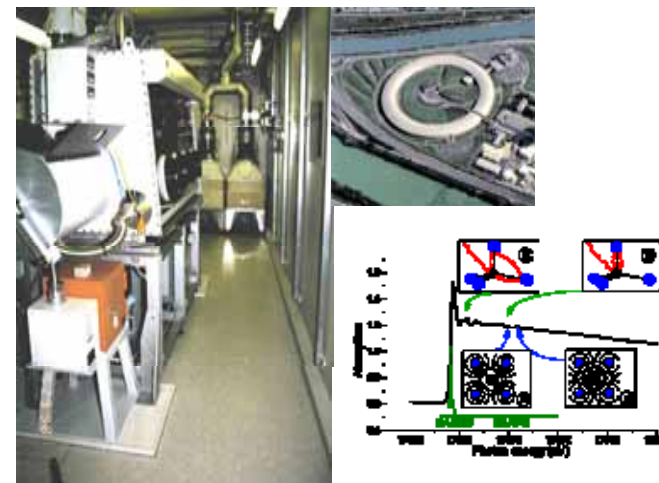


## Microscopy



## Spectroscopy

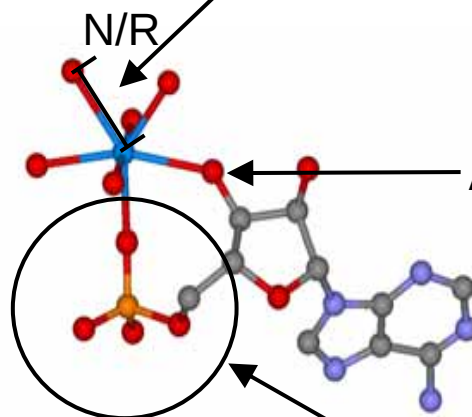
## XAS

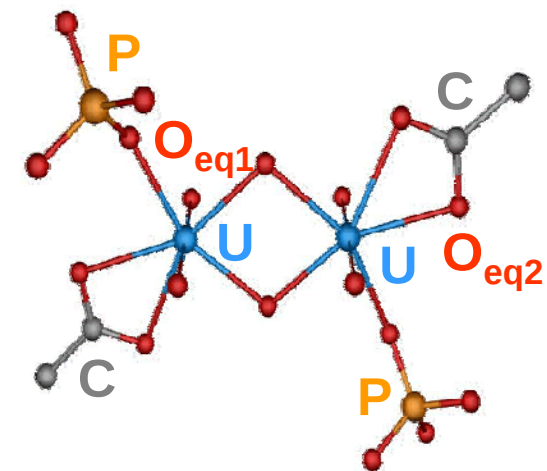
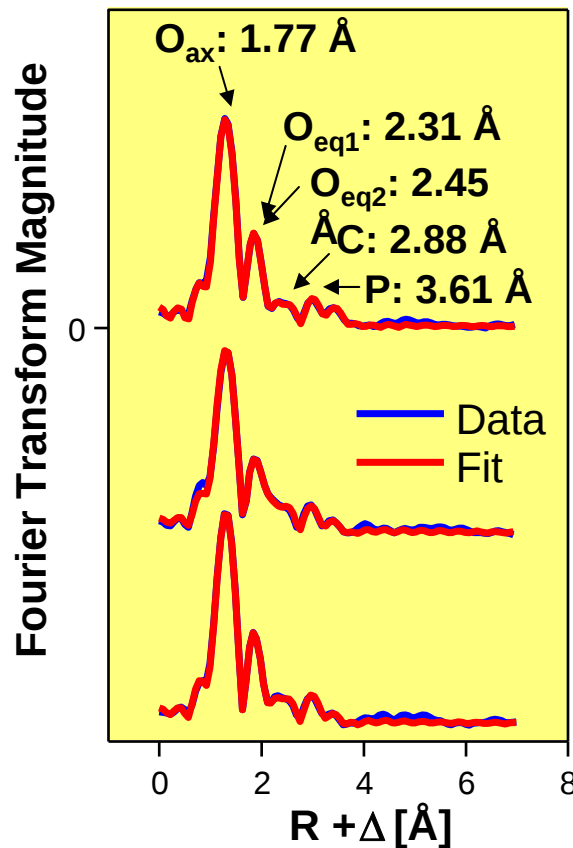
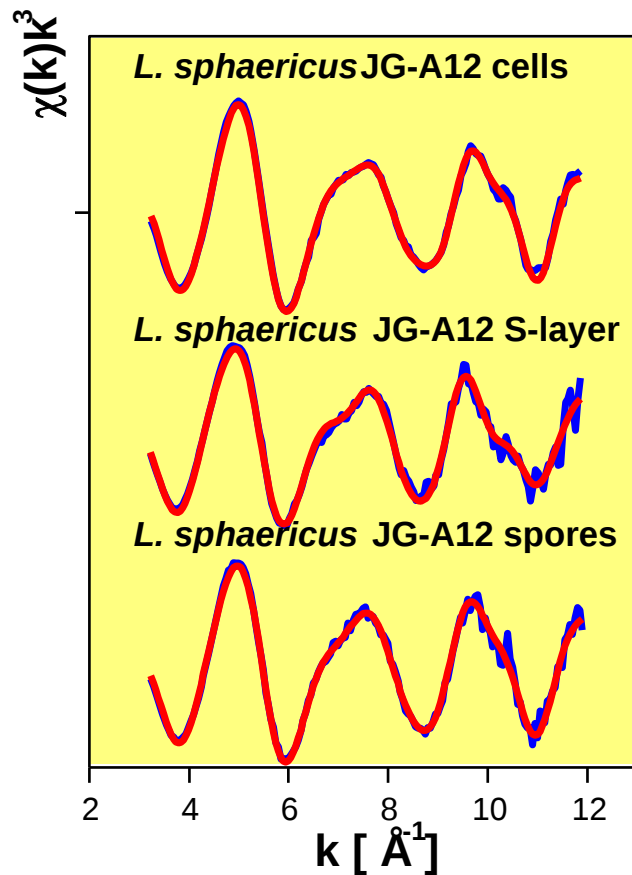


## ATR-FTIR



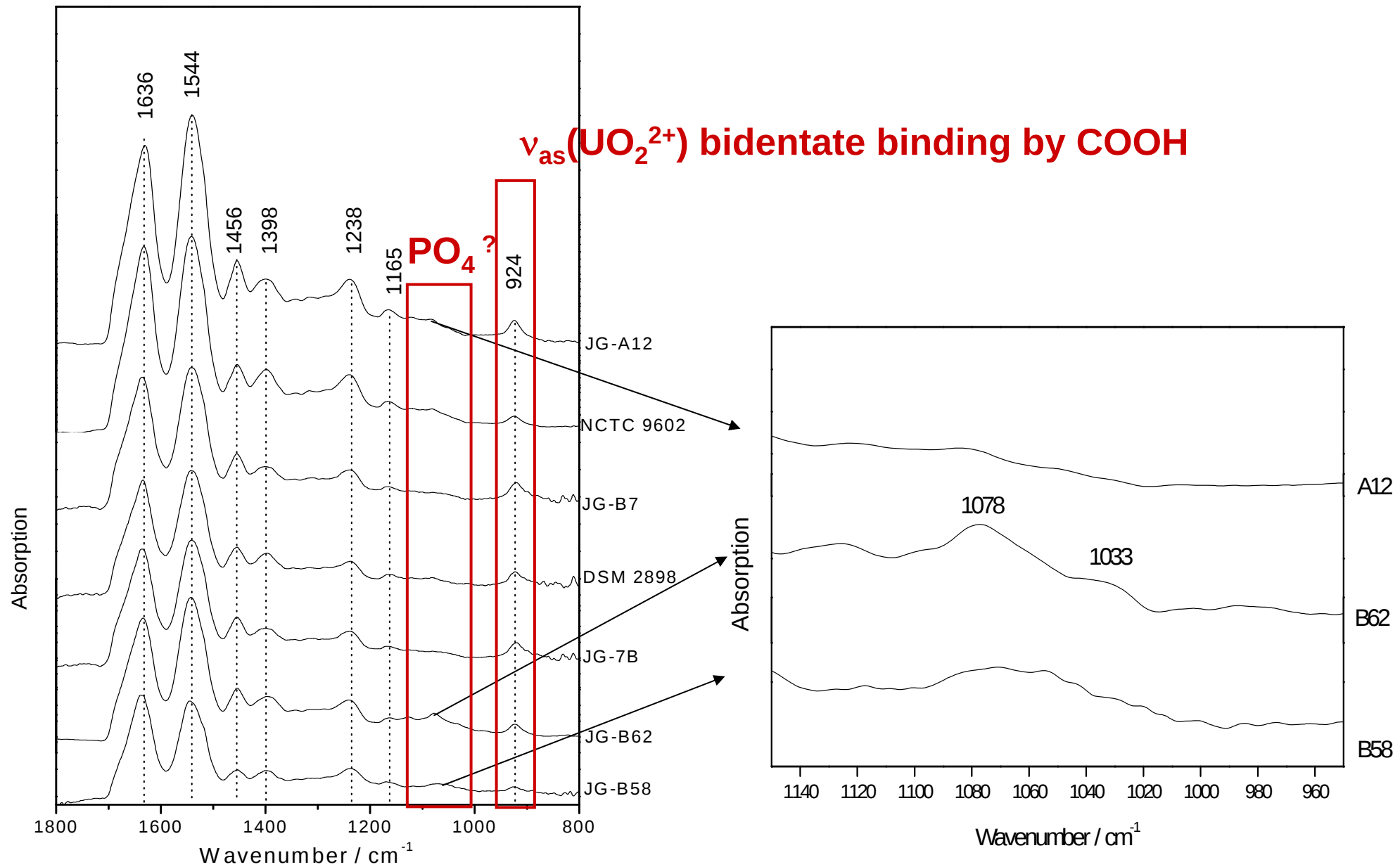
## TRLFS





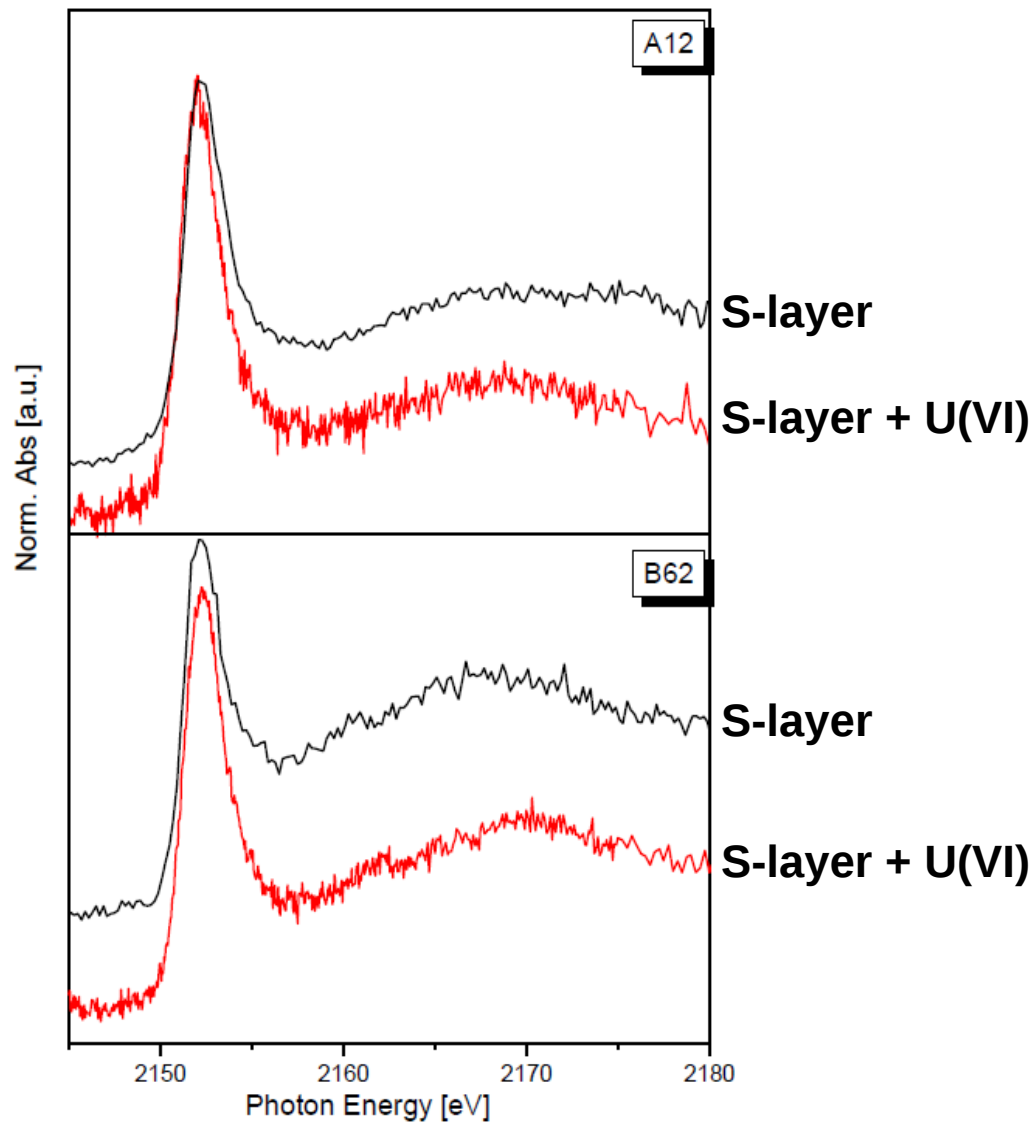
Merroun, M. et al. (2005) Appl. Environ. Microbiol. 71(9): 5532-5543

Raff, J. et al. (2004) In Water-Rock Interaction, Richard B. Wanty und Robert R. Seal II, A.A. Balkema Publishers, New York, p 97-701

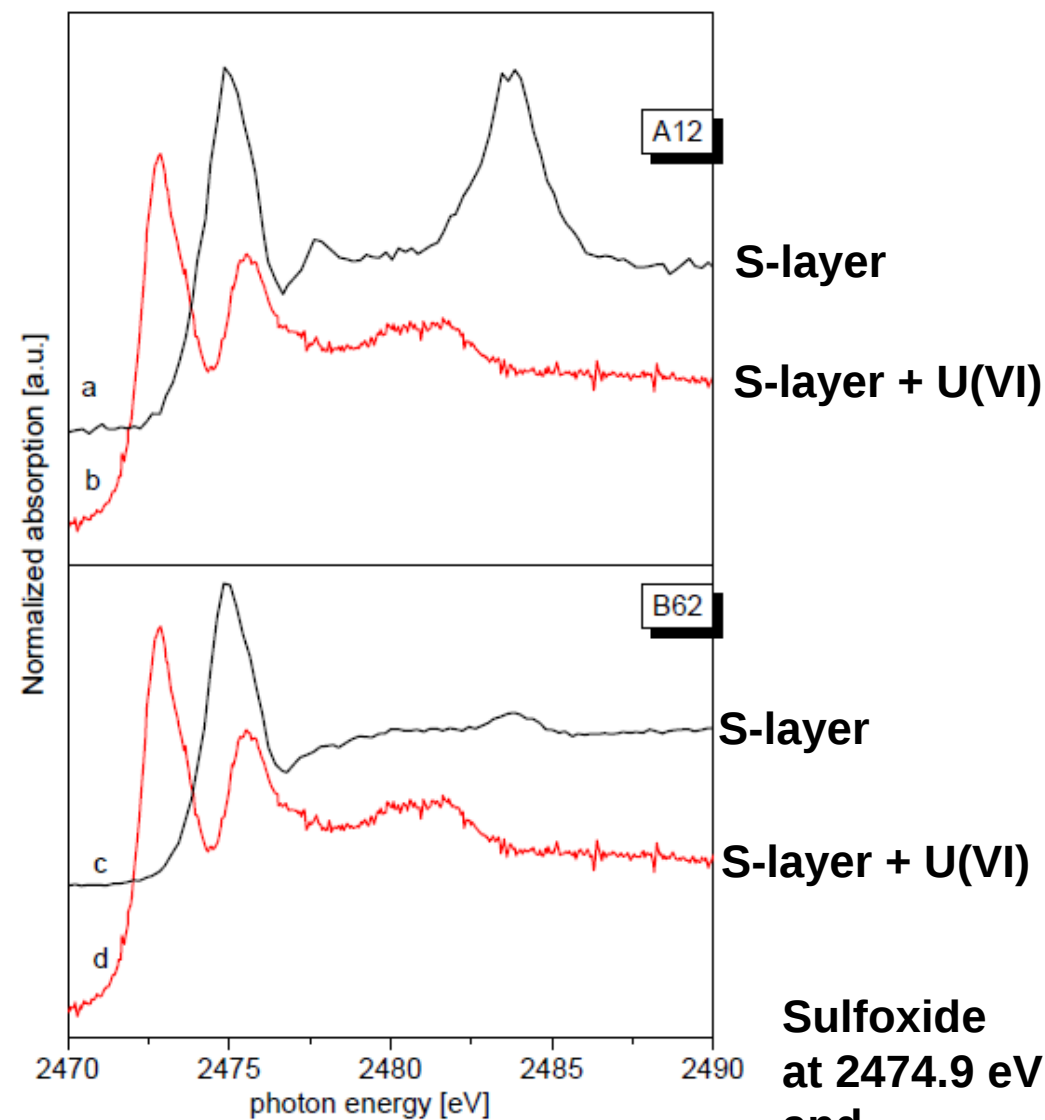




## P K-edge XANES

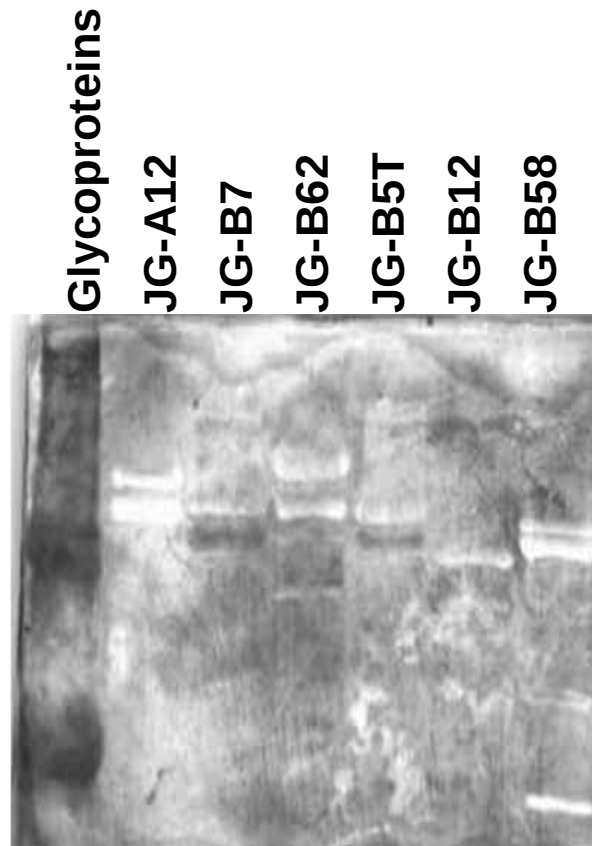


## S K-edge XANES



**Sulfoxide**  
at 2474.9 eV  
and  
**sulfate**  
at 2483.8 eV

→ No evidence, that Phosphate groups are not involved in the U(VI) binding, but sulfur-species interact also.



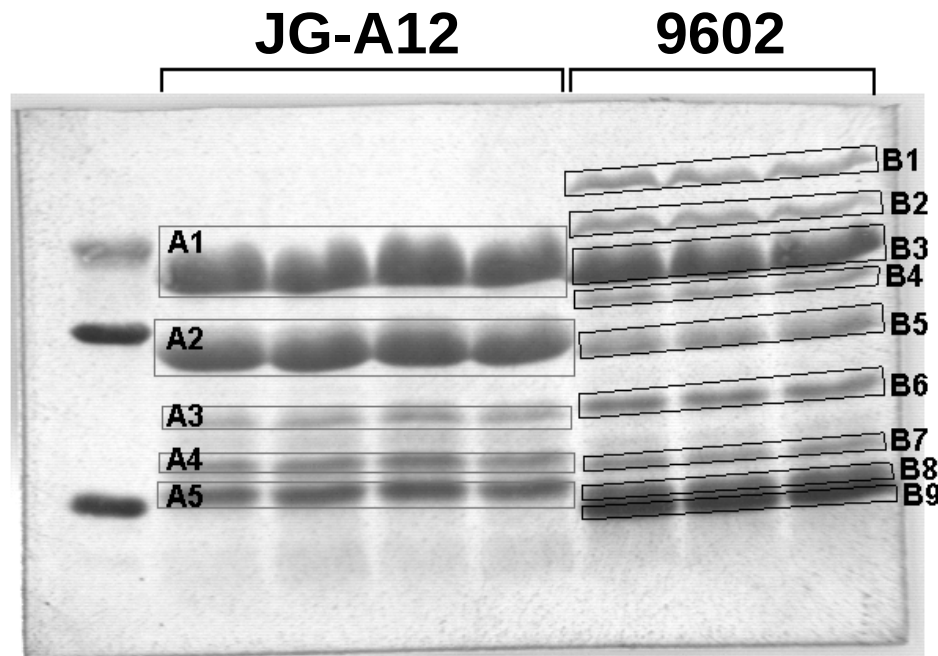
**Detection of glycosylated proteins by means of the “DIG glycan detection kit”. Dark bands indicate glycosylation.**

| S-layer | Absorption maxima | Sugar residue |
|---------|-------------------|---------------|
|         | (nm)              | (mol/mol)     |
| JG-A12  | -                 | 0,03          |
| JG-B5T  | 477               | 2,46          |
| JG-B7   | 482               | 1,93          |
| JG-B12  | 482               | 4,74          |
| JG-B58  | 477               | 2,33          |
| JG-B62  | 482-485           | 3,79          |

**Modified colorimetric determination of sugars after Dubois.**

Weinert, U. et al.(2010) Anal Biochem (in preparation)

## Proteolytic digestion of native S-layers from *L. sphaericus* JG-A12 and NCTC 9602 using the endoproteinase Glu-C (–Glu-P<sub>1</sub>′– and –Asp-P1– in Phosphate buffer)



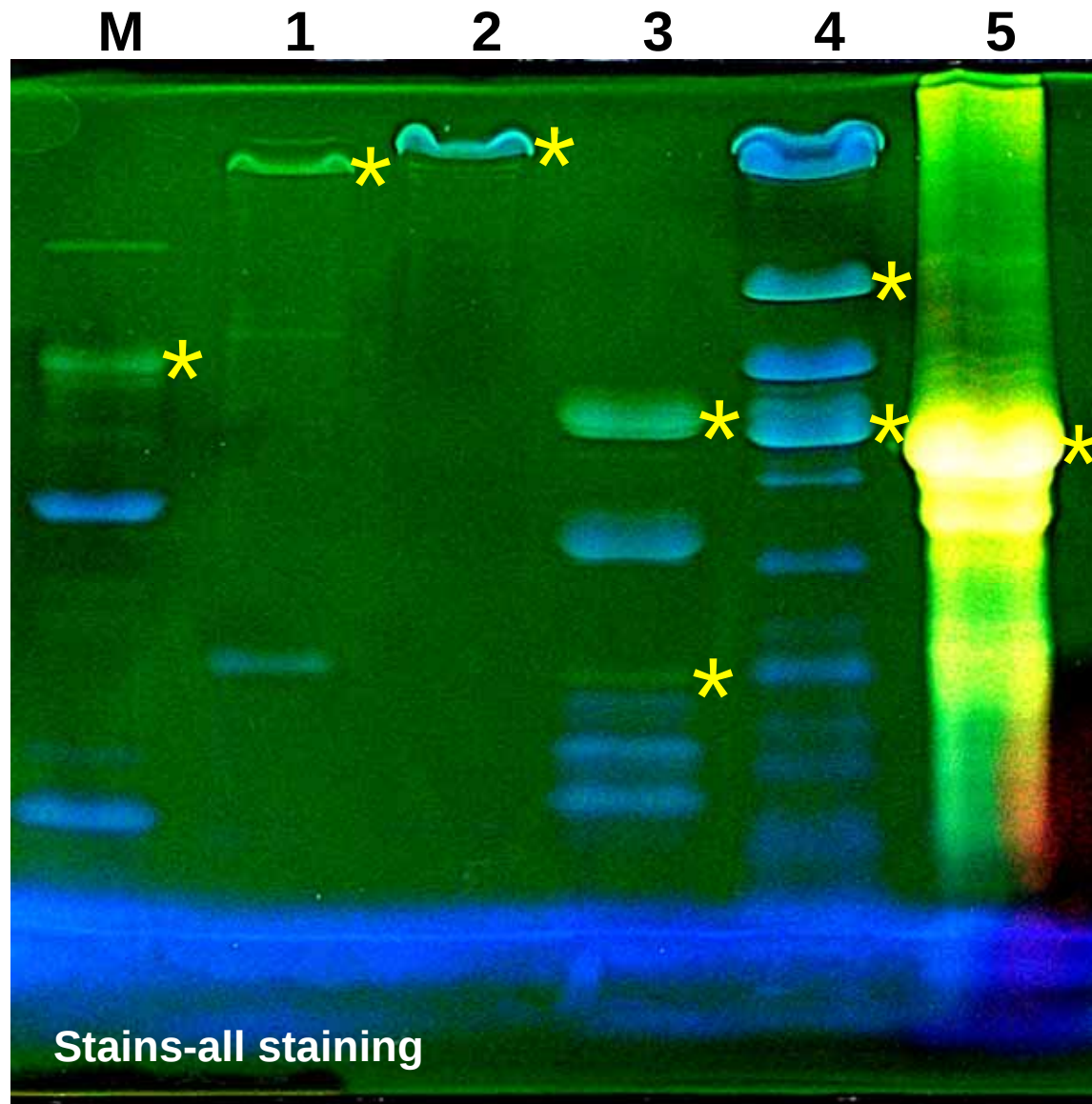
### JG-A12

```
MANQPKKYKKFVATAATATLVASAIVPVSAAGFSDVAGNDHEVAINALADAGIINGYADGSFKPNQITIN
RGQVVLLGRYLEAQGGQIPADWNSKQRFNDLPVTAEEELVKYAAALAKDAGVFNNGSNGNLNASQTMQRQQ
MAVVLVRAIKEIAGVDLVAEYKKANFVTEIGDLKAYSAEQRTAIVALEYAGITNVAHFNPNGSVTRGQF
ASFLYRTIENVVNNPEAGVAAVKAINNTTVEVTFDEEVDNVQALNFLISDLEVNAAVKLTNKKVVVLT
AAQTADKEYTVSLGEEKIGTFKGIAAVVP TKVDLVEKSVQGGKLGQQVTLKAQVTVAEQGTAKAGIPVTFI
PGSANGVKTPVTVAEATDEEGIATYSYTRYAATNDTVTVYANGDRSKFSTGYVFWAVDQTLLEIETVTDG
IINNGANKTYKVYKHPETGKPVSGKVLNVSVKENIDVTVDKLQNVTVNGIAVQTSNNTTAAQITTD
KGEATFTTSGSNAEVPVVFEEATPVQVNTNGAVTTTSYTKYTADILQASAAKVTFGAVQAAATLEVTR
DGGEVAATGVENGRKYNLVKDKDKLAANE TVNVAFNEDIDGVI STVTSAEFVKVENDKQVRYEGKKIT
VKTNKSGEASFVISSEAVNTIATPIAWIDINNQS AKDANLDKGEPSAVAPISYFQAEYLDGSKLVSYKDT
TETDKFTGSETATFKVQLTNQSGKVVKNSGYTTPNVSYTVYNTGANNVKVDGVEIAPNRVHTVVAKDGV
NVTTVDNKSSSVKVLATGVAKQTNGNKEFAFTSKEATATFTATNEV NPY GAVKHYNTDKKTIITFDNKD
AIKYAGVKGKTYKYFGLGSTPIANADAFIATLSGGAGNQVTVTYKEEDDVVSFYIISVVNGGIGNPTTD
PALANGVTGTIAFTNTSFNASANQITTVKDDADLVNATVADTTTVDTSAKITFNITLTETGVNIGSET
TITSAQLALLKDGVI TATYNDAKNANNVAATATATATLNTTVGAVNFAAKATTEYSEGVGTAAKVEGST
VLTISKVDGDLVLVDGVQFTTVVTAIDPGTSATASLTAVVAEINAAKVGFTADVATVDADKIVLTSP
TKGTSSSVEVKDLGTTTGLGLTKAPEVKGTAGAAVATQWKFTVTTTPAVGETITVKVGTKESHKVVAGN
DLAAIATALNTAIGADYNIALVTGSNVITVTQATPAKTTDELSTVTVK
```

### 9602

```
MANQPKKYKKFVATAATATLVASAIVPVSAAGFSDVAGNDHEVAINALADAGIINGYADGSFKPNQITIN
RGQVVLLGRYLEAQGGQIPADWNSKQRFNDLPVTAEEELVKYAAALAKDAGVFNNGSNGNLNASQTMQRQQ
MAVVLVRAIKEIAGVDLVAEYKKANFVTEIGDLKAYSAEQRTAIVALEYAGITNVAHFNPNGSVTRGQF
ASFLYRTIENVVNNPEAGVAAVKAINNTTVEVTFDEEVDNVQALNFLISDLEVNAAVKLTNKKVVVLT
AAQTADKEYTVSLGEEKIGTFKGIAAVVP TKVDLVEKSVQGGKLGQQVTLKAQVTVAEQGTAKAGIPVTFI
PGSANGVKSPVTVAEATDEEGIATYSYTRYAATNDTVTVYANGDRSKFSTGYVFWAVDQTLLEIETVTDG
IINNGANKTYKVYKDPETGKPVSGKVLNVSVKENIDVTVDKLQNVTVNGIAVQTSNNTTAAQITTD
KGEATFTTSGSNAEVPVVFEEATPVQVNTNGAVTTTSYTKYTADILQASAAKVTFGAVQAAATLEVTR
DGGEVAATGVENGRKYNLVKDKDKLAANE TVNVAFNEDIDGVI STTDAKFVKLDNEDNVQVWDTTVVDKDAKKITV
KTDKSGEASFIIIGNTKENTYTIPIAWIDVNYQSGKVGTLQDQGEPSATAPISYFQSPITDGLKLVANEDKG
KKPSDFEDKDAIFKVVVLNVQSGKEMKNHNYKTEKVSYTVNNTGSNEAVVEYTYNGKVESETLAPNRTSV
VADNGTITVPKNGKTSVKVLATGVAKEDKTNKGEYAF TAKEATAKFTATKEISNPYIGAIKYVYDIDDKT
ITFDGKDAIKYAGESGKKYKYSNGIEVATEKAFIDLLANASGKVTVTYKVEDDTKSFHINVGTDVANK
PVEKKDDKPNAPVAKQVADQVIA SNGSKLFTANDVASDADKDLKLYTATYTTNSTVATATTDANQL
GFSVEPKGEGSTTVLVVVDGKDNINVSKVNVSTAYTSVNSGNPVKDFAPAVPTAATAKLPEFTSVA
KAGTIKVGQFDIPLTLGSTQAQVLETLNFI TDYKINASAKLVDAKIVISSTETGNAATLTVEGDSTLEL
VAGTVLGTVSTPGSVGTATPAKYAVKVLNGVNGTKVDFKFTIGAKTVTVTVDATAGAKAVSDVVDTLA
AATLDGYSITKNGVDIELTQTTPATTTDKLVVETKKAN
```

### Proteolytic digestions of native S-layer endoproteinase Glu-C



#### Samples

M: Marker

1: S-layer JG-A12

2: S-layer NCTC 9602

3: Glu-C digested JG-A12 SL

4: Glu-C digested 9602 SL

5: Phosvitin

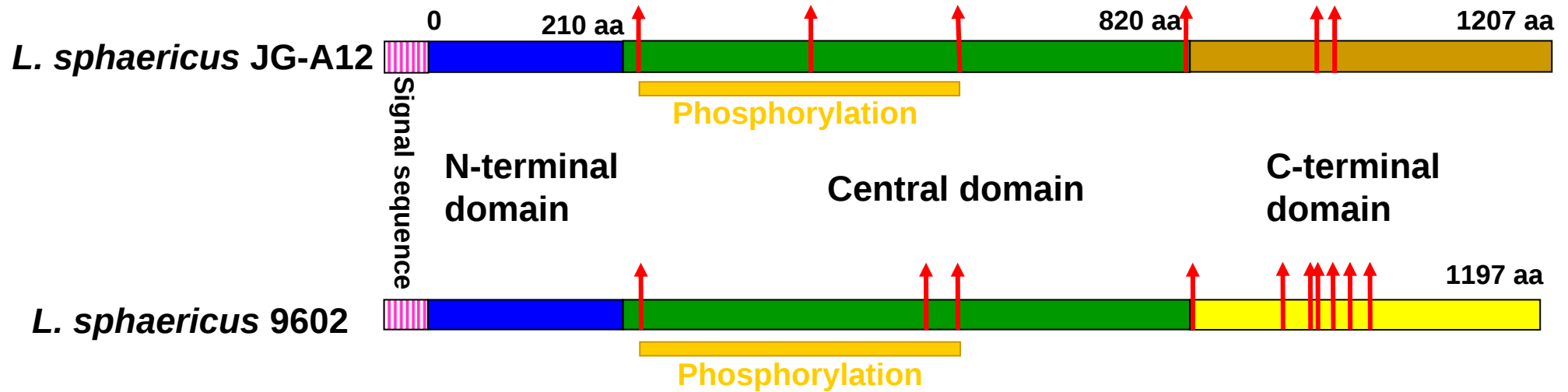
#### P-positive references in lane...

M: Ovalbumine 1,73 P:1 (0.12 %)

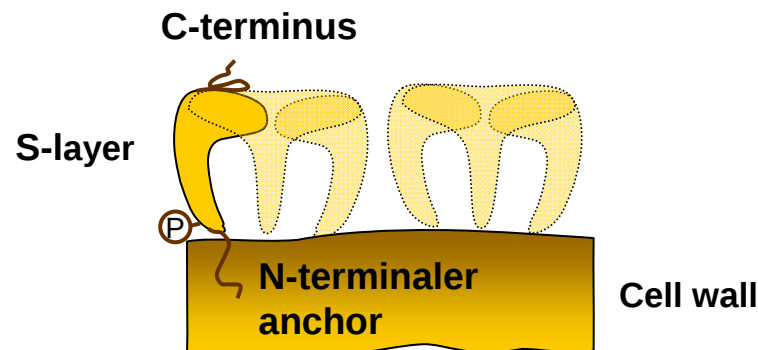
5: Phosvitin 100 P:1 (10 %)

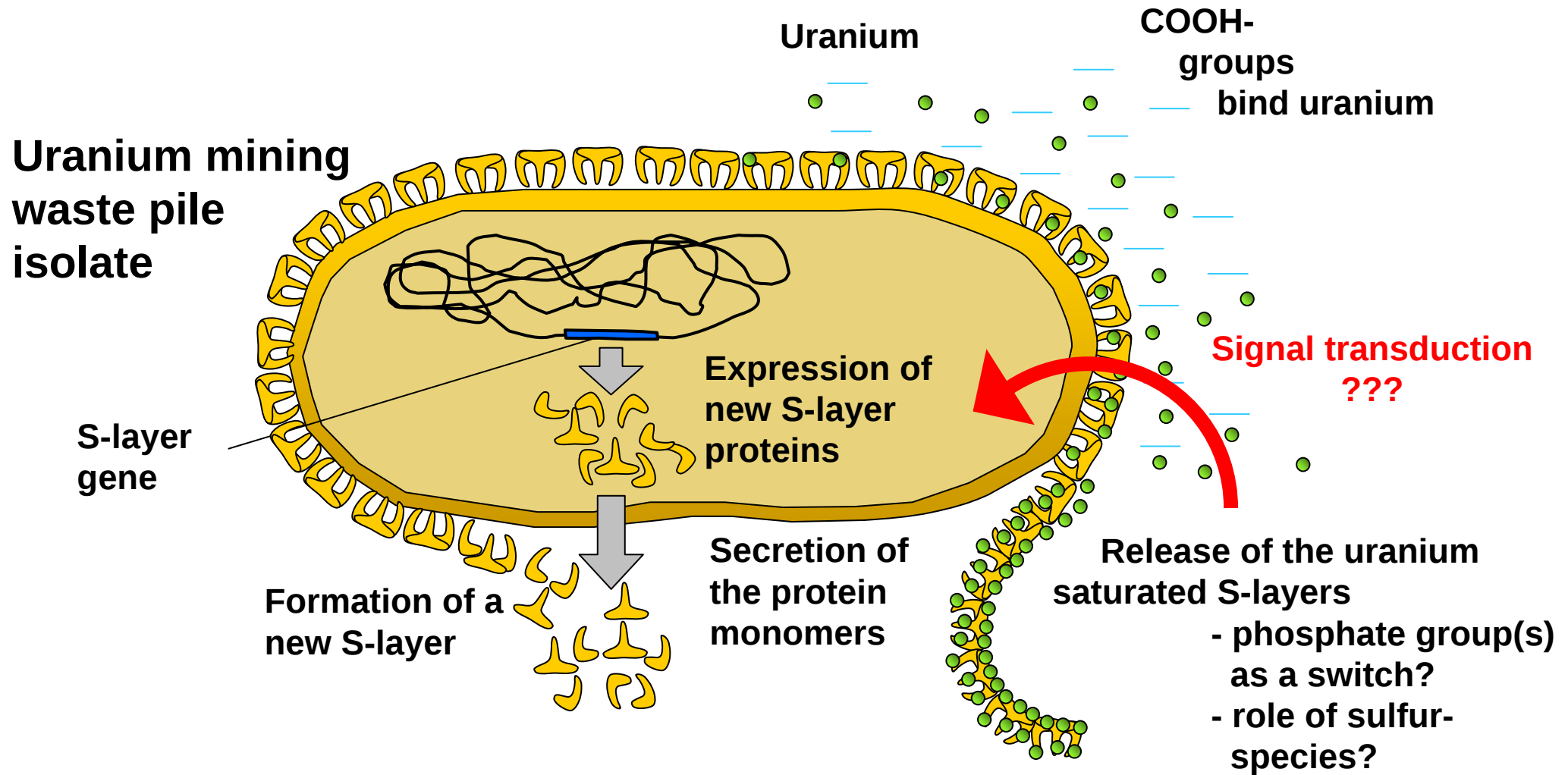
P-positive bands are indicated by an asterix

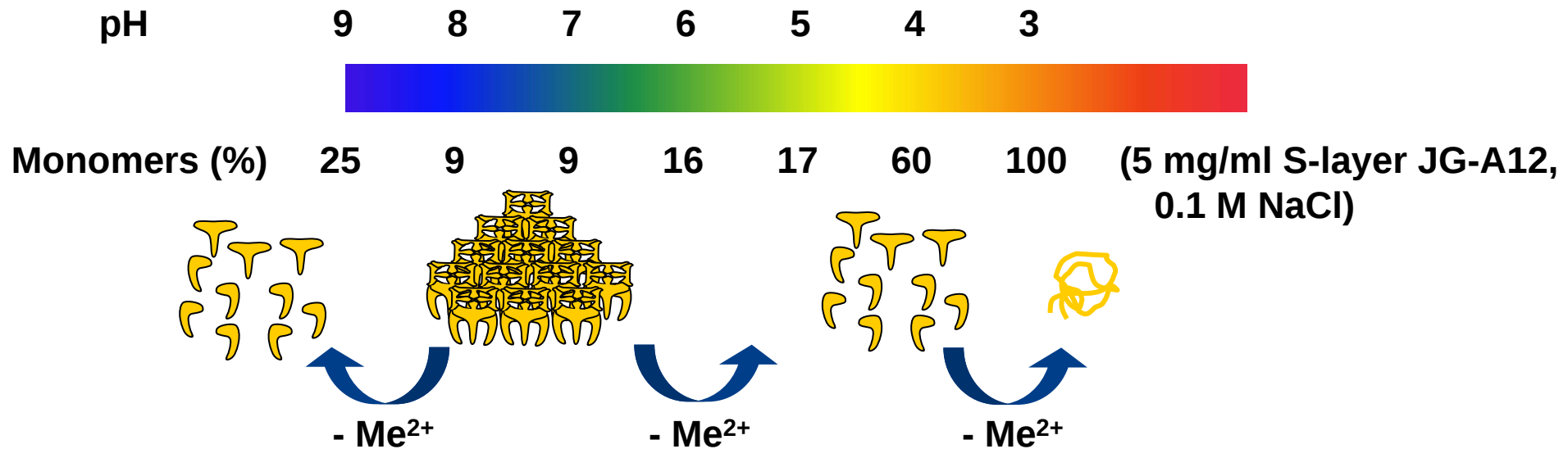
### N-terminal sequencing of the fragments



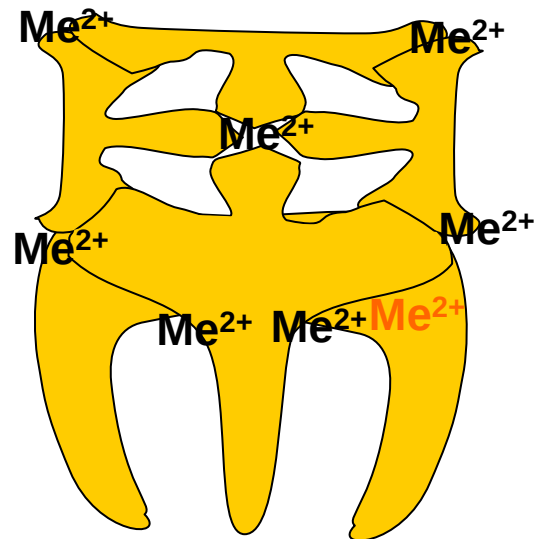
Glu-C cleaving sites in the native protein;  
theoretical 125 (JG-A12) and 141 (9602)  
Possible sites.





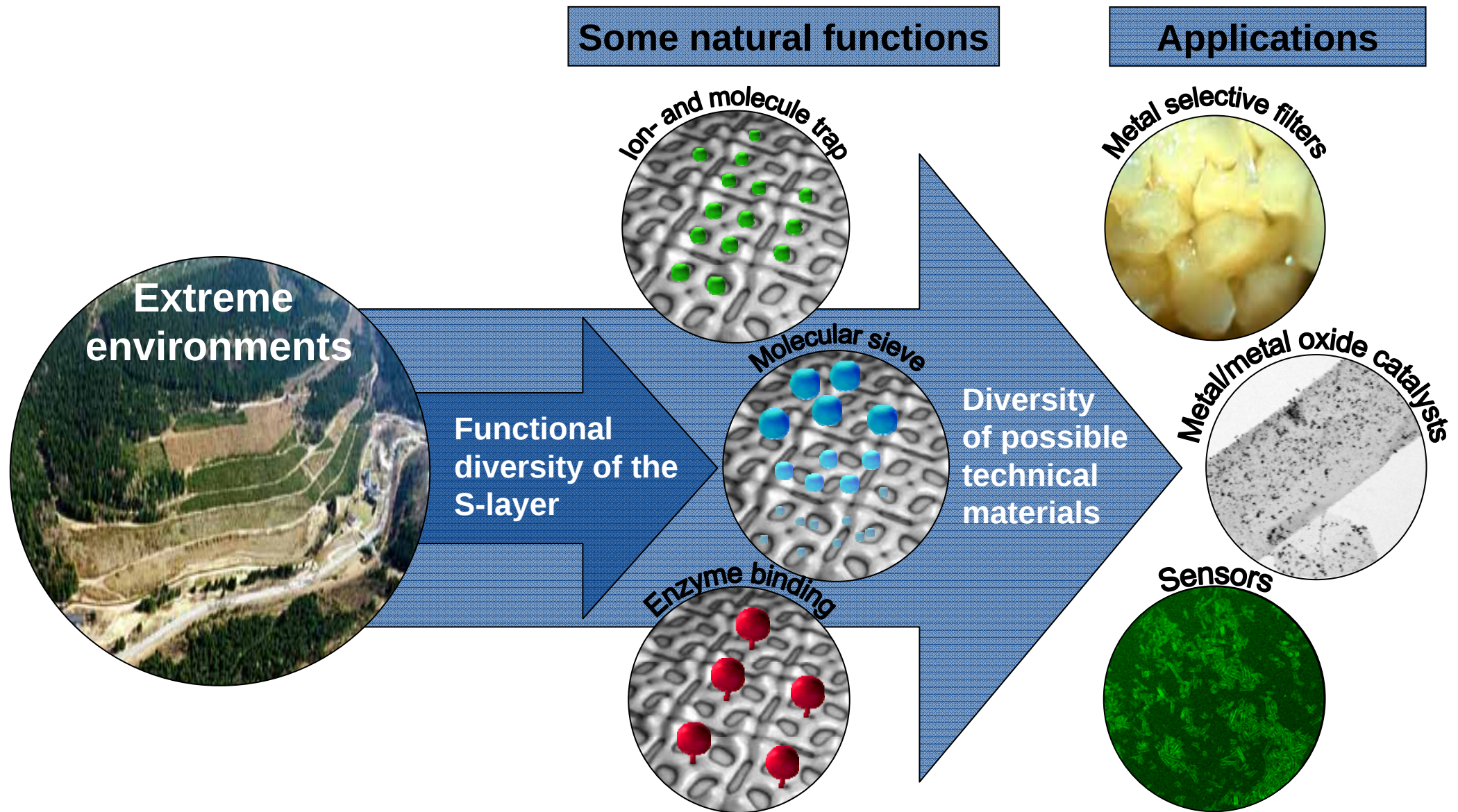


Hypothesis:

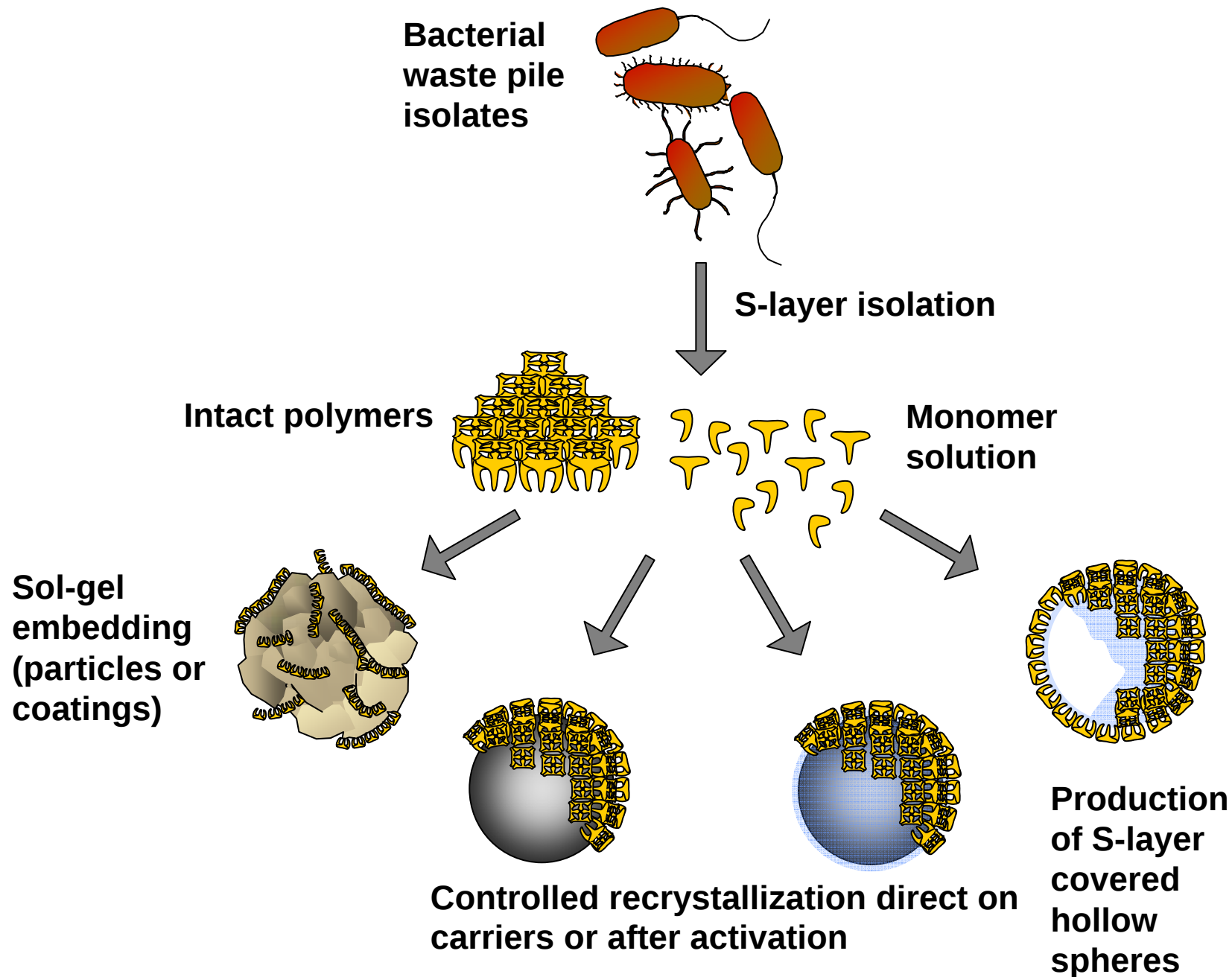


# Application oriented research

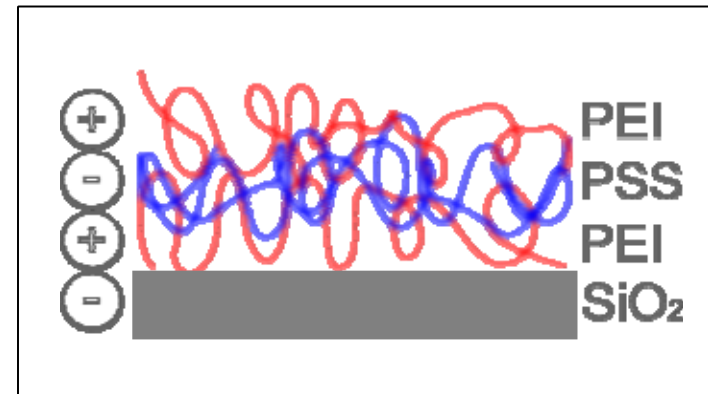
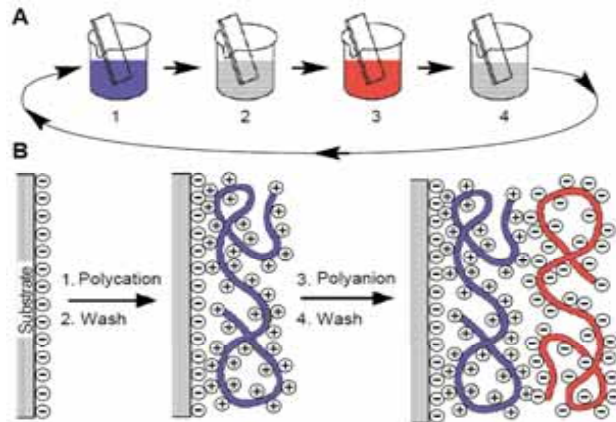




Pollmann, K. et al. (2006) Biotechnology Advances 24 (1), 58-68

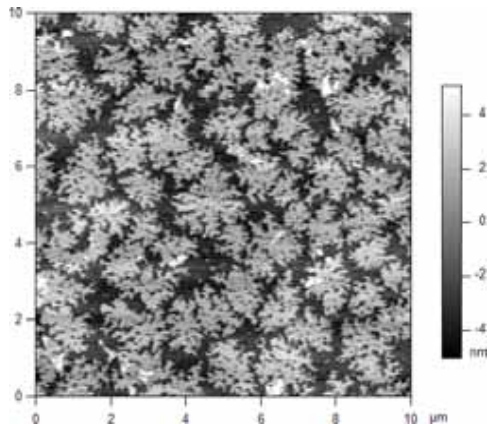


- To improve the surface properties for S-layer recrystallization a polyelectrolyte interface can be introduced
- Simple LbL-coating
- Layer composition influences the stability of the protein coating



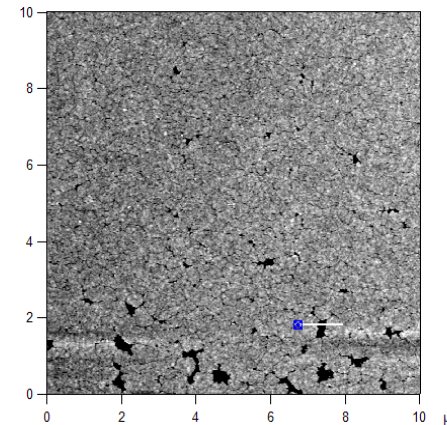
Modified after Decher, G. SCIENCE 277(5330), 1232-1237

## Without polyelectrolyte support



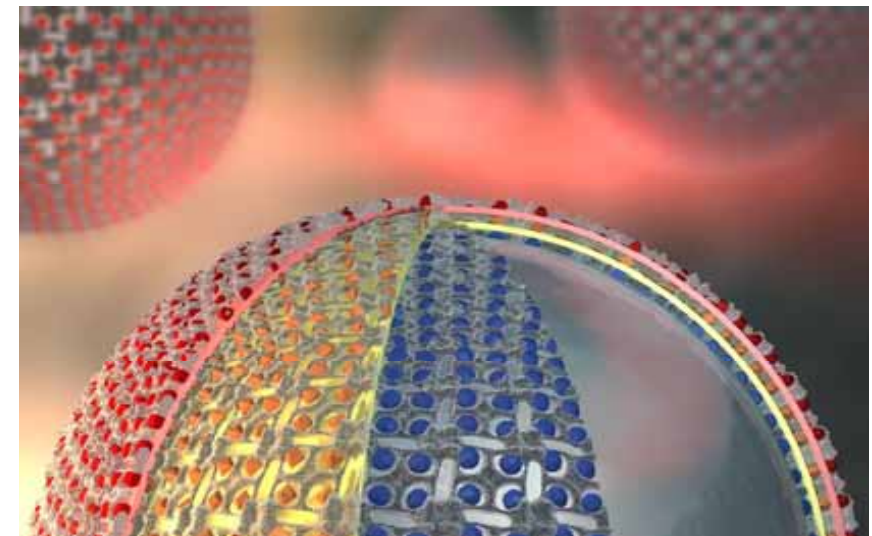
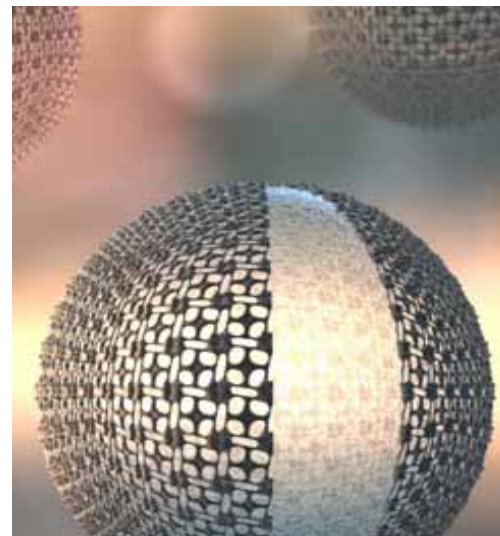
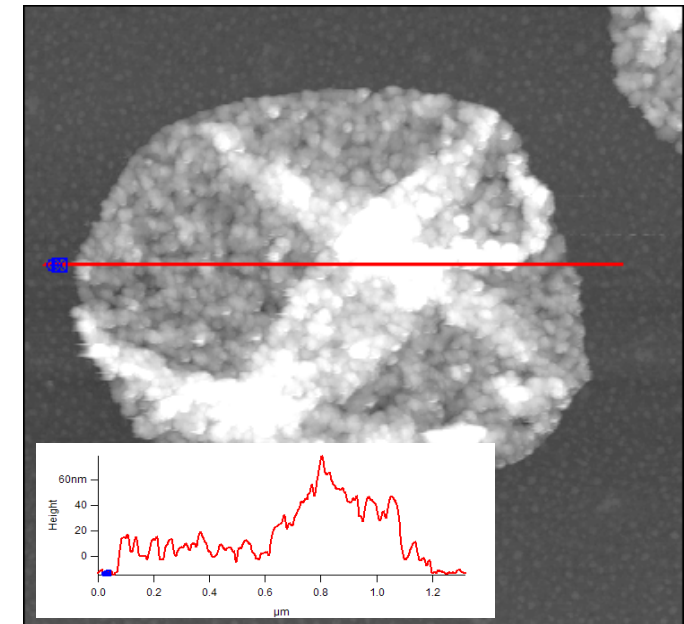
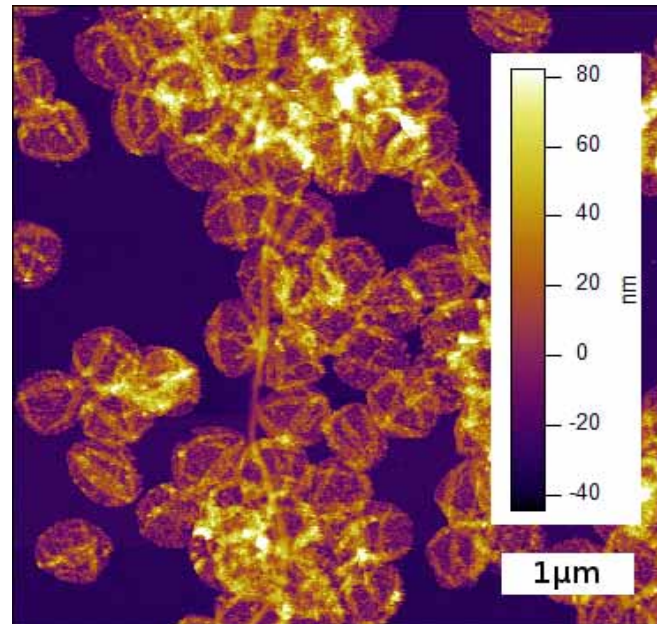
- Recrystallization on  $\text{SiO}_2$
- 57 % coverage
- 12 h incubation

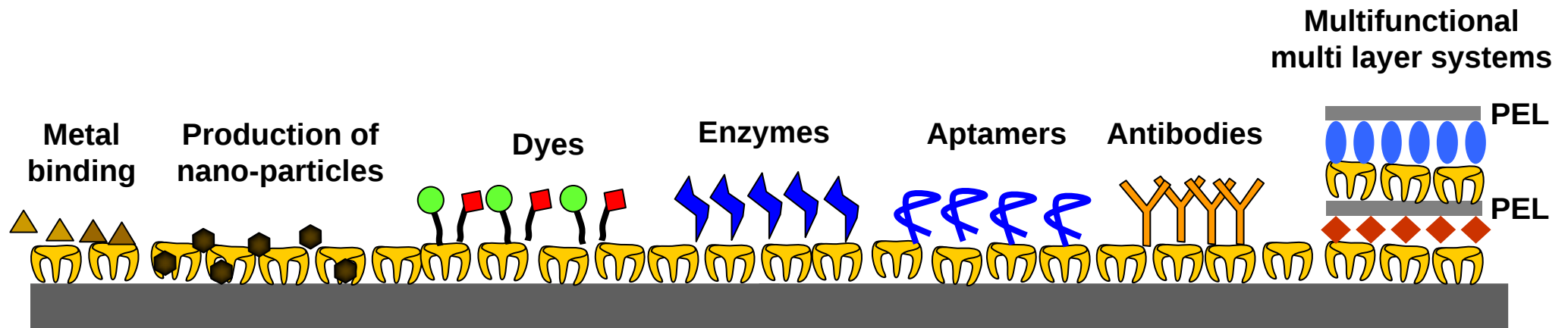
## with polyelectrolyte support



- Recrystallization on PE-coated Si
- >90 % coverage
- < 30 min incubation
- Monolayer

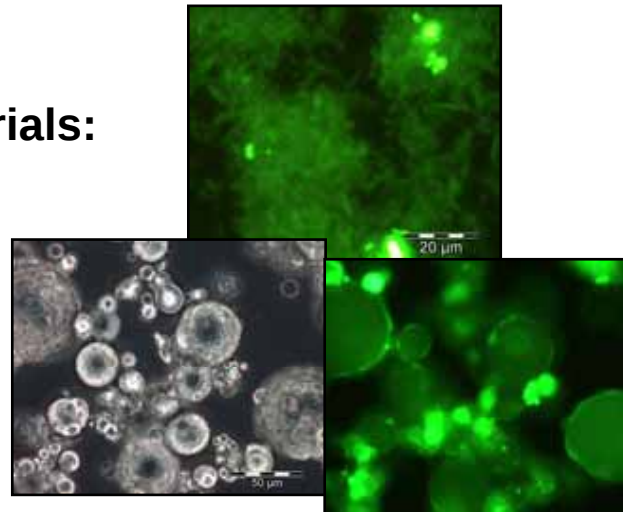
- Coating of spheric templates ( $\text{CaCO}_3$ , CaP) with multiple layers (4-10-...) of polyelectrolytes (PSS, PAH)
- Dissolution of the core material by HCl
- Coating of the spheres with S-layer(s) via recrystallisation and LbL-techniques
- Mono- or multifunctional transparent spheres
  - Magnetic “beads”
  - Combination of binding molecules and catalysts
  - ...

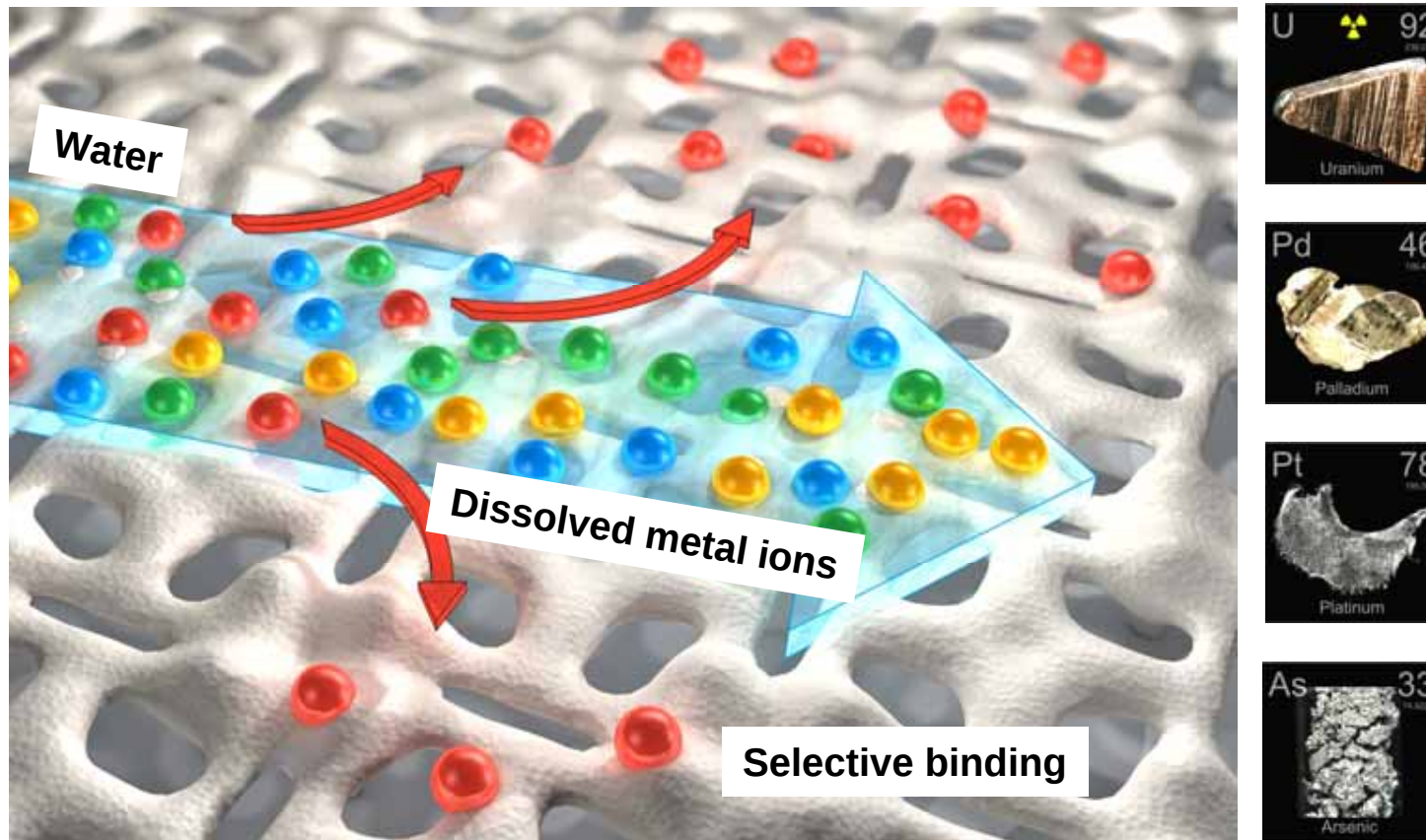




**Possible carrier materials:**

Glas  
Ceramics  
Metal  
Plastics



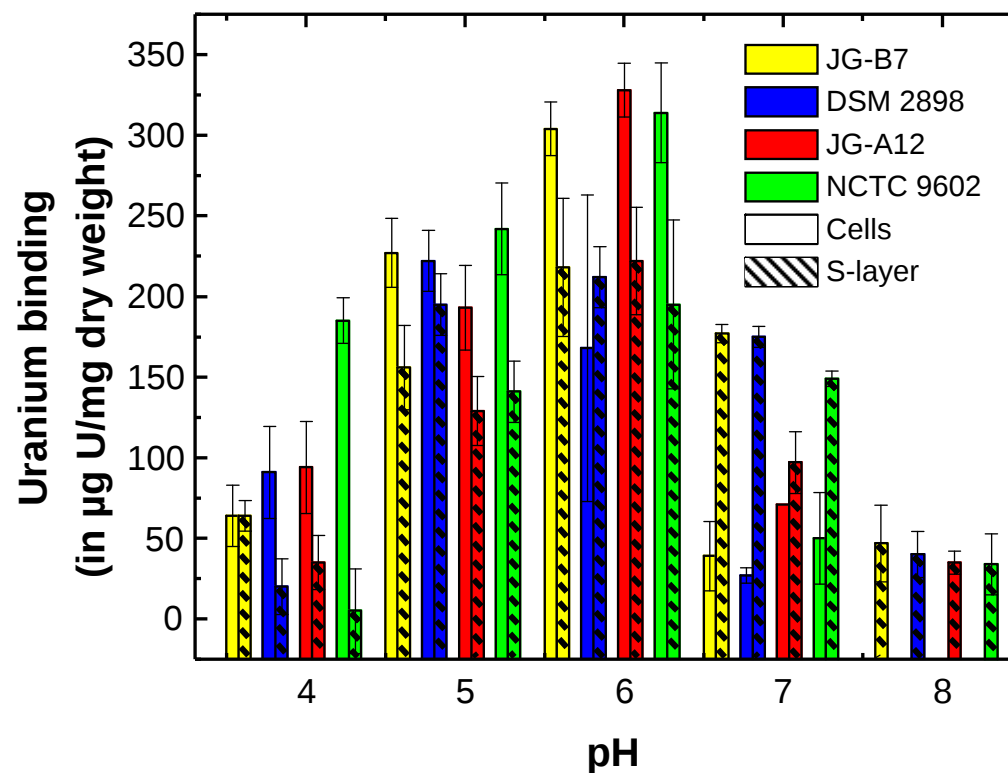
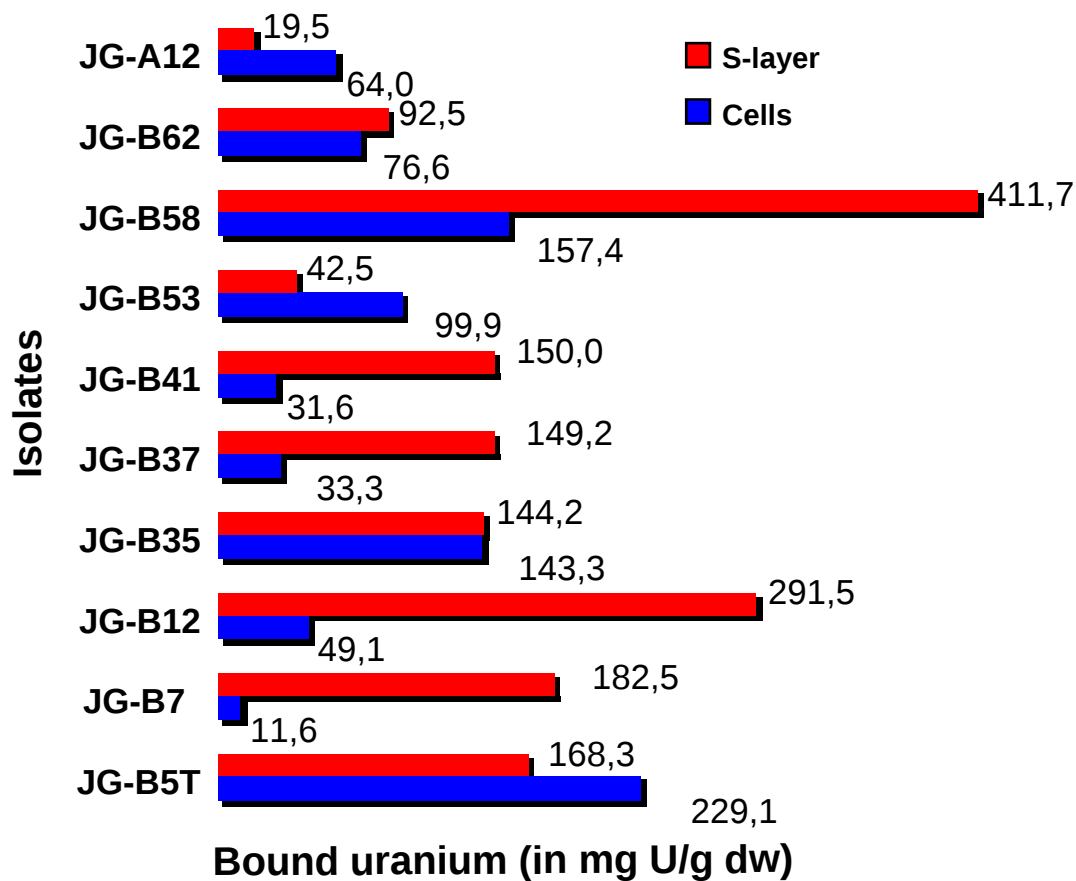


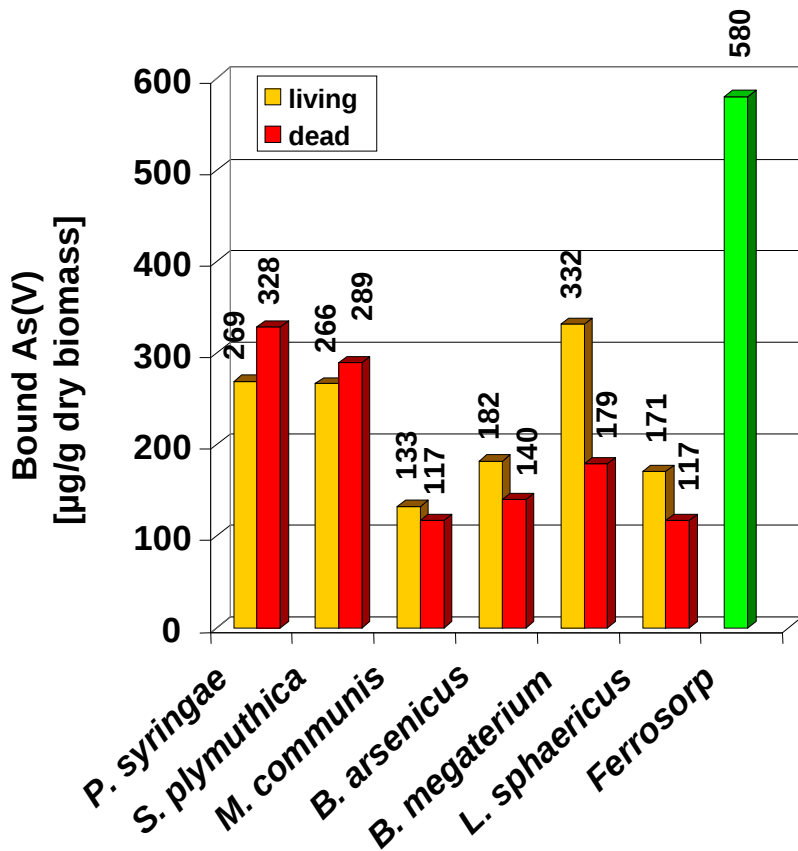
**Advantages compared to previous approaches:**

- Binding properties
- Environmental-friendly disposal
- Selectivity ?

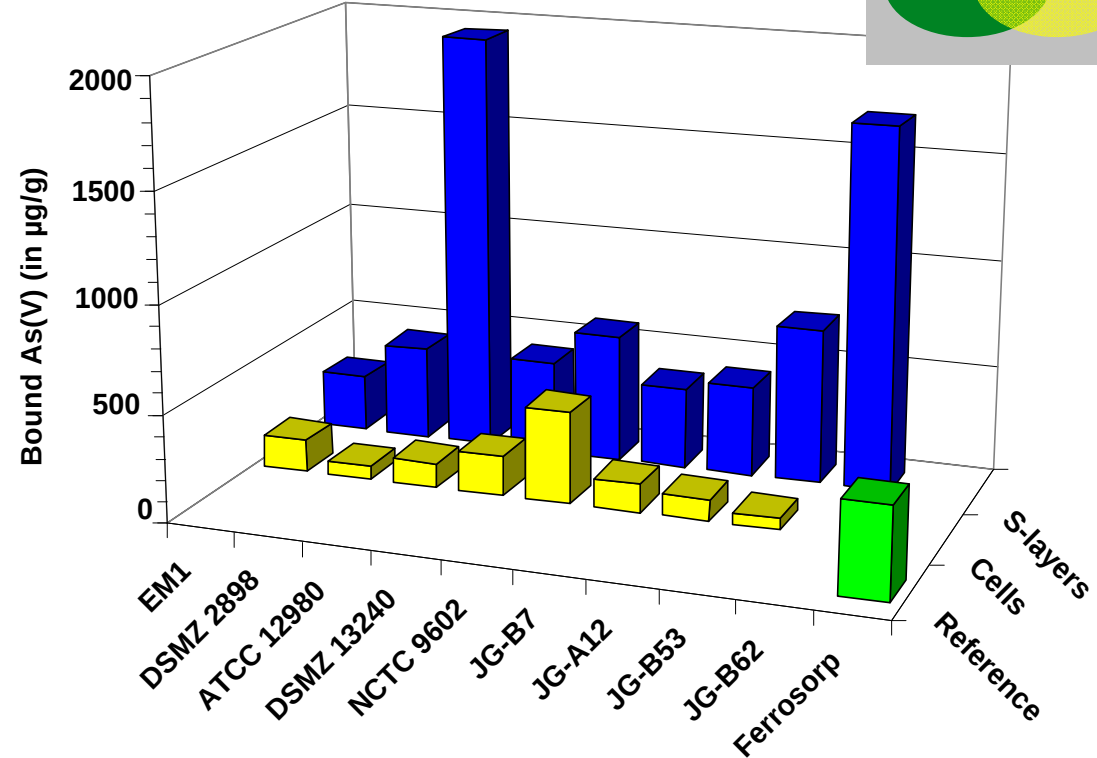
<http://www.periodictable.com>

Raff, J. et al. (2003) Chem Mater 15(1): 240-244





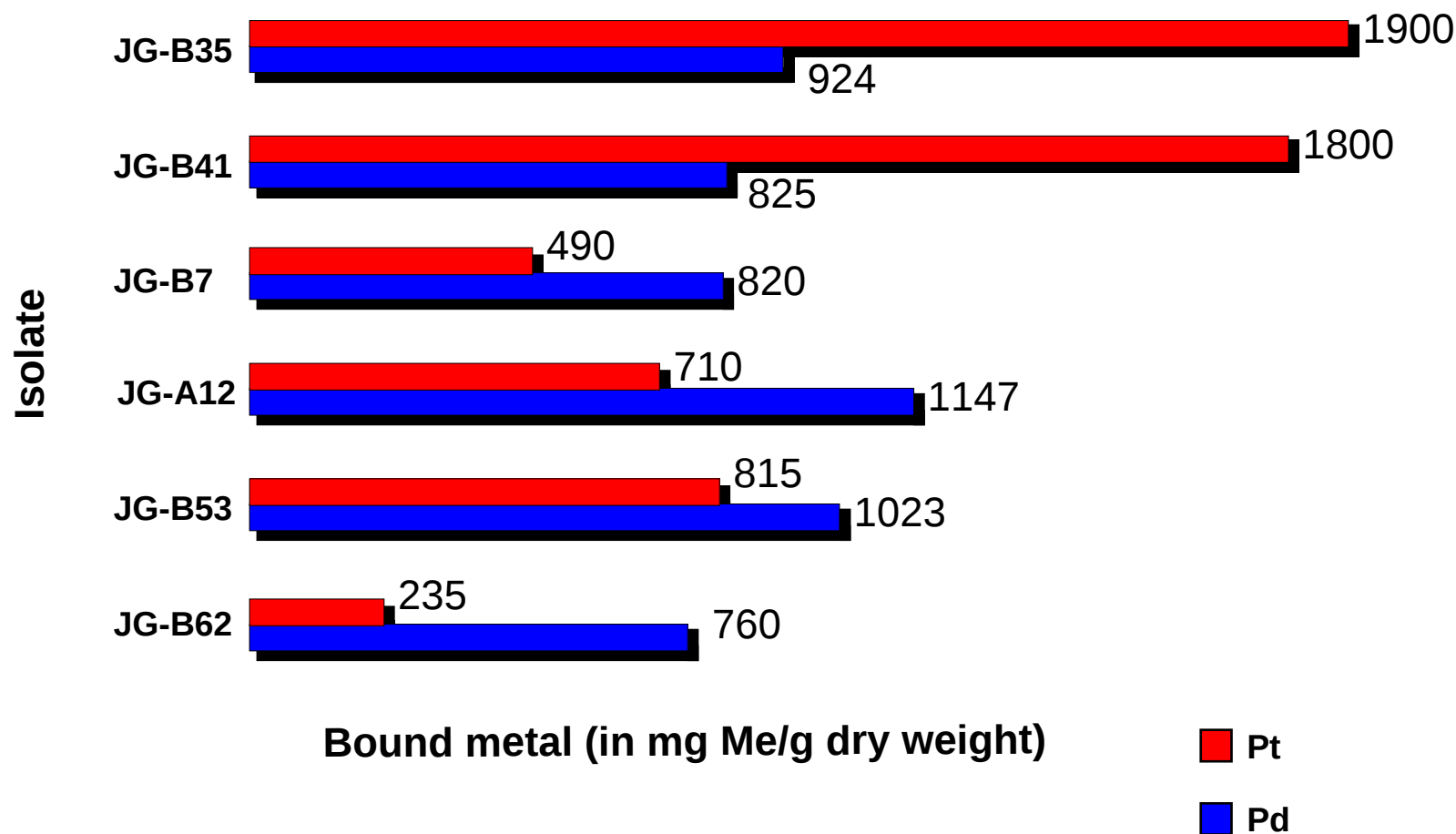
As(V) binding by living and dead biomass, using a As(V) solution with  $[\text{As}] = 10 \text{ mg/l}$



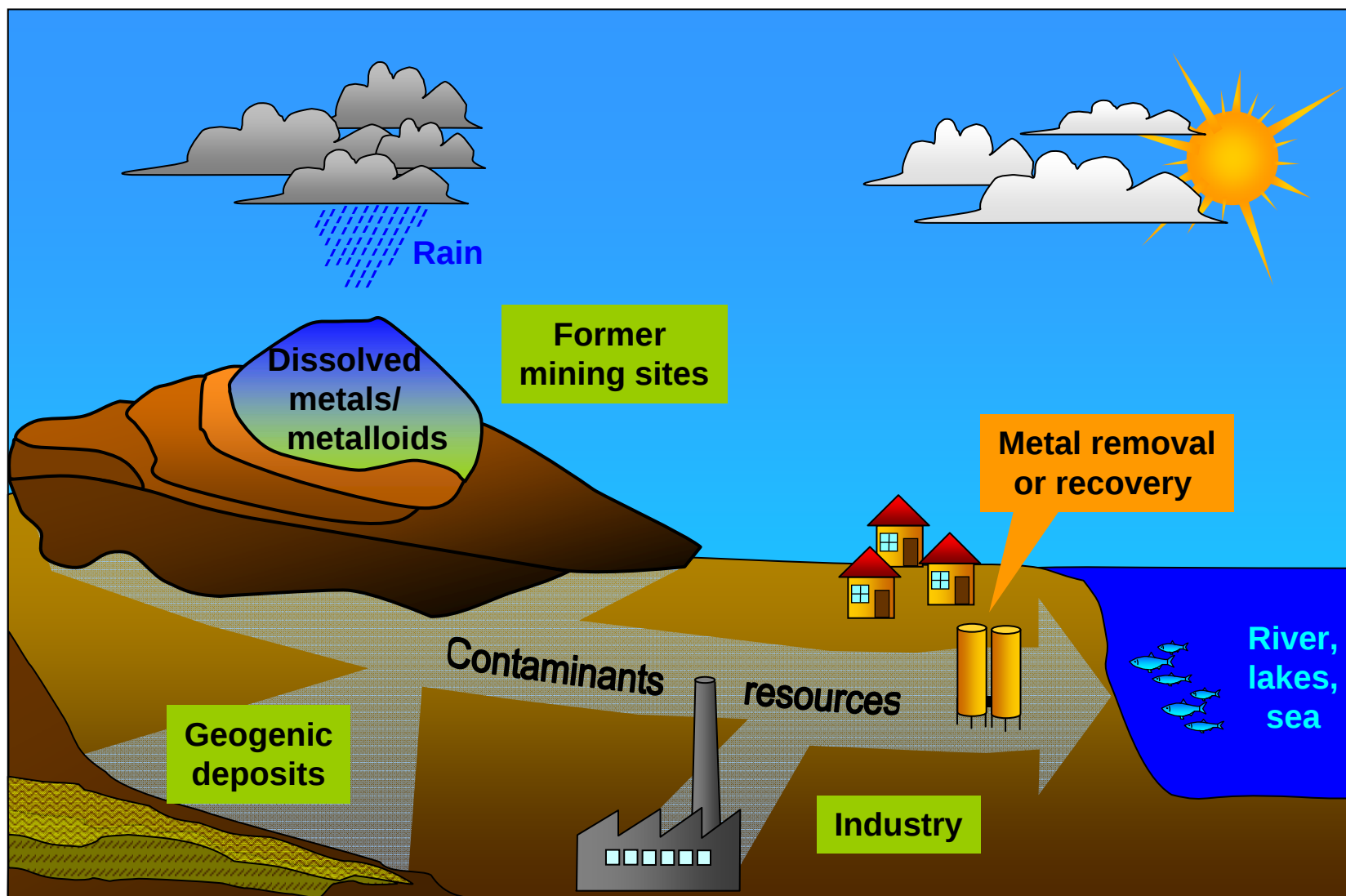
As(V) binding by cells and S-layers of reference strains and uranium mining waste pile isolates, using a As(V) solution with  $[\text{As}] = 10 \text{ mg/l}$

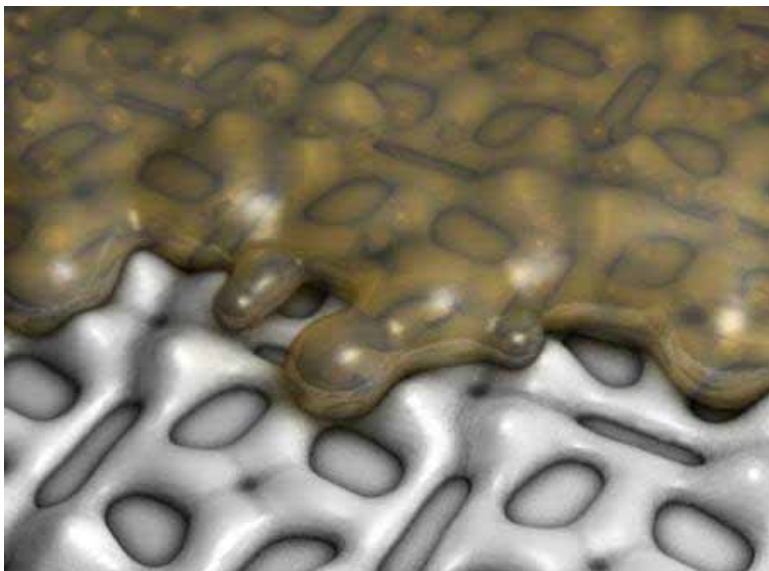
Reference material: Ferrosorp (granulated ferric hydroxide)

Matys, S. et al. (2010) FZD-Report 530

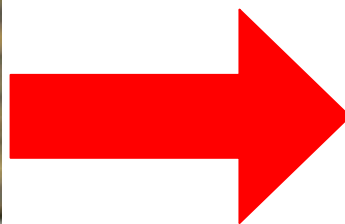


Fahmy, K. et al. (2006) Biophysical Journal 91 (3), 996-1007

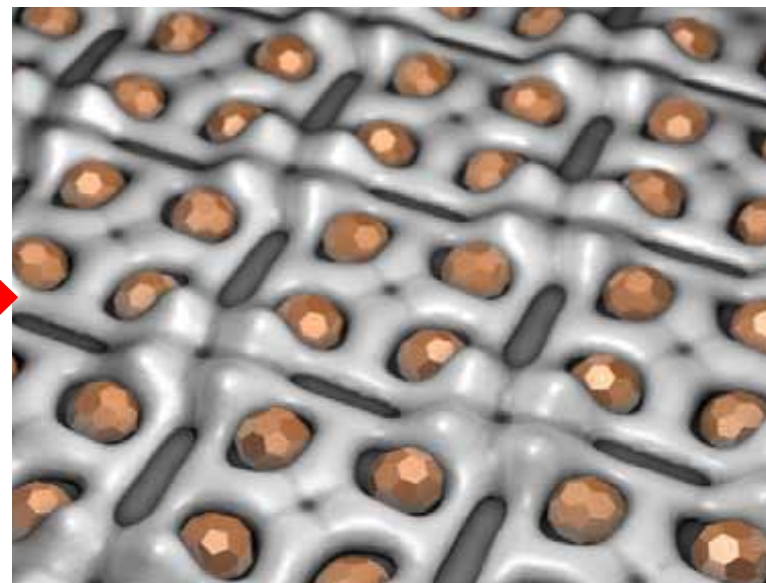




Incubation with a metal salt solution

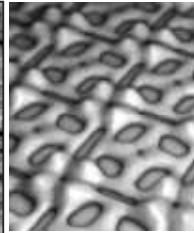
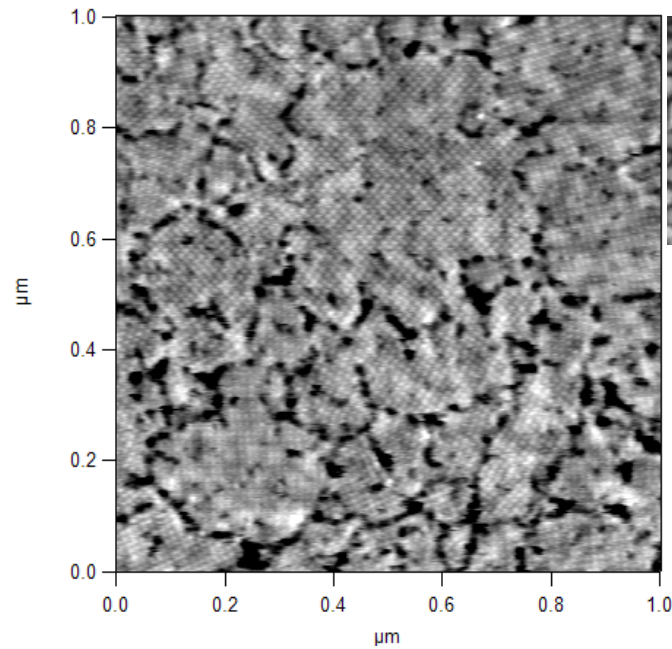


+ Addition of  
reducing /  
precipitation  
agents

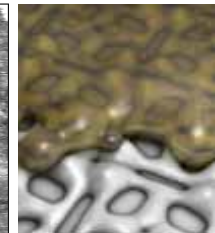
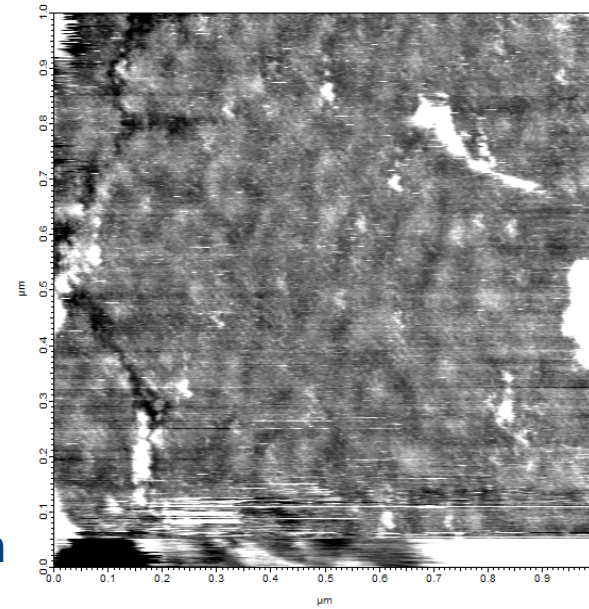


Defined catalytic active nanoparticles e.g.

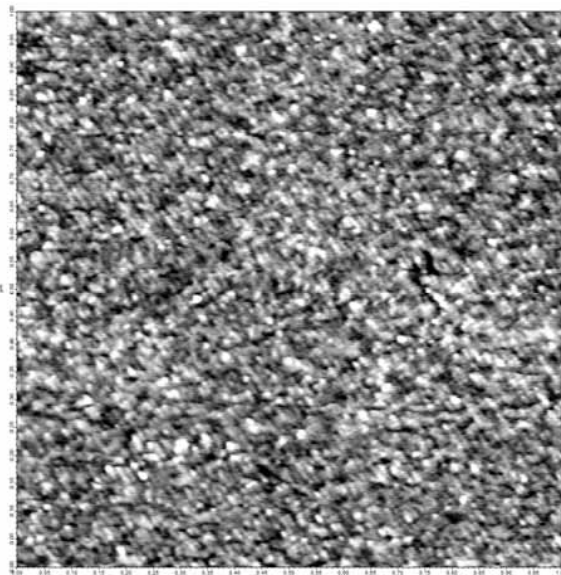
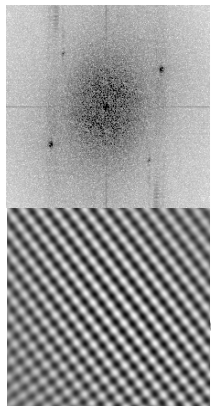
Pt, Pd, Fe/Pd, Fe/Pt  
ZnO, TiO<sub>2</sub>  
with sizes of 1-26 nm



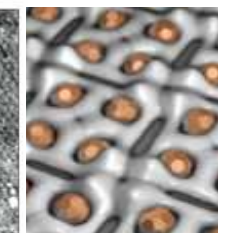
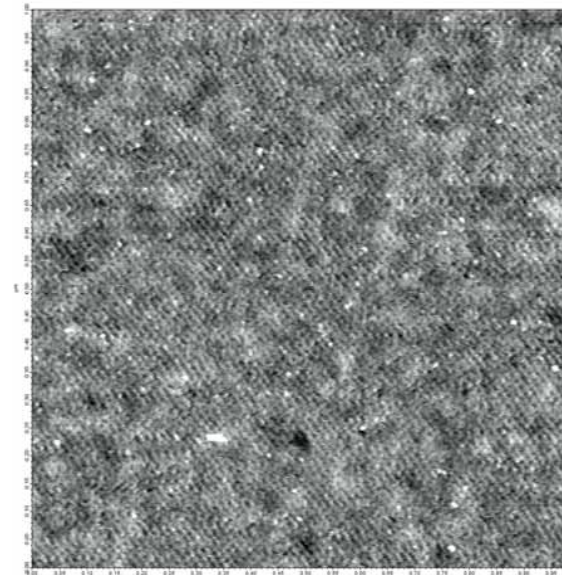
Adding  
metal  
salt  
solution



Adding reducing  
reagents

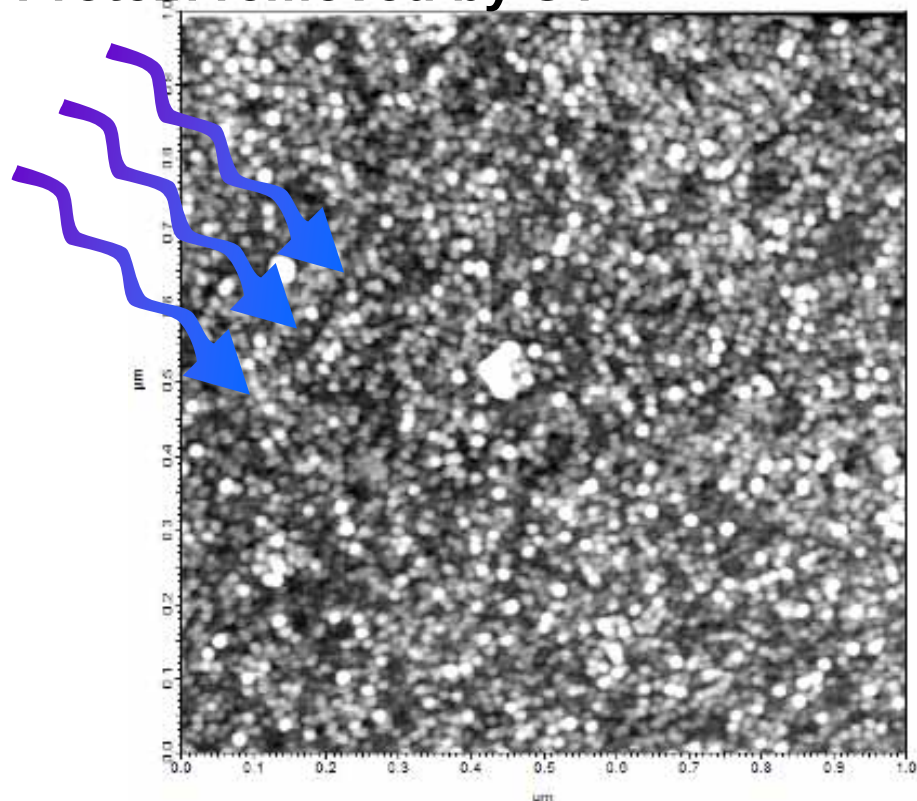


Protein  
removal  
via UV





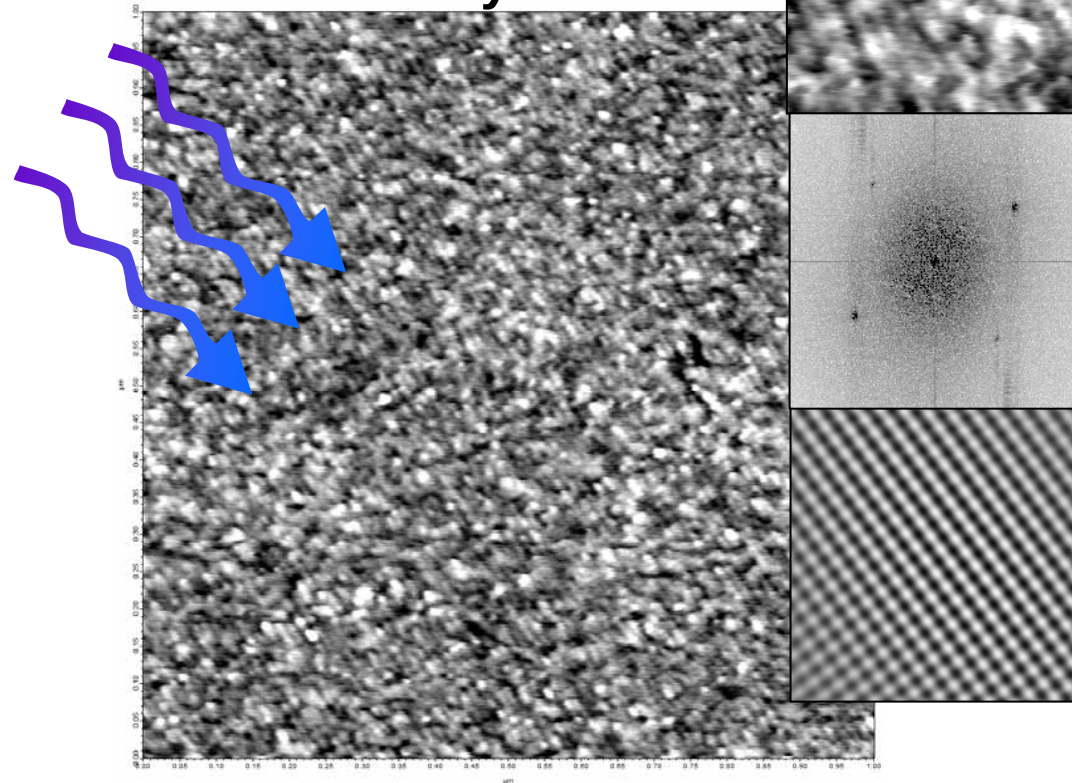
Protein removed by UV



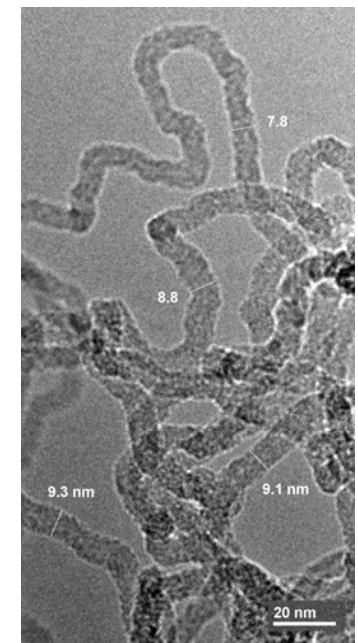
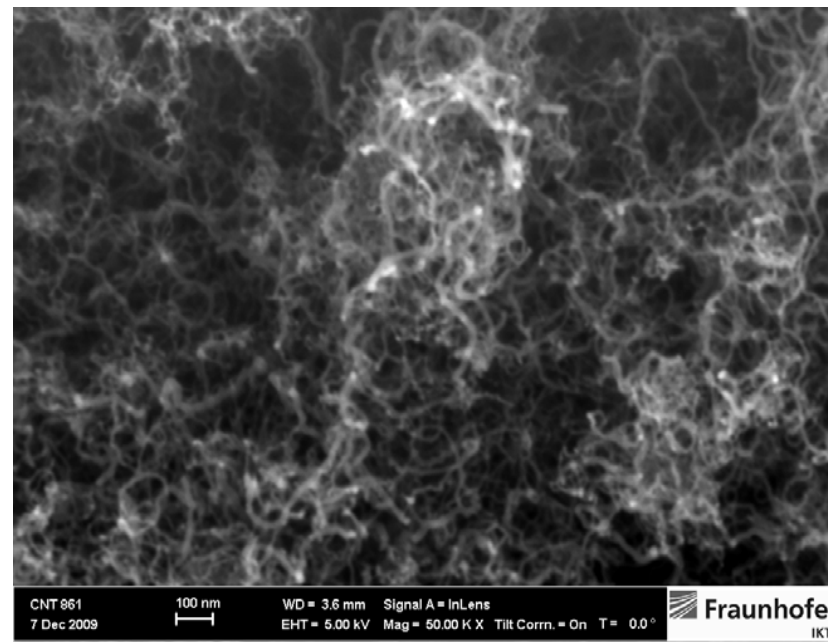
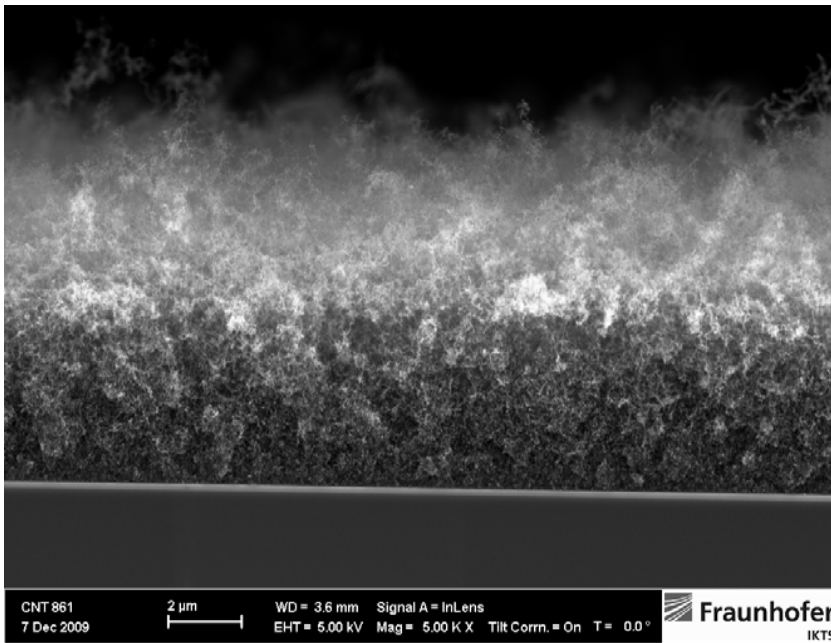
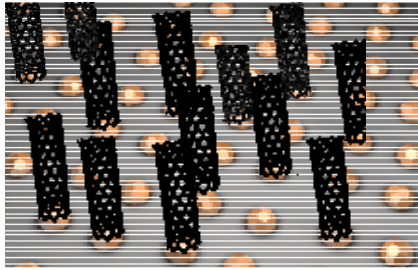
Nanoparticles produced on  
PE without S-layer



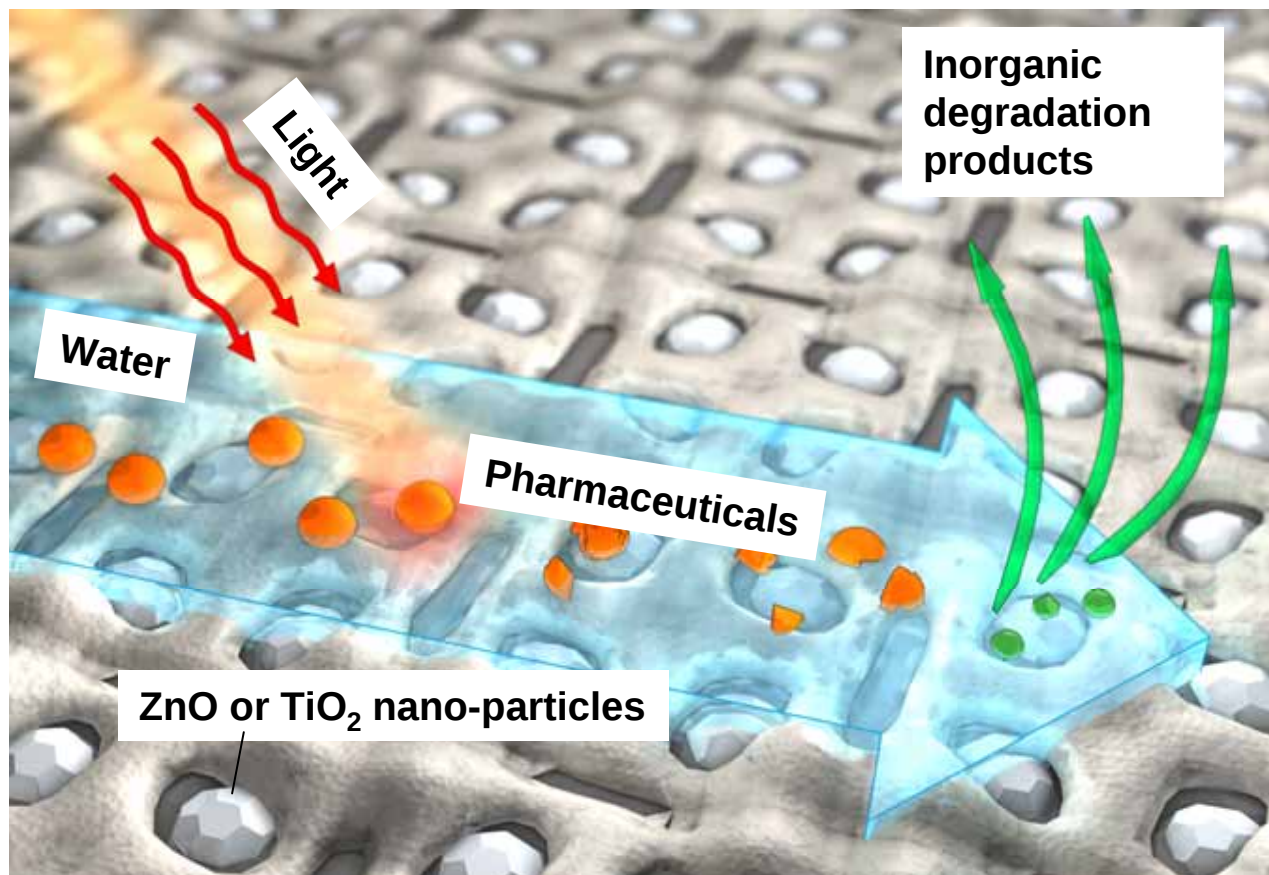
Protein removed by UV



Nanoparticles produced on  
PE + S-layer



- Possibly migration and agglomeration of the particles
- Further improvement necessary



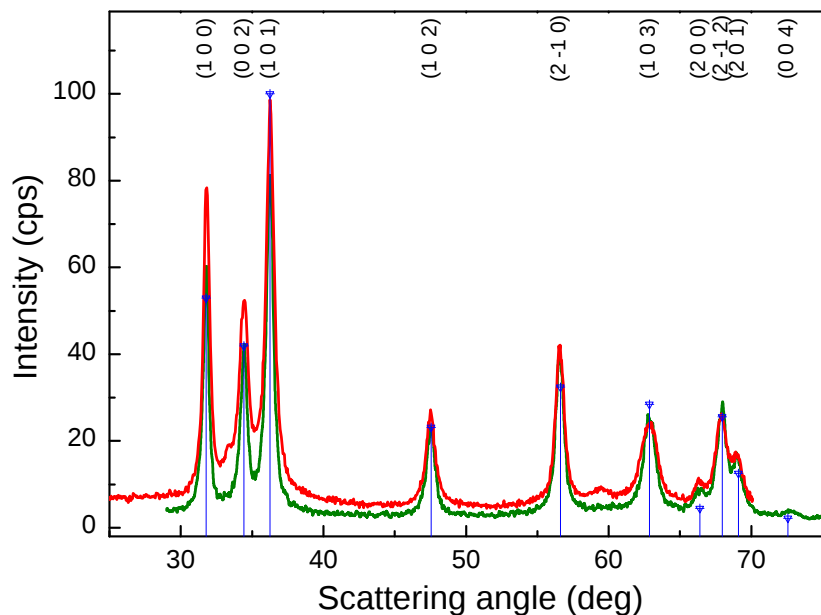
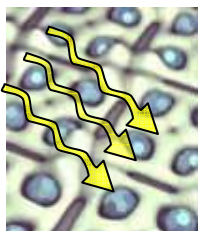
Projects:



NanoAqua

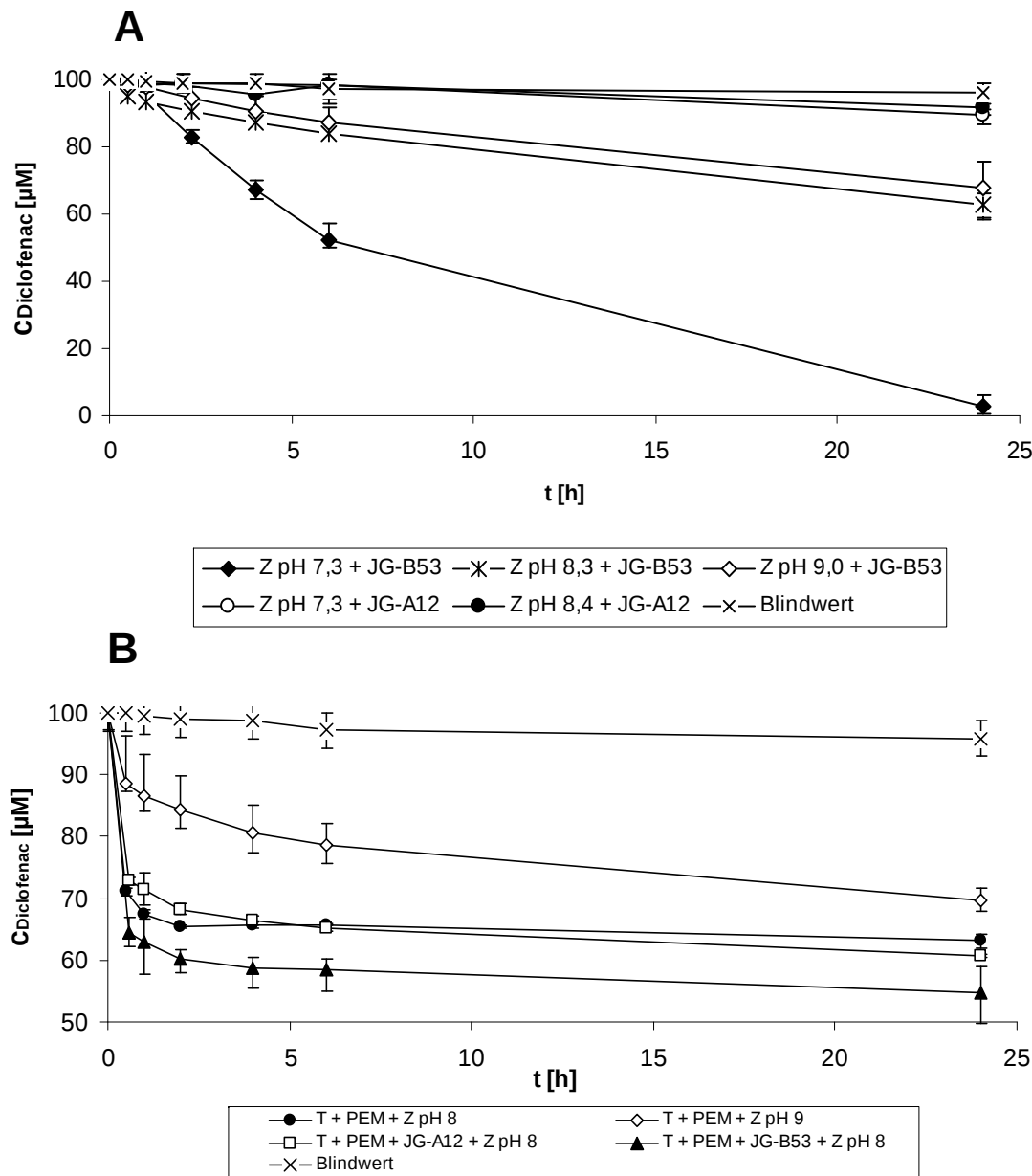
**Advantages compared to previous approaches:**

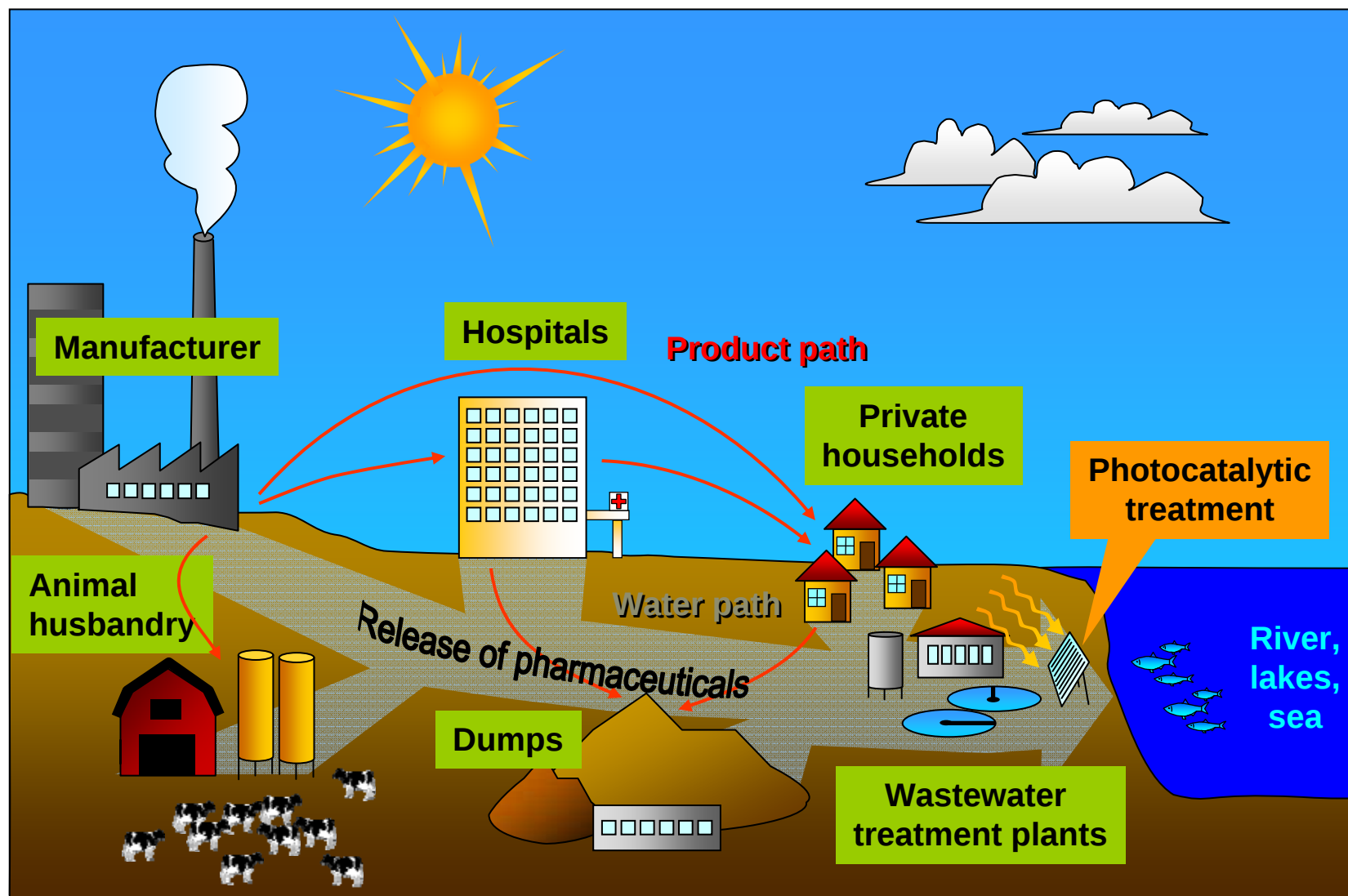
- Environmental-friendly (disposal, immobilization of nano-particles)
- Higher efficiency (required energy, day light sensitivity) ?

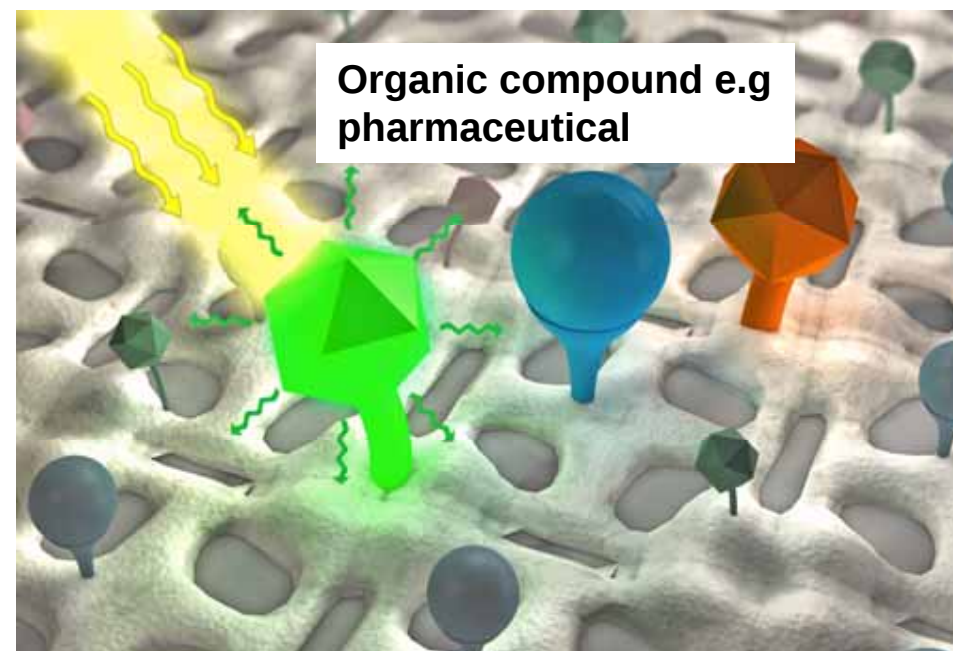
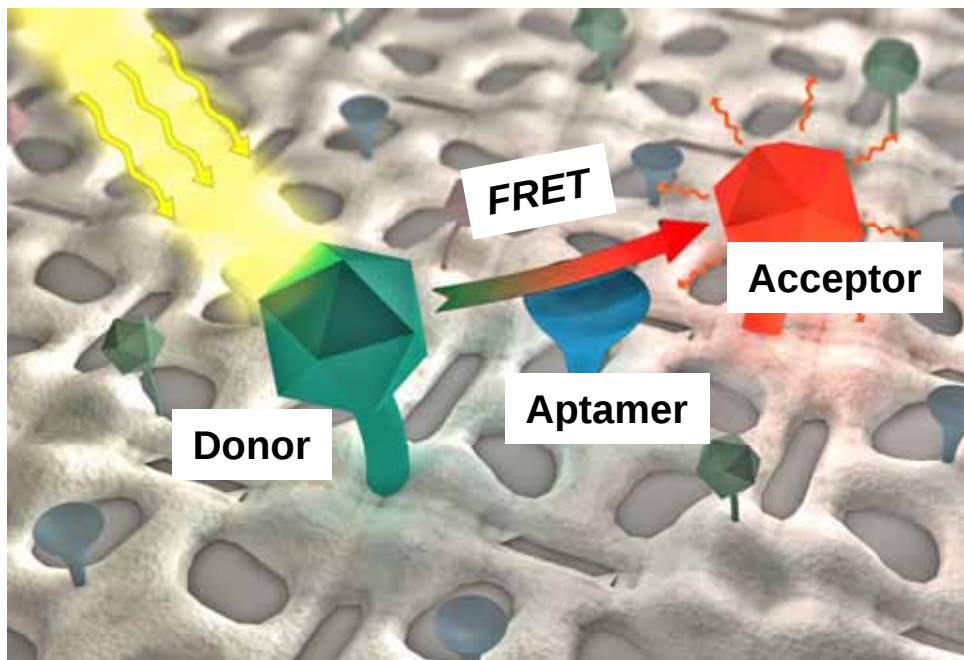


Top: XRD-pattern of S-layer supported ZnO (red graph) and equally prepared ZnO without protein (green graph). S-layer prepared ZnO-particles have a size of about 14 nm.

Right: Diclofenac elimination by S-layer supported ZnO in suspension (A) and immobilized on a carrier (B).







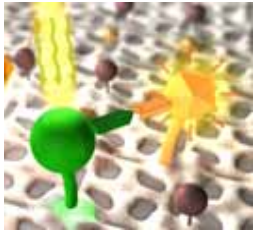
**Project:**



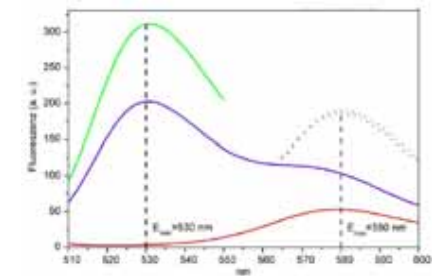
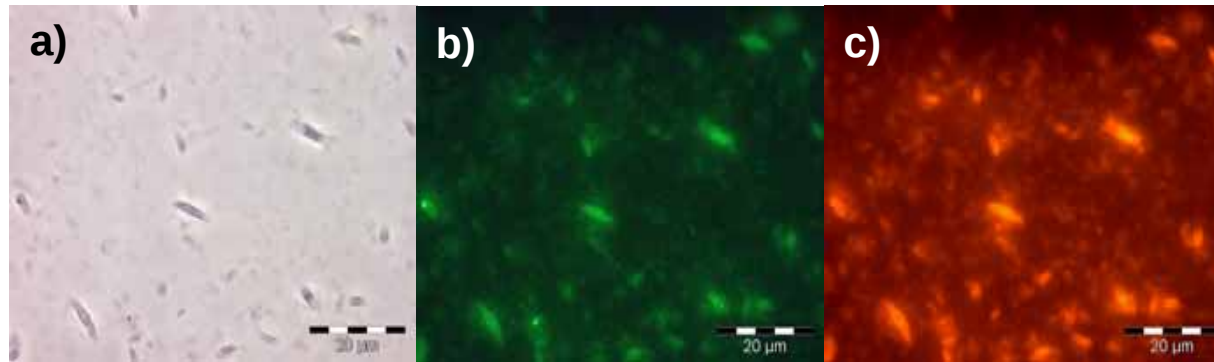
**AptaSens**

**Advantages compared to previous approaches:**

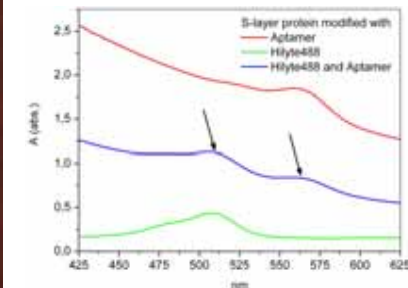
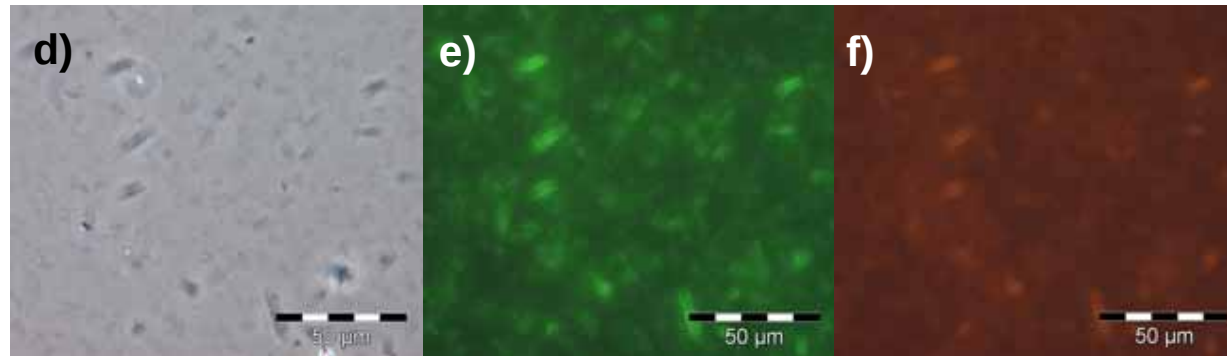
- Transferability
- Chip-based quantification of several analytes ?
- lower unspecific binding ?



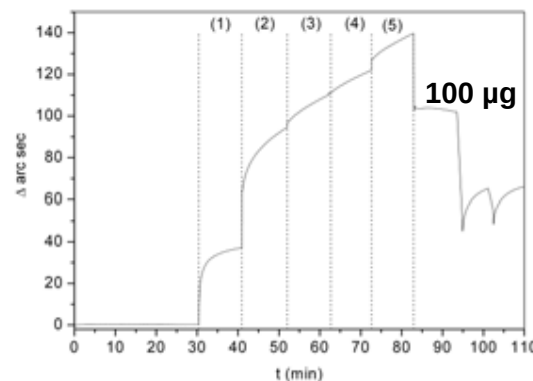
## FRET-pair



## S-layer aptamer composite



## Thrombin binding by the S-layer bound aptamers (IASys)





| Process           | Yield in g/l<br>(z.B. JG A12) |         | Manpower<br>requirements | €/g<br>S-layer |
|-------------------|-------------------------------|---------|--------------------------|----------------|
|                   | Bio-<br>mass                  | S-layer |                          |                |
| Conven-<br>tional | 3                             | 0,03    | 6,9                      | 19000          |
| Pilot plant       | 6                             | 0,16    | 1,4                      | 730            |
| Optimisation      | 12                            | 0,85    | 1,4                      | 100            |
| Aim               |                               |         |                          | <10            |

**Possible strategies:**

- High density cultivation
- Heterologous expression

- **S-layers are essential for many bacteria to survive in extreme environments.**
- **S-layer proteins are highly complex proteins with many different functional groups and modifications, directly affecting their molecular behavior (conformation, hierarchic organization, metal binding).**
- **Their metal binding and self-assembly properties make S-layers very worthwhile for the development of new materials for**
  - **environmental techniques**
  - **biotechnology**
  - **nanotechnology,****but for their application we need to know more about them.**

- **Submitted EU-project “ActiveBrane”**  
Development of reactive nanomembranes with low biofouling for decomposition of organic pollutants in water streams by applying a novel sandwich technology
- **Arsenic binding by S-layers**
  - **Planned DFG project**
  - **Arsenic binding aptamers**
- **Aptamer based sensors for different pollutants**
  - **Planned project**

## Research:

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Dr. Katrin Pollmann

Dr. Harald Foerstendorf  
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Dr. André Rossberg

Dr. Sabine Matys (IfWW)  
Dr. Ulrich Soltmann (GMBU)

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(Universität Granada, Spanien)

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5 Institutes



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Thank you for your attention ...



... questions, remarks?

